

## The Microbial Contamination of Ready-To-Eat Vended Fruits in Abakpa Main Market, Abakaliki, Ebonyi State, Nigeria

<sup>1</sup>Orji Jerry Okechukwu, <sup>1</sup>Orinya Chinedu I., <sup>1</sup>Okonkwo Eucharia O.,  
<sup>2</sup>Uzoh Chukwuma V., <sup>1</sup>Ekuma Uchechukwu O., <sup>3</sup>Ibiam Gideon Ama And  
<sup>4</sup>Onuh Euslar N.

<sup>1</sup>Department of Applied Microbiology, Ebonyi State University, Abakaliki, Nigeria.

<sup>2</sup>Department of Biology, Microbiology and Biotechnology Faculty of Science, Federal University Ndufu-Alike Ikwo, Nigeria.

<sup>3</sup>Department of Medical Laboratory Science, Ebonyi State University Abakaliki, Nigeria.

<sup>4</sup>Ebonyi State School of Health Mgbo, Ohaukwu Local Government Area, Ebonyi State, Nigeria.

---

**Abstract:** The microbial contamination of ready-to-eat vended fruits in Abakpa Main market, Abakaliki was examined using standard microbiological methods. A total of thirty (17) samples of vended fruits were screened for total bacterial and fungal count. From examination five (5) bacterial species were isolated namely: *Escherichia coli*, *Staphylococcus aureus*, *Salmonella sp*, *Shigella sp* and *Pseudomonas sp* while one (1) fungal species, *Mucor sp*, was isolated from the vended fruit samples. The total aerobic plate count ranged from  $3.5 \times 10^5$ - $1.03 \times 10^6$  CFU ml<sup>-1</sup> with tiger nuts having the highest count and cucumber having the lowest count. The total fungal count ranges from  $1.1 \times 10^5$ - $1.42 \times 10^6$  CFU ml<sup>-1</sup> with carrot having the highest count and pineapple (sliced) having the lowest count apart from tiger nuts that had no significant growth. The isolated organisms from the vended fruits showed that contamination occurred due to poor hygiene and environmental factors like contaminated air. Therefore adequate tutorials on sanitary practices on both individuals and environment should be encouraged by concerned government officials to reduce the level of contamination in vended fruits.

---

### I. Introduction

Fruits are an extraordinary dietary source of nutrients, micronutrients, vitamins and fibre for humans and are also vital for health and well-being. Well balanced diets, rich in fruits have been reported to help to prevent Vitamin C and Vitamin A deficiencies and to reduce the risk of several diseases (Kalia and Gupta, 2006). Fruits are widely exposed to microbial contamination through contact with soil, dust and water and also by mishandling during harvest or post harvest processing. They therefore harbour a diverse range of microorganisms including pathogens (Kalia and Gupta, 2006).

Vended fruits are fruits that have been cut or sliced open and carried around by street vendors or hawkers at local markets or streets and such fruits are eaten immediately that is they are eaten without necessarily having to cut, peel or rinse them before consumption because they have already been prepared or packaged by the vendors (Kaplan and Campbell, 1982; Lund, 1992; De Roeber, 1998). They are usually packaged in small polyethylene bags for sale.

Over the last few years, there has been a significant increase in the consumption of vended fruits in Nigeria. This is because they are easily accessible, conveniently, and most importantly, they are cheaper than the whole fruits. Other reasons include modern lifestyle, industrialisation, economic downturn, materialism and lack of time to prepare proper meal (Nielsen, 2006). The increased consumption coupled with the associated risk of disease to which consumers may be exposed, is a matter of great concern. Most times, it is difficult for one to attest to the hygiene of the processors or the sanitary conditions during preparation. This is worsened by the fact that vended fruits are done without adequate storage conditions, thereby, exposing the fruits to flies, dust and other pathogens (Barro *et al.*, 2007). These vended fruits such as watermelon, pineapple, carrots, cucumber, tiger nuts (also known as *aki hausa*) are sold by unlicensed vendors or local hawkers who have little or no knowledge on food hygiene (Muinde and Kuria, 2005). This therefore increases the risk of food-borne diseases caused by a wide range of pathogens such as bacteria (*Salmonella sp*, *Staphylococcus aureus*, *Enterobacteriaceae*), fungi, viruses and parasites (Mensah *et al.*, 1999). And these pathogens could invade these fruits during washing, peeling, slicing, trimming, packaging, handling and marketing (Barro *et al.*, 2007; Khali *et al.*, 2007). The use of dirty utensils encourages rare visits of cockroaches, flies and rats (Bryan *et al.*, 1992).

### Aim Of The Study

To assess the microbial contaminants of some vended fruits sold in Abakpa, Main Market, Abakaliki, Ebonyi State, Nigeria.

### **Objectives Of The Study**

Determination of total aerobic bacteria plate count and total fungal count.

Isolation and identification of bacteria and fungi contaminants from the different ready-to-eat vended fruits.

Determination of the percentage frequency distribution of the microbial isolates on the ready-to-eat vended fruits.

### **Statement Of Problem**

Fruits continue to remain a source of nutrients that are very essential to human health and these nutrients help to reduce the risk of some life threatening diseases like cancer and cardiovascular diseases. Fruits have played a good role in the human health and that is why today they are highly purchased in the markets and streets as well. Some of these fruits are sold in a packaged way that the buyer or consumer does not need to wash or cut the fruits before eating. And most of these vendors are in experienced based on personal hygiene that is to say that during processing, these ready-to-eat vended fruits are exposed to contaminated air, unclean utensils and unclean environment. This is why it is necessary to check for the microbial contamination of such fruits to have an insight on the risk people expose their health to these vended fruits when purchased.

### **Significance**

This study will help give an idea on how contamination of these ready-to-eat vended fruits can pose a threat to human health based on food poisoning.

It will also create awareness to the role of concerned government health officials in the control of microbial contamination in these ready-to-eat vended fruits.

## **II. Materials And Methods**

### **Study Area**

This study was conducted in Applied Microbiology Laboratory Unit, Ebonyi State University, Abakaliki while the samples were collected from different fruit vendors in Abakpa Main Market, Abakaliki, Ebonyi State. Abakpa Main Market, Abakaliki, also known as “meat market” is the largest market in Ebonyi State with different people selling different items like foodstuffs, fruits, vegetables, wears and other exciting goods. A great number of traders there are involved in fruit selling. And most of them are sliced or processed because most of their customers may not be able to afford or have time to process the fruits properly.

### **Materials and Reagents**

The materials and reagents used during the course of this research include: weighing balance, beakers, conical flasks, autoclave, petri-dishes, 70% ethanol, non-absorbent cotton wool, aluminium foil, test tubes, wire loops, incubators, microscope, blender, nutrient agar, potato dextrose agar, mannitol salt agar, salmonella-shigella agar, macConkey agar, peptone water and distilled water.

### **Collection of samples**

A total of seventeen (17) vended fruit samples consisting of carrot, cucumber, tiger nuts, sliced watermelon and pineapple were collected. The sliced watermelon and carrot were collected from four different fruit vendors while the cucumber, sliced pineapple and tigernuts were collected from three different fruit vendors. They were all collected and put into different white polyethene bags to differentiate them based on the vendors they were bought from.

### **Analysis procedure**

**Media preparation:** The different media which included nutrient agar, potato dextrose agar, mannitol salt agar, macConkey agar and salmonella-shigella agar; and peptone water were prepared according to the manufacturer’s instruction.

**Isolation of micro-organisms from the vended fruit samples:** About 10g of each of the fruit samples were weighed and homogenised in 90ml of sterile distilled water using an electric blender. Then, ten-fold dilutions of the homogenates were made with sterilized peptone water; after that 1ml of the  $10^{-4}$  dilutions of the homogenates were dispensed into the petri-dishes that were labelled based on the agar used by pour plate method and allowed to gel. After gelling, the petri-dishes that contained mannitol salt agar, nutrient agar, macConkey agar and salmonella-shigella agar were incubated at 37°C for 24hours while the petri-dishes that contained potato dextrose agar were incubated at 25°C for 3days.

The nutrient agar, macConkey agar, mannitol salt agar and salmonella-shigella agar were used to check for total bacterial count, total coliform count, presence of *Staphylococcus aureus*, *Salmonella* and *Shigella spp* respectively.

At the end of the incubation period, the plates were brought out of the incubators and the colonies were counted using a colony counter device and each count was expressed in colony forming unit per ml (CFU ml<sup>-1</sup>).

#### **Isolation of the cultured micro-organisms**

The distinct colonies on nutrient agar and potato dextrose agar were carefully examined using microscope for their morphological characteristics like colour. Then these colonies were subcultured on nutrient agar using streaking method and were incubated at 37°C for 24hours.

#### **Identification of Isolates**

Gram staining and other biochemical tests were carried out based on the method of Cheesbrough (2006). The biochemical tests performed here included catalase test, oxidase test, indole test and coagulase test.

##### **Biochemical tests**

**Catalase test:** The discrete colonies of each of the isolates were collected with a wooden stick and emulsified in a drop of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>). Bubbles of gas indicated a positive result according to Cheesbrough (2006).

**Indole test:** Here a little portion of each of the isolates was inoculated into 5ml of sterilised prepared peptone water which was contained in different test tubes using a wire loop. And then, the test tubes containing the organisms were left to incubate at 37°C for 48hours. After incubation period, 3-4drops of indole reagent known as Kovac's reagent was added and shook gently. A positive result gave a red surface layer after 10minutes while a negative result gave no red surface layer after 10minutes according to Cheesbrough (2006).

**Oxidase test:** A piece of filter paper was placed in a clean petri dish and 2-3drops of freshly prepared oxidase reagent was added. With the aid of a wooden stick, discrete colonies of the isolates were collected separately and smeared on the filter paper. A positive result gave a purple-blue colouration after 10seconds while a negative result gave no such colour after 10seconds according to Cheesbrough (2006).

**Coagulase test:** A drop of distilled water was placed on each end of a slide and a colony of the test organism was emulsified in each of the drops to form a thick suspension. Then a loopful of plasma was added to one of the suspensions and swirled gently. A positive result showed clumping after 10seconds while a negative result showed no clumping after 10seconds according to Cheesbrough (2006).

**Gram staining:** A thin smear of the isolates were made on different slides with the aid of a wire loop and left to dry and after they were heat fixed and allowed to cool. Then the different smears were covered with crystal violet stain for 30-60seconds and rapidly washed off with clean water. Then the smears were covered with Lugol's iodine for 30-60seconds and rapidly washed off with clean water. The smears were decolourised rapidly with alcohol and washed out immediately with clean water. Then the smears were covered with safranin for 30-60seconds and washed immediately with clean water. The stained smears were then allowed to air-dry. After drying, a few drops of oil immersion were dropped on the stained smears and viewed with the aid of a microscope (×100 oil objective lens) to check for the microscopic properties of the organisms like the Gram reaction, morphology (Cheesbrough, 2006).

For the fungal isolate, a drop of lactophenol cotton blue stain was dropped in the centre of a clean slide. And then a fragment of the fungus was collected with the aid of a wireloop and placed in the drop of the stain and teased gently and covered with a coverslip. The coverslip was not pushed down or tapped to avoid the dislodging of the conidia from the conidiophores. Then the stained isolate was viewed under the microscope with ×10 and ×40 objective lens for its morphological characteristics (Cheesbrough, 2006).

### **III. Results**

The results of the microbial contamination of the processed vended fruit samples collected from different fruit vendors in Abakpa Main market, Abakaliki are presented in the following tables.

Table 1 shows the result of the average microbial load of the vended fruit samples in Colony forming unit per ml (CFU ml<sup>-1</sup>). It reveals that tiger nuts has the highest average total aerobic plate count of  $1.03 \times 10^6$ , followed by watermelon (sliced),  $1.0 \times 10^6$ , while cucumber has the lowest,  $3.5 \times 10^5$ . Moreover, in the total fungal count carrot has the highest of  $1.42 \times 10^6$ , followed by watermelon,  $1.26 \times 10^6$ . No significant fungal growth was observed in tiger nuts.

Table 2 shows the result of the morphological and biochemical characteristics of the microbial isolates from the ready-to-eat vended fruit samples. It reveals that a total of six (6) micro-organisms were isolated. Out of these isolates, five were bacterial isolates namely: *Salmonella sp*, *Pseudomonas sp*, *Escherichia coli*, *Shigella sp* and *Staphylococcus aureus*; and one fungal isolate, *Mucor sp*. All the bacterial isolates are rod-shaped and Gram negative except for *Staphylococcus aureus* that is cocci in shape and Gram positive. Moreover, all bacterial isolates are catalase positive while four bacterial isolates are coagulase positive except for *Pseudomonas sp* which is coagulase negative.

Figure 1 shows the percentage frequency of occurrence of the microbial isolates in the ready-to-eat vended fruit samples. It shows that *Escherichia coli* has the highest occurrence, 5(83.3%), followed by *Salmonella sp.*, *Staphylococcus aureus* and *Mucor sp* with 4(66.7%) respectively. *Pseudomonas sp* and *Shigella sp* has the least occurrence of 2(33.3%).

**Table 1:** Average microbial load of ready-to-eat vended fruit samples (CFU ml<sup>-1</sup>)

Vended Fruit Samples	Vendors	Total Aerobic Plate Count (TAPC)	Average TAPC	Total Fungal Count (TFC)	Average TFC
Carrot	A	9.6 × 10 <sup>5</sup>	7.97 × 10 <sup>5</sup>	1.44 × 10 <sup>6</sup>	1.42 × 10 <sup>6</sup>
	B	6.7 × 10 <sup>5</sup>		TNTC	
	C	TNTC		8.3 × 10 <sup>5</sup>	
	D	7.6 × 10 <sup>5</sup>		1.99 × 10 <sup>6</sup>	
Watermelon (sliced)	E	TNTC	1.0 × 10 <sup>6</sup>	1.77 × 10 <sup>6</sup>	1.26 × 10 <sup>5</sup>
	F	1.6 × 10 <sup>6</sup>		6.4 × 10 <sup>5</sup>	
	G	9.7 × 10 <sup>5</sup>		2.02 × 10 <sup>6</sup>	
	H	4.4 × 10 <sup>5</sup>		6.1 × 10 <sup>5</sup>	
Pineapple (sliced)	I	4.2 × 10 <sup>5</sup>	5.6 × 10 <sup>5</sup>	TNTC	1.1 × 10 <sup>5</sup>
	J	9.0 × 10 <sup>4</sup>		1.1 × 10 <sup>5</sup>	
	K	1.17 × 10 <sup>6</sup>		TNTC	
Cucumber	L	3.2 × 10 <sup>5</sup>	3.5 × 10 <sup>5</sup>	4.5 × 10 <sup>5</sup>	3.0 × 10 <sup>5</sup>
	M	5.4 × 10 <sup>5</sup>		NG	
	N	1.8 × 10 <sup>5</sup>		9 × 10 <sup>4</sup>	
Tiger nuts	O	1.04 × 10 <sup>6</sup>	1.03 × 10 <sup>6</sup>	NG	NG
	P	9.4 × 10 <sup>5</sup>		NG	
	Q	1.12 × 10 <sup>6</sup>		NG	

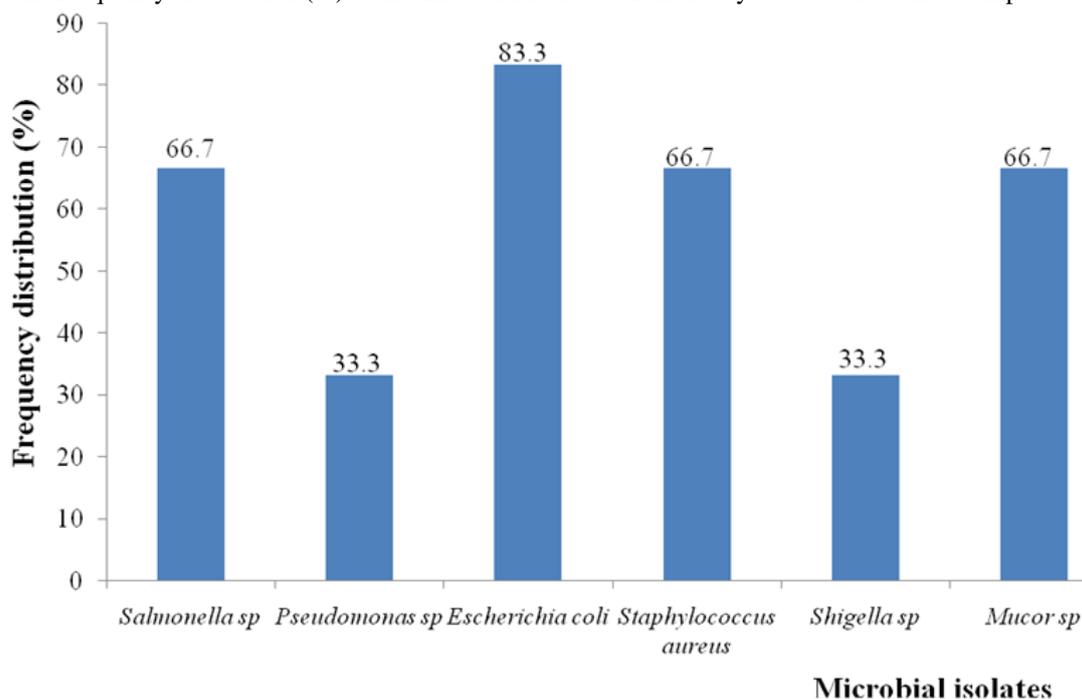
Keys: TNTC: To Numerous To Count  
NG: No Growth

**Table 2:** Morphological and Biochemical characteristics of the microbial isolates from the ready-to-eat vended fruit samples.

S/N	Vended Fruit samples	Biochemical test				GRAM REACTION			Morphological characteristics	Probable organisms
		CA	CO	OX	IND	+ve/-ve	Shape	Arrangement		
1	Carrot	+	-	-	-	-	Rod	Single	Pale white with black edges	<i>Salmonella species</i>
		+	+	+	-	-	Rod	Single	Yellow-green	<i>Pseudomonas species</i>
		+	-	-	+	-	Rod	Single	Pink	<i>Escherichia coli</i>
		+	-	+	+	+	Cocci	Group	Yellow	<i>Staphylococcus aureus</i>
										Cottony dark-grey branched with round sporangiospores
2	Watermelon (Sliced)	+	-	-	+	-	Rod	Single	Pink	<i>Escherichia coli</i>
		+	-	-	-	-	Rod	Single	Pale white with black edges	<i>Salmonella species</i>
		+	-	+	+	+	Cocci	Single	Yellow	<i>Staphylococcus aureus</i>
		+	-	-	-	-	Rod	Single	Pale white	<i>Shigella species</i>
										Cottony dark-grey branched with round sporangiospores
3	Pineapple (Sliced)	+	-	-	+	-	Rod	Single	Pink	<i>Escherichia coli</i>
		+	-	-	-	-	Rod	Single	Pale white with black edges	<i>Salmonella species</i>
		+	-	+	+	+	Cocci	Group	Yellow	<i>Staphylococcus aureus</i>
		+	-	-	-	-	Rod	Single	Pale white	<i>Shigella species</i>
										Cottony dark-grey branched with round sporangiospores
4	Cucumber	+	-	-	+	-	Rod	Single	Pink	<i>Escherichia coli</i>
		+	-	+	+	+	Cocci	Group	Yellow	<i>Staphylococcus aureus</i>
										Cottony dark-grey branched with round sporangiospores
5	Tigemuts	+	-	-	+	-	Rod	Single	Pink	<i>Escherichia coli</i>
		+	+	+	-	-	Rod	Single	Yellow-green	<i>Pseudomonas sp</i>
		+	-	-	-	-	Rod	Single	Pale white with black edges	<i>Salmonella species</i>

Keys: CA= Catalase test, + = Positive, CO= Coagulase test, - = Negative, OX= Oxidase test, IND= Indole test

**Figure 1:** Frequency distribution (%) of the microbial isolates on the ready-to-eat vended fruit samples.



#### IV. Discussion

Bacteria and fungi are the common contaminants of our fruits and they could be easily transferred from the vendors to the processed fruits through mishandling. The consumption of ready-to-eat fruits directly from street vendors or hawkers potentially increase the risk of food-borne diseases caused by a wide variety of pathogens, because it is difficult to attest to the hygiene of these vendors or to the sanitary conditions at points of processing as well as the packaging materials. This could pose a threat to human health and this helps to throw light to the microbial contamination of ready-to-eat vended fruits that were collected from different fruit vendors in Abakpa Main market, Abakaliki.

These micro-organisms isolated were *Escherichia coli* (83.3%), *Salmonella sp* (66.7%), *Pseudomonas sp* (33.3%), *Staphylococcus aureus* (66.7%), *Shigella sp* (33.3%) and *Mucor sp* (66.7%). All the microbial isolates apart from *Shigella sp* was reported in the work of Odebisi-Omokanye *et al.*, (2015) in the microbial quality of pre-cut fruits sold in Ilorin, Kwara state; Jolaoso *et al.*, (2010) isolated *Staphylococcus aureus*, *Salmonella sp* and *Escherichia coli* from sliced pineapple and paw-paw. This is further supported by the work of Oranusi and Olurunfemi, (2011) that isolated *Staphylococcus aureus*, *Pseudomonas sp*, *Salmonella sp* and *Escherichia coli* from ready-to-eat fruits sold in Otta, Ogun state; Tambeker *et al.*, (2009) also isolated *Staphylococcus aureus*, *Pseudomonas sp*, *Salmonella sp* and *Escherichia coli* from street vended fruits juices in Amravati, India. Moreover, the result of this study is in line with the report of Fowoyo, (2012) from air-contaminated vended foods sold in Lokoja, Kogi state.

Most of the isolates in this study may have been introduced into these fruits through faecally polluted water used in washing utensils like knives, trays and polyethene bags used for the packaging of the fruits after slicing or cutting and also exposure of these fruits to low temperatures which encourage the microbial growth of these pathogens (Daniyan and Ajibo, 2011). The presence of *Staphylococcus aureus*, *Pseudomonas sp*, *Salmonella sp* and *Escherichia coli* was in line with the work of Odebisi-Omokanye *et al.*, (2015) from pre-cut fruits sold in Ilorin. *Staphylococcus aureus*, *Salmonella sp*, *Shigella sp*, *Pseudomonas sp* and *Escherichia coli* are environmental isolates and they have been isolated from plants, human skin, animal and dairy products. Their presence in these ready-to-eat fruits may have been through unclean hands of the vendors, contact with sewage and contaminated water (De Roeve, 1998). This implies that the fruit samples could serve as a vehicle in the transmission of these pathogens to the consumers of these contaminated fruits.

The presence of *Staphylococcus aureus* may have been introduced into the ready-to-eat fruits through body contact of vendors with the fruits because the organism is a normal flora of the nasal passage, hands and skins of healthy individuals (Nester *et al.*, (2006). Odebisi-Omokanye *et al.*, (2015) and Ganguli, (2006) reported *Staphylococcus aureus* to have the highest occurrence in fruits and foods respectively. It was recorded to be the second highest occurring isolate with the frequency of occurrence of 4(66.7%). Aboloma, (2008) and Wada-Kura *et al.*, (2009) have also reported that the incidence of *Staphylococcus aureus* in food is an indication of

environmental and human contamination. This high incidence may have occurred due to the use of polyethylene bags for the packaging of these fruits after slicing or cutting them (Little and Mitchell, 2004).

In this study, *Mucor sp*, *Salmonella sp* and *Staphylococcus aureus* had the same incidence of 4(66.7%). Oviasogie *et al.*, (2015) reported such incidence of *Mucor spin* the assessment of fungal pathogens associated with orange spoilage sold in Benin, Edo state while Oluwatoyin *et al.*, (2015) reported such high incidence in *Salmonella sp* and *Staphylococcus aureus* in assessment of the microbial safety of polyethylene packaged sliced fruits sold in Abeokuta, Ogun state. The presence of *Mucor sp* promotes the contamination and because they are ubiquitous they can be found on fresh vegetables, fruits and other substances that give nutrients. They are also able to withstand high concentration of sugar and they can survive in the absence of water or moisture. Such high occurrence may have occurred as a result of the exposure of these ready-to-eat fruits to dusty or muddy areas. Most of these fruit vendors stay near stagnant water of gutters which may serve as an entry for fruit contamination. Frank and Warribor, (2006) reported that the microbial load on leafy vegetables and fruits increase with time during storage. When these fruits are stored at inappropriate temperatures, they tend to attain temperatures that are suitable for the microbial growth of these pathogens to cause diseases when ingested (Bryan *et al.*, 1992; Muinde and Kuria, 2005).

The results show that *Escherichia coli* had the highest frequency of occurrence of 5(83.3%) and it conforms to the report by Daniyan and Ajibo, (2011) and Daniel *et al.*, (2014) in sliced fresh fruits sold in Minna and Bida metropolis respectively. *Escherichia coli* is regarded as primary indicator for microbiological quality of food and water and this shows that these fruits are not safe for human consumption. According to CDC, (2011), the main transmission of *Escherichia coli* was through faecally contaminated food or water. The high occurrence may have occurred in the contact of contaminated water with the fruits during washing of the fruits and also the inadequate washing of hands by the fruit vendors (Tambekar *et al.*, 2007). Some of these fruit vendors get their water from unclean sources like dirty streams and also they could use very little quantity of water to wash or rinse all the fruits. The low occurrence of *Pseudomonas sp* and *Shigella sp* was also reported by Fowoyo, (2012) in the assessment of air contaminated vended foods sold in Lokoja, Kogi state.

These ready-to-eat fruits may get contaminated from knives used for cutting or slicing, improper human handling and processing, tables or trays used during peeling and cutting, rinsed water, washing buckets and packaging materials as these fruits are cut, washed, wrapped with transparent polyethylene bags and sold to the consumers. The presence of these possible pathogens in the analysed fruit samples should be of great importance to the vendors, consumers and concerned arms of government.

## V. Conclusion

In conclusion, the result from this study has shown that poor hygiene of the vendors and environmental factors could cause the microbial contamination of these processed vended fruits sold in Abakpa Main market, Abakaliki. From time to time, government health officials should give attention to the market especially these fruit vendors, at least to put on check how these vended fruits are processed which includes the type and source of water used, the condition of the utensils and most especially the personal hygiene of the fruit vendor to reduce help the rate of vended fruit contamination. Public awareness programs can also be used as a measure to educate these fruit vendors on personal and environmental hygiene to reduce contamination.

## References

- [1]. Abano, E.E., Amoah, K.K., (2011). "Effect of moisture content on the physical properties of tiger nut (*Cyperus esculentus*)". *Asian Journal of Agricultural Research* 5:56-66.
- [2]. Abbott, Catherine (2012). *The Year-Round Harvest: A Seasonal Guide to Growing, Eating, and Preserving the Fruits and Vegetables of Your Labour*. Adams Media. pp. 54-55.
- [3]. Aboloma, R.I., (2008). Microbiological analysis of bread samples from bakery to sale points in Ado-Ekiti, Ekiti state, Nigeria. *Biol. Environ. Sci. J. Tropics*, 5:77-81.
- [4]. Adebayo-Tayo, B.C., Okonko, I.O., Esen, C.U., Odu, N.N., (2012). Microorganisms Associated with Spoilage of Stored Vegetables in Uyo Metropolis, Akwa Ibom State, Nigeria. *Nature and Science*,10(3):23-32.
- [5]. Adejuyitan, J.A., Otunola, E.T., Akande, E.A., Bolarinwa, I.F. and Oladokun, F.M., (2009). Some Physicochemical properties of Flour obtained from fermentation of tiger nut (*Cyperus esculentus*) sourced from a market in Ogbomoso, Nigeria. *Afric. J. of Food Science*, 3: 51-55.
- [6]. Al-Ghazali, M.R. and Al-Azawi, S.K., (1990). *Listeria monocytogenes* contamination of crops grown on soil treated with sewage sludge cake. *J. Appl. Bacteriol.*, 69:642-647.
- [7]. Angela, O.E., Ibukunoluwa, A.O. and Oranusi, U.S., (2010). Microbial quality of fruits and vegetables sold in Sango Ota, Nigeria. *Afr. J. Food Sci.*, 4:291-296.
- [8]. Anon (1996). *Ministry of the Environment Guidelines for the Utilization of Biosolids and Other Wastes on Agricultural Land*, 1996. Toronto, ON: Ministry of the Environment, <http://www.ene.gov.on.ca/envision/gp/3425e.pdf>.
- [9]. Anon (2001). *Analysis and Evaluation of Preventive Control Measures for the Control and Reduction / Elimination of Microbial Hazards on Fresh and Fresh-Cut Produce, Chapter IV, Outbreaks Associated with Fresh Produce: Incidence, Growth, and Survival of Pathogens in Fresh and Fresh-Cut Produce*. Rockville, MD: US Food and Drug Administration, <http://www.cfsan.fda.gov/~comm/ift3-4o.html>.
- [10]. Anon (2001). Diagnosis and Management of Foodborne Illnesses: A Primer for physicians. *MMWR January*26, 50:1-69.

- [11]. Anon (2003). *Water for People, Water for Life: Executive Summary*. United Nations World Water Development Report 2003. Paris, France: UNESCO Publ., from <http://unesdoc.unesco.org/images/0012/001295/129556e.pdf>.
- [12]. Arshad, Z.I., Amid, A., Yusof, F., Jaswir, I., Ahmad, K., Loke, S.P., (2014). "Bromelain: an overview of industrial application and purification strategies". *Appl. Microbiol. Biotechnol.* 98(17):7283-7297.
- [13]. Avery, L.M., Killham, K. and Jones, D.L., (2005). Survival of *E. coli* O157: H7 in organic wastes destined for land application. *J. Appl. Microbiol.*, 98:814–822.
- [14]. Baeza, R., Rössler, C.E., Mielnicki, D.M., Zamora, M.C., Chirife, J., (2007). Simplified prediction of *Staphylococcus aureus* growth in a cooked meat product exposed to changing environmental temperatures in warm climates. *Rev Argent Microbiol.*, 39(4):237-42.
- [15]. Baranska, Malgorzata; Schulz, Hartwig; Baranski, Rafal; Nothnagel, Thomas; Christensen, Lars P., (2005). In situ simultaneous analysis of polyacetylenes, carotenoids and polysaccharides in carrot root tissue. *Physiological Plant Pathology* 13(2): 241-246.
- [16]. Barro, N., Bello, A.R., Savadogo, A., Ouattara, C.A.T., Ilboudo A.J. and Traore A.S., (2006). Hygienic status assessment of dish washing waters, utensils, hands and pieces of money from street food processing sites in Ouagadougou (Burkina Faso). *African Journal Biotechnology* (5): 1107-1112.
- [17]. Bartholomew, D.P., Paul, R.E., and Rohrbach K.G., (2002). The pineapple: botany, production and uses. *Commonwealth Agricultural Bureau, Intl.* pp. 320.
- [18]. Bean, N.H. and Griffin, P.M., (1990). Foodborne disease outbreaks in the United States, 1973-1987: pathogens, vehicles and trends. *J. Food Prot.*, 53(9):804-817.
- [19]. Belew, M.A. and Belew, K.Y., (2007). Comparative physicochemical evaluation of tiger nut, soyabean and coconut milk sources. *Int. Journal Agric. Biol.*, 9:785-787.
- [20]. Benjamin, L.R., McGarry, A., Gray, D., (1997). "The root vegetables: Beet, carrot, parsnip and turnip". *The Physiology of Vegetable Crops*. Wallingford, UK: CAB International, pp. 553-580.
- [21]. Beuchat, L.R., (1996) Pathogenic microorganisms associated with fresh produce. *J. Food Prot.* 59:204–216.
- [22]. Blostein, J., (1991). An outbreak of *Salmonella javiana* associated with consumption of watermelon. *J. Environ. Health.* 56:29-31.
- [23]. Brickell, Christopher (ed) (1992). *The Royal Horticultural Society Encyclopedia of Gardening (Print)*. London: Dorling Kindersley. p. 333
- [24]. Bryan, F.L., Teufel, P., Riaz, S., Roohi, S., Qadar, F. and Malik, Z.U.R., (1992). Hazards and critical control points of street-vended chat, a regionally popular food in Pakistan. *Journal Food Prot.*, 55: 708-713.
- [25]. CDC, (1990). CDC surveillance summaries; March 1, 2000. *MMWR*, 39(SS-1):15-23.
- [26]. CDC, (2000). CDC surveillance summaries; March 17, 2000. *MMWR*, 49(SS-1):1-51.
- [27]. CDC, (2005). Outbreaks of *Salmonella* infections associated with eating tomatoes – United States and Canada, 2004. *Morb. Mortal WklyRep.*, 54:325–328.
- [28]. CDC, (2006). Ongoing multistate outbreak of *Escherichia coli* serotype O157:H7 infections associated with consumption of fresh spinach – United States, September 2006. *Morb. Mortal Wkly Rep.*, 55:1045–1046.
- [29]. Chan, Y.K., (2006). Hybridisation and selection in pineapple improvement: The Research Experience. *Acta Horticulturae* 702:87-92.
- [30]. Chaurasiya, R. ., Sakhare, P.Z., Bhaskar, N., Hebbar, H.U., (2015). "Efficacy of reverse micellar extracted fruit bromelain in meat tenderisation". *J. Food Sci Technol.* 52(6):3870-3880.
- [31]. Cheesbrough, M., (2006). District Laboratory Practice in Tropical countries. *Cambridge University Press, Cambridge, U.K.*, pp. 62-70.
- [32]. Chenghui, L., Beiwei, Z., Xiuping, D., and Ligu, C., (2007). Study on the separation and antioxidant activity of enzymatic hydrolysates from sea cucumber. *Food Ferment. Ind.*, 2007, vol. 33, pp. 50–53.
- [33]. Chiang, Y.C., Liao, W.W., Fan, C.M., Pai, W.Y., Chiou, C.S., Tsen, H.Y., (2008). PCR detection of Staphylococcal enterotoxins (SEs) N, O, P, Q, R, U, and survey of SE types in *Staphylococcus aureus* isolates from food-poisoning cases in Taiwan. *Int J Food Microbiol.*, 121(1):66-73.
- [34]. Coppens d' Eeckenbrugge, G. and Leal, F. (2003). Morphology, Anatomy and Taxonomy. In: Bartholomew, DP, Paull, RE and Rohrbach, KG (eds) *The Pineapple: Botany, Production and Uses*. CABI Publishing, Oxon, UK, pp 13-32.
- [35]. Coyne, T., Ibiebele, T.I., Baade, P.D., Dobson, A., McClintock, C., Dunn, S., Leonard, D. and Shaw, J. (2005). Diabetes Mellitus and Serum Carotenoids: Findings of a Population-Based Study in Queensland, Australia. *The American Journal of Clinical Nutrition*, 82:685-693.
- [36]. Cunningham, Sally Jean (2000). *Great Garden Companions: A Beautiful, Chemical-Free Vegetable Garden*. pp 195-196.
- [37]. Czepa, Andreas; Hofmann, Thomas (2003). "Structural and sensory characterisation of compounds contributing to the bitter off taste of carrots and carrot puree". *Journal of Agricultural and Food Chemistry*, 51(13): 3865-3873.
- [38]. Dane, F. and Liu, J., (2006). "Diversity and origin of cultivated and citron type watermelon (*Citrullus lanatus*)". *Genetic Resources and Crop Evolution* 54(6):1255.
- [39]. Dane, Fenny; Liu, Jiarong (2006). "Diversity and origin of cultivated and citron type watermelon (*Citrullus lanatus*)". *Genetic Resources and Crop Evolution* 54 (6): 12-55
- [40]. Daniel, A.A., Danfulani, S., Barnabas, B.B., Peter, G. and Ajewole, A.E., (2014). Microbiological quality of sliced fresh fruits sold in Bida, Nigeria. *Global Journal of Biology, Agriculture and Health Sciences*, 3(3):178-180.
- [41]. Daniyan, S. Y. and Ajibo, C. Q., (2011). Microbiological examination of selected fruit sold in Minna metropolis. *International Research Journal of Pharmacy*, 2(7), pp 124-129.
- [42]. Davis, R., (2004). "Carrot diseases and their management". *Diseases of Fruits and Vegetables: Diagnosis and Management*. pp. 397-439.
- [43]. De Rover, C., (1998). Microbial Safety Evaluations and Recommendation on Fresh Produce. *Food Control* 9(6): 321-347.
- [44]. Defelice, M.S., (2002). "Yellow nutsedge, *Cyperus esculentus* L. - snack food of the gods. *Weed Technology*, 16:901-907.
- [45]. Dent, D. R., (1995). Control measures. In: Integrated Pest Management. *chapman and Hall, London*. pp. 47-81.
- [46]. Dias, J.S., (2012). Nutritional Quality and Health Benefits of Vegetables: A Review. *Food and Nutrition Sciences*, 3:1354-1374.
- [47]. Doijode, S. D., (2001). Seed Storage of Horticultural crops. *Haworth Press*. pp. 281.
- [48]. Duff, S.B., Scott, E.A., Mafilios, M.S., Todd, E.C., Krilov, L.R., Gedded, A.M. and Ackerman, S.J., (2003). Cost effectiveness of a targeted disinfection program in household kitchens to prevent food-borne illnesses in the United States, Canada and United Kingdom. *J. food Prot.*, 2:2103-2115.
- [49]. Ekam, V.S., Udosen, E.O. and Chighu, A.E., (2006). Comparative Effect of Carotenoid Complex from Goldenro- Life Dynamite and Carrot Extracted Carotenoids on Immune Parameters in Albino Wistar Rats. *Nigerian Journal of Physiological Sciences*, 21:1-4.

- [50]. Elzer-Peters, K., (2014). Midwest Fruit and Vegetable Gardening: Plant, Grow, and Harvest the Best Edibles. *Cool Springs Press*. pp 136.
- [51]. Enujeke E. C., Ojeifo I. M., Nnaji G. U. (2013). Residual effects of organic manure and inorganic fertilizer on maize grain weight and some soil properties in Asaba area of Delta State. *International J. Advanced Biol. Res.* 3(3):433-442.
- [52]. Enujeke E.C., (2013). Growth and yield responses of cucumber to five different rates of poultry manure in Asaba area of Delta state, Nigeria. *Int. Res. J. Agric. Sci. Soil Sci.* 3(11):369-375.
- [53]. Faruque, S.O., Albert, M.J., Mekalanos J.J., (1998). Epidemiology, genetics and ecology of toxigenic *Vibrio cholerae*. *Microbiology and Molecular Biology Reviews* 62: 1301-1314.
- [54]. FDA, (2001). Hazard Analysis and Critical Control Point (HACCP); Procedures for the Safe and Sanitary Processing and Importing of Juices. *Federal Register: January 19, 2001*, 66:6137–6202.
- [55]. Fowoyo, P.T., (2012). Microbiological assessment of air contamination of vended foods sold in the main market in Lokoja, Kogi state, Nigeria. *Research Journal of Biological Sciences*, 7(9-12):355-360.
- [56]. Francis, G.A., Thomas, C. and O'Beirne, T., (1999). The microbiological safety of minimally processed vegetables. *International Journal of Food Science and Technology*, 34: 1-22.
- [57]. Francis, S.O., Morufu, E.B. and Adedeji, G.T., (2013). Antisecretory Effects of Watermelon (*Citrullus lanatus*) Juice in Male Albino Rats. *Annual Review & Research in Biology*. 3(4): 358-366.
- [58]. Frank-Peterside, N. and Warribor, O., (2006). Bacteria-associated spoilage of fluted pumpkin leaves and their effect on the chlorophyll content. *Nig. Journal of Microbiol.*, 20(1):751-756.
- [59]. Gilani, A.H., Shaheeri, F., Saeed, S.A., Bibi, S., Irfamillah-Sadiq, M. and Faiz, S. (2000). Hypotensive Action of Coumarin Glycoside from *Daucus carot*. *Phytomedicine*, 7:423-426.
- [60]. Gill, N.S., Bansal, R.K., Garg, M., Sood, S., Muthuraman, A., and Bali, M., (2010). Evaluation of antioxidant, antiinflammatory and analgesic potential of *Citrullus lanatus* seed extract in rodent model. *The Internet Journal of Nutrition and Wellness*, 9 Number 2.
- [61]. Gordon, T.R. and Martyn, R.D., (1997). The evolutionary biology of *Fusarium oxysporum*. *Annual Review of Phytopathology*, 35:111-128.
- [62]. Gramenzi, A., Gentile, A., Fasoli, M., Negri, E., Parazzini, F. and La Vecchia, C. (1999). Association between Certain Foods and Risk of Acute Myocardial Infarction in Women. *Biol. Microbiol. J*, 300:771-773.
- [63]. Greene, W., (2012). Vegetable Gardening the Colonial Williamsburg Way: 18<sup>th</sup>- Century Methods for Today's Organic Gardeners. pp. 81.
- [64]. Hartke, A., Bouche, S., Laplace, J.M., Benachour, A., Boutibonnes, P. and Auffray, Y., (1995). UV-inducible proteins and UV-induced cross-protection against acid, ethanol, H<sub>2</sub>O<sub>2</sub> or heat-treatments in *Lactococcus-lactis* subsp *lactis*. *Arch. Microbiol.*, 163:329–336.
- [65]. Hedberg, C.W. and Osterholm, M.T., (1993). Outbreaks of food-borne and water-borne viral gastroenteritis. *Clinical Microbiology Reviews* 6: 199–210.
- [66]. Hill, D. S. and Waller, J. M., (1999). Pests and Diseases of Tropical Crops, Vol. 2 (ed.). *Longman, Ghana*. p.179-182.
- [67]. Huang, S., Zhang, Z., Lucas, W., Ruan, J., Qian, W., Wang, M., (2009). The genome of the cucumber, *Cumis sativus* L. *Nature Genetics* 4(12):1275-1281.
- [68]. Hutchison, M.L., Walters, L.D., Moore, A., Crookes, K.M. and Avery, S.M., (2004). Effect of length of time before incorporation on survival of pathogenic bacteria present in livestock wastes applied to agricultural soil. *Appl. Environ. Microbiol.*, 70:5111–5118.
- [69]. Islam, M., Morgan, J., Doyle, M.P., Phatak, S.C., Millner, P. and Jiang, X., (2004). Fate of *Salmonella enteric* serovars *typhimurium* on carrots and radishes grown in fields treated with contaminated manure composts or irrigation water. *Appl. Environ. Microbiol.*, 70:2497–2502.
- [70]. Janisiewicz, W.J., Conway, W.S., Brown, M.W., Sapers, G.M., Fratamico, P. and Buchanan, R.L., (1999). Fate of *Escherichia coli* O157:H7 on fresh-cut apple tissue and its potential for transmission by fruit flies. *Appl. Environ. Microbiol.*, 65:1–5.
- [71]. Jiyun, A., Wonhee, C., Suna, K., and Taeyoul, H., (2011). Anti-diabetic effect of watermelon (*Citrullus vulgaris*, Schrad) on Streptozotocin-induced diabetic mice. *Food Science and Biotechnology*, Volume 20(1): 251-254.
- [72]. Kalia A., Gupta, R.P., (2006). *Handbook of fruit and fruit processing*, 1st Edition, Blackwell publishing, pp. 3-28.
- [73]. Kaplan, J.E., Campbell, D.S., (1982). Frequency of Norwalk like pattern of illness in outbreak of acute gastroenteritis. *Am. J. Pub Health*, 72:1329 – 1332.
- [74]. Kaplan, J.E., Feldman, R., Campbell, D.S., Lookabaugh, C., (1982). The frequency of a Norwalk-like pattern of illness in outbreaks of acute gastro-enteritis. *American Journal of Public Health* 72:1329-1332.
- [75]. Kim, K.Y., Kimb, H.T., Kim, D., Nakajimad, J. and Higuchi, T., (2009). Distribution characteristics of airborne bacteria and fungi in the feedstuff-manufacturing factories. *J. Hazard Mat.*, 169:1050-1060.
- [76]. Krech, S., McNeill, J. R., Merchant, Carolyn (2004). *Encyclopedia of World Environmental History*. p. 1071.
- [77]. Kudva, I.T., Blanch, K. and Hovde, C.J., (1998). Analysis of *Escherichia coli* O157:H7 survival in ovine or bovine manure and manure slurry. *Appl. Environ. Microbiol.*, 64:3166–3174.
- [78]. Larkin, R.P. and D.R. Fravel. 1998. Efficacy of various fungal and bacterial biocontrol organisms for control of *Fusarium* wilt of tomato. *Plant Dis.* 82:1022-1028.
- [79]. Leyer, G.J. and Johnson, E.A., (1993). Acid adaptation induces cross-protection against environmental stresses in *Salmonella typhimurium*. *Appl. Environ. Microbiol.*, 59:1842–1847.
- [80]. Lindqvist, R., Andersson, Y., De Jong, B., Norberg, P., (2000). A summary of reported foodborne disease incidents in Sweden, 1992-1997. *Journal of Food Protection*, 63:1315-1320.
- [81]. Little, C.L. and Mitchell, R.T., (2004). Microbiological quality of pre-cut fruits, sprouted seeds and unpasteurized fruit from retail and production premises in UK. *Public Health*, 7(3): 567-70.
- [82]. Loiy Elsir Ahmed Hassan; Hasnah Mohd Sirat; Sakina M. Ahemd Yagi1; Waleed, S. Koko; and Siddig Ibrahim Abdelwahab, (2011). In vitro Antimicrobial activities of chloroformic, hexane and ethanolic extracts of *Citrullus lanatus* var. *citroides* (Wild melon). *Journal of Medicinal Plants Research*. 5(8):1338-1344.
- [83]. Lund, B.M., (1992). Ecosystems in vegetable foods. *J Appl Bact.* Vol. 73(21): 115-135.
- [84]. Mabey, R. (1997). *Flora Britannica*. London: Chatto and Windus. p. 298.
- [85]. Mangila E., Tabiliran F.P., Naguit M.R.A., Malate R., (2007). Effects of Organic Fertilizer on the Yield of Watermelon. *Threshold* 2. *January-December, 2007*, pp 27-35.
- [86]. McKenzie, G., (2010). "A Little Bit of History". *Journal of the Bromeliad Society* 39(3):187-189.
- [87]. Meagan, M. Kennelly; Francisco, M. Cazorla; Antonio de Vicente; Cayo Ramos; and George, W. Sundin, (2007). *Pseudomonas syringae*, diseases of fruit trees: Plant Diseases. *The American Phytopathological Society*, 91(1): 4-17.

- [88]. Mensah, P., Owusu-Darko, K., Yeboah-Manu, D., Ablordey, A., Nkrumah, F.K., Kamiya, H., (1999). The role of street food vendors in transmission of enteric pathogens, *Ghana med. J.* 33:19-29.
- [89]. Ministry of Health and Welfare of Japan, (1997). National Institute of Infectious Diseases and Infectious Disease Control Division. Verocytotoxin producing *Escherichia coli* (enterohaemorrhagic *E. coli*) infection, Japan, 1996-June 1997, *Infectious Agents Surveillance Report* 18: 153-154.
- [90]. MMWR-Report. (2001). Norwalk-like Viruses. *Public Health Consequences and Outbreak Management, CDC, Atlanta, Georgia*, 30333. June 1. Vol 50, No RR-9.
- [91]. Moller Nielsen, E., Skov, M.N., Madsen, J.J., Lodal, J., Brochner Jespersen, J. and Baggesen, D.L., (2004). Verocytotoxin-producing *Escherichia coli* in wild birds and rodents in close proximity to farms. *Appl. Environ. Microbiol.*, 70:6944–6947.
- [92]. Muinde, O. K., and Kuria, E., (2005). Hygienic and safety practices of vendors of street foods in Nairobi, Kenya. *A. J. Food Agric. Nutritional Development*, 5:1-18.
- [93]. NACMCF, (1999). National Advisory Committee on Microbiological Criteria for Foods. Microbiological safety evaluations and recommendations on fresh produce. *Food Control* 10: 117-143.
- [94]. NACMCF, (1999). National Advisory Committee on Microbiological Criteria for Foods: Microbiological safety evaluations and recommendations on sprouted seeds. *International Journal of Food Microbiology*, 52: 123-153.
- [95]. Natvig, E.E., Ingham, S.C., Ingham, B.H., Cooperband, L.R. and Roper, T.R., (2002). *Salmonella enterica* serovar typhimurium and *Escherichia coli* contamination of root and leaf vegetables grown in soils with incorporated bovine manure. *Appl. Environ. Microbiol.*, 68:2737–2744.
- [96]. Nester, E.W., Anderson, D.G., Roberts, C.E., Pearsall, N.W. and Nester, M.T., (2010). *Microbiology: A Human Perspective*, (3rd ed). *McGraw Hill Plc., New York*. p. 590.
- [97]. Nicholson, F.A., Groves, S.J. and Chambers, B.J., (2005). Pathogen survival during livestock manure storage and following land application. *Biores. Tech.*, 96:135–143.
- [98]. Nicolle, C., Cardinault, N., Aprikian, O., Buserrolles, J., Grolier, P., Rock, E., Demigné, C., Mazur, A., Scalbert, A., Amouroux, P. and Révész, C., (2003). Effect of Carrot Intake on Cholesterol Metabolism and on Antioxidant Status in Cholesterol-Fed Rat. *European Journal of Nutrition*, 42: 254-261.
- [99]. Nielsen, A.C., (2006). Consumers and ready-to-eat meals: A global ACNielsen report. December 2006. ACNielsen Inc., USA. <http://dk.nielsen.cpm/reports/GlobalRTE ReportDec06.pdf>.
- [100]. Nweze, E.I., (2010). Aetiology of diarrhoea and virulence properties of diarrhoeagenic *Escherichia coli* among patients and healthy subjects in Southeast Nigeria. *J. Health Popul. Nutri.* 28:245-252.
- [101]. Odebisi-Omokanye, M.B., Oke, M.A., Ahmed El-Imam, A. M., Ajijolakewu, A.K. and Salaudeen, B.I., (2015). Microbiological Quality and Safety of pre-cut fruit retailed in Ilorin, Kwara State, Nigeria. *Fountain Journal of Natural and Applied Sciences*, 4(1): 19 – 26.
- [102]. Odoemelan, S.A., (2003). Chemical composition and functional properties of conophor nut flour (*Tetracarpidium conophorum*) flour. *Int. J. Food Sci. Technol.*, 38:729-734.
- [103]. Ogugbue, C.J., Mbakwem-Aniebo, C., Akubuenyi, F., (2011). Assessment of microbial air contamination of post processed gari on sale in markets. *Afri. J. Food Sci.*, 5:503-512.
- [104]. Okafor, J.N.C., Mordi, J.I., Ozumba, A.U., Solomon, H.M. and Olatunji, O., (2003). Preliminary studies on the characterisation of contaminants in tiger nuts (yellow variety). *Proceedings of 27<sup>th</sup> Annual Nigerian Institute of Food and Science and Technology (NIFST), conference*, pp. 210-211.
- [105]. Okonko, I.O., Adejoke, O.D., Ogunnusi, T.A., Fajobi, E.A. and Shittu, O.B., (2008). Microbiological and physicochemical analysis of different water samples used for domestic purposes in Abeokuta and Ojota, Lagos State, Nigeria. *Afri. J. Biotechnol.*, 7:617-621.
- [106]. Okonmah L.U., (2011). Effects of different types of staking and their cost effectiveness on the growth, yield and yield components of cucumber (*Cumumis sativa* L). *Int. J. of Agric. Sci.* Vol. 1 (5): 290-295.
- [107]. Oluwatoyin Afolabi, Adejare Oloyede, Wasiu Abibu, Adeola Adeyanju, (2015). Microbial Safety of Polyethylene Packaged Sliced Fruits Sold in Abeokuta, South-West Nigeria. *Journal of Natural Sciences Research*, 5(13):16-21.
- [108]. Olsen, S.J., MacKinon, L.C., Goulding, J.S., Bean, N.H., Slutsker, L., (2000). Surveillance for Foodborne Disease Outbreaks-United States, 1993-1997. *MMWR* March 17 49(SS01):1-51.
- [109]. Oranusi, S. and Olorunfemi, O.J., (2011). Microbiological safety evaluation of street vended ready-to-eat fruits sold in Ota, Ogun state, Nigeria. *Int. J. Biol. Sci.*, 1:27-32.
- [110]. Oviasogie, F.E., Ogofure, A.G., Beshiru, A., Ode, J.N. and Omeje F.I., (2015). Assessment of fungal pathogens associated with orange fruit spoilage sold in Benin metropolis. *Afr. J. Microbiol. Res.* Vol. 9(29), pp. 1758-1763.
- [111]. Parry, C.M., Hien, T.T., Dougan, G., White, N.J. and Farrar, J.J., (2002). Typhoid fever. *New Engl. J. Med.*, 347:1770-1782.
- [112]. Pascual-Seva, N., San-Bautista, A., López-Galarza, S., Maroto, J.V. and Pascual, B., (2012). Yield and irrigation water use efficacy for ridge and bed cultivated chufa (*Cyperus esculentus* L. *sativus boeck*). *Acta Hort.* 936:125-132.
- [113]. Pasquerella, C., Pitzurra, O. and Savino, A., (2000). The index of microbial air contamination. *J. Hosp. Infect.*, 46:241-256.
- [114]. Patil, M.V., Kandhare, A.D. and Bhise, S.D., (2012). Pharmacological Evaluation of Ethanolic Extract of *Daucus carota* Linn Root Formulated Cream on Wound Healing Using Excision and Incision Wound Model. *Asian Pacific Journal of Tropical Biomedicine*, 2:646-655.
- [115]. Pegg, K.G., (1993). Diseases. In Broadley, R.H., Wassman, R.C. and Sinclair, E. (eds). Pineapple: Pests and Disorders. *Queensland Department of Primary Industries*, Brisbane, pp. 21-29.
- [116]. Rezende, B., Filho, A., Junior, A., Porto, D., (2011). Economic analysis of cucumber and lettuce intercropping under greenhouse in the winter-spring. *Ann. Braz. Acad. Sci.* 83(2):705-717.
- [117]. Rohrbach, K.G., Bartholomew, D.P., Paul, R.E., (2003). Pests, diseases and weeds. In. *CABI*, New York. pp. 203-252.
- [118]. Rohrbach, K.G., Leal, F. and Coppens d'Eeckenbrugge, G. (2003). History, distribution and world production. In: Bartholomew, DP, Paull, RE and Rohrbach, KG (eds). *The Pineapple: Botany, Production and Uses*. *CABI Publishing*, Oxon, UK, pp. 1-12.
- [119]. Rose, F., (2006). *The Wild Flower Key*. London: Frederick Warne. p. 346.
- [120]. Ross, M.A., Lembi, C.A., (2008). Applied Weed Science (3<sup>rd</sup> edition). *Prentice Hall*, pp. 322.
- [121]. Rubatsky, V.E., Quiros, C.F., Siman, P.W., (1999). *Carrots and Related Vegetable Umbelliferae*. pp 6-28.
- [122]. Sánchez-Zapata, E., Fernández-López, J., Angel Pérez-Alvarez, J., (2012). "Tiger nut (*Cyperus esculentus*) commercialisation: Health Aspects, Composition, Properties and Food applications". *Comprehensive Reviews in Food Sci. and Food Safety*, 11:366-377.
- [123]. Sanewski, G. and Scott, C., (2000). The Australian pineapple industry. In: Subhadrabandhu, S. and Chairidchai, P. (eds) Proceedings of the Third International Pineapple Symposium. *International Society for Horticultural Science*, Pattaya, Thailand, pp 53-55.

- [124]. Simon, P.W. and Goldman, I.L., (2007). Carrot. In: Sing, R.J., Ed., *Genetic Resources, Chromosome Engineering, and Crop Improvement*, CRC Press, Boca Raton, pp. 497-516.
- [125]. Simon, P.W., (2000). Domestication, Historical Development and Modern Breeding of Carrot. *Plant Breeding Reviews*, 19:157-190.
- [126]. Simon, Philipp W., Freeman, Roger E., Viera, Jairo V., Boiteux, Leonardo S., Briard, M., Nothnagel, T., Michalik, B., Kwon, Young-Seok, (2008). Carrot: Vegetables II. Handbook of Plant Breeding 2. *New York, NY: Springer*. pp 327-357.
- [127]. Singh, B., Kumar, M., (2006). Techno-economic feasibility of Israeli and indigenously designed naturally ventilated greenhouses for year-round cucumber cultivation. *Acta. Hort.* 710:535-538.
- [128]. Sivapasingham, S., Friedman, C.R., Cohen, L. and Tauxe, R.V., (2004). Fresh produce: a growing cause of outbreaks of foodborne illness in the United States, 1973 through 1997. *J. Food Prot.* 67:2342-2353.
- [129]. Slutsker, L. and Schuchat A., (1999). Listeriosis in humans In: Ryser, E.T., Marth, E.H., eds. *Listeria, Listeriosis, and Food Safety*. *New York: Marcel Dekker*; 75-95.
- [130]. Slutsker, L., Ries, A.A., Greene, K.D., (1997). *Escherichia coli* O157:H7 diarrhoea in the United States: clinical and epidemiologic features. *Annals of Internal Medicine* 126:505-513.
- [131]. Sowonola, O.A., Akintunde, T.Y. and Adedeji, F., (2005). Nutritional and sensory qualities of soymilk- kunnu blends. *African J. Food and Nutr. Sci.*, 5: 1-12.
- [132]. Steele, M. and Odemeru, J., (2004). Irrigation water as a source of foodborne pathogens on fruit and vegetables. *J. Food Prot.*, 67:2839-2849.
- [133]. Stopforth, J.D., Ikeda, J.S., Kendall, P.A. and Stofos, J.N., (2004). Survival of acid-adapted or nonadapted *Escherichia coli* O157:H7 in apple wounds and surrounding tissue following chemical treatments and storage. *Int J Food Microbiol.*, 90:51-61.
- [134]. Swapnil S., Sarvesh, P., Jaya D., and Amita, T., (2011). First report on laxative activity of *Citrullus lanatus*. *Pharmacologyonline*, 2: 790-797.
- [135]. Tambekar, D.H., Jaiswal, V.J., Dhanorkar, D.V., Gulhane, P.B., Dudhane, (2009). Microbial Quality and safety of street vended fruit juices: A case study of Amravati city. *Internet J. Food Safety*, 10: 72 – 76.
- [136]. Tambekar, D.H., Shirsat, S.D., Suradkar, S.B., Rajankar, P.N. and Banginwar, Y.S., (2007). Prevention of transmission of infectious disease: Studies on hand hygiene in health-care among students. *Continental J. Biomedical Sciences* 1: 6-10: 2007.
- [137]. Taulo, S., Westlesen, A., Abrahamsen, R., Mkakosya, R. and Kululanga, G., (2008). Microbiological quality of water, associated management practices and risks at source, transport and storage points in a rural community of Lungwena, Malaysia. *Afri. J. Microbiol. Res*, 7:131-137.
- [138]. Tlili, I., Hdider, C., Lenucci, M.S., (2011). Bioactive compounds and antioxidant activities during fruit ripening of watermelon cultivars. *Journal of Food Composition and Analysis*, 24(7):923-928.
- [139]. Todd, C. Wehner (2008). "Watermelon". In: Jaime Prohens and Fernando Nuez Handbook of plant breeding. *Volume 1, Vegetables. I, Asteraceae, Brassicaceae, Chenopodiaceae, and Cucurbitaceae*. Springer. pp. 381-418.
- [140]. Tyrell, S.F., Knox, J.W. and Weatherhead, E.K., (2006). Microbiological water quality requirements for salad irrigation in the United Kingdom. *J. Food Prot.* 69:2029-2035.
- [141]. Wachtel, M.R., Whitehand, L.C. and Mandrell, R.E., (2002). Association of *Escherichia coli* O157:H7 with preharvest leaf lettuce upon exposure to contaminated irrigation water. *J. Food Prot.* 65:18-25.
- [142]. Wada-Kura, A., Maxwell, R.G., Sadiq, H.Y., Tijani, M.B., Abdullahi, M.O., Aliyu, M.S. and Adetunji, O.A., (2009). Microbiological quality of some ready-to-eat foods and formules in some cafeterias in Ahmadu Bello University, Zaria. *Biol. Environ. Sci. J. Trop.*, 6:6-9.
- [143]. Wallace, J.S., Cheasty, T. and Jones, K., (1997). Isolation of verocytotoxigenic producing *Escherichia coli* O157 from wild birds. *J. Appl. Microbiol.*, 82:399-404.
- [144]. Walls, I. and Scott, V.N., (1997). Use of predictive microbiology in microbial food safety risk assessment. *Int. J. Food Microbiol.*; 36(2-3):97-102.
- [145]. Wells, J.M. and Butterfield, J.E., (1999). Incidence of *Salmonella* on fresh fruits and vegetables affected by fungal rots or physical injury. *Plant Diseases*, 83:722-726.
- [146]. Zaini, R., Clench, M.R. and Maitre, C.L. (2011). Bioactive Chemicals from Carrot (*Daucus carota*) Juice Extracts for the Treatment of Leukemia. *Journal of Medicinal Food*, 14:1303-1312.
- [147]. Zhang, D. and Hamauzu, Y., (2004). Phenolic Compounds and Their Antioxidant Properties in Different Tissues of Carrots (*Daucus carota* L.). *Journal of Food, Agriculture and Environment (JFAE)*, 2: 95-100.
- [148]. Zohary, M., Daniel, M. and Hopf, M., (2000). Domestication of plants in the Old World (3<sup>rd</sup> edition). *Oxford University Press*, pp. 193.