

# Comparative Evaluation Of Antibacterial Efficacy Of Metapex, Metapex With Nanosilverfluoride, Metapex With Chitosan Against Streptococcus Mutans And Enterococcus Faecalis As Obturating Material:An Invitro Study

Dr Hemanthakrishna Shivala, Dr Neena I E, Dr Mallikarjuna Kenchappa

(Postgraduate, Department Of Pediatric And Preventive Dentistry, College Of Dental Sciences, Davangere, India)

(Professor, Department Of Pediatric And Preventive Dentistry, College Of Dental Sciences, Davangere, India)

(Professor And Head, Department Of Pediatric And Preventive Dentistry, College Of Dental Sciences, Davangere, India)

---

## Abstract:

**Background:** Successful endodontic treatment in primary teeth relies heavily on effective elimination of pathogenic microorganisms, particularly *Streptococcus mutans* and *Enterococcus faecalis*, which are commonly implicated in persistent root canal infections. Metapex, a calcium hydroxide and iodoform-based paste, is widely used as an obturating material due to its resorbable and antimicrobial properties. However, limitations in its spectrum of antimicrobial action have led to the exploration of enhanced formulations. Nanotechnology-based additives such as Nanosilverfluoride and Chitosan have shown promising antibacterial effects due to their high surface area and sustained release capabilities. Incorporating these agents into Metapex may improve its efficacy against resistant endodontic pathogens.

**Materials and Methods:** This in vitro study was conducted in the Department of Pediatric and Preventive Dentistry, Davangere, in collaboration with Maratha Mandal Dental College, Belgaum. *Streptococcus mutans* (MTCC 890) and *Enterococcus faecalis* (MTCC 3159) strains were used. Three obturating materials were tested: Metapex, Metapex with Nanosilverfluoride (NSF), and Metapex with Chitosan, prepared in a 3:1 ratio. The antibacterial efficacy was assessed using the agar diffusion method. Standardized wells were filled with test materials and incubated anaerobically. Zones of inhibition were measured after 24 hours using a Vernier calliper to evaluate antimicrobial activity.

**Results:** Metapex showed the largest inhibition zone against *Streptococcus mutans* (mean: 37.73 mm), followed by NSF (23.73 mm) and Chitosan (14.33 mm). In contrast, for *Enterococcus faecalis*, NSF had the highest zone (34.47 mm), followed by Chitosan (25.60 mm), and Metapex (14.80 mm). ANOVA and Tukey's post-hoc tests confirmed significant differences between all groups ( $p < 0.001$ ). NSF significantly enhanced the efficacy of Metapex against *Enterococcus faecalis*, a more resistant organism, while plain Metapex was more effective against *Streptococcus mutans*.

**Conclusion:** The study concluded that while Metapex alone is highly effective against *Streptococcus mutans*, its efficacy against *Enterococcus faecalis* improves markedly when combined with Nanosilverfluoride. NSF exhibited the most balanced and broad-spectrum antibacterial action. Metapex with Chitosan showed moderate effectiveness, particularly against *Enterococcus faecalis*. These findings suggest that modifying obturating materials with Nanosilverfluoride or Chitosan may enhance their antimicrobial potential and clinical applicability in pediatric endodontics.

**Key Word:** Antibacterial efficacy, Agar diffusion method, Chitosan, *Enterococcus faecalis*, Metapex, Nanosilverfluoride, Obturating material, *Streptococcus mutans*, Pediatric endodontics, Primary teeth

---

Date of Submission: 19-09-2025

Date of Acceptance: 29-09-2025

---

## I. Introduction

Pulp therapy preserves primary teeth by removing infected pulp<sup>1</sup>. *E. faecalis* resists treatment and survives in root canals. *S. mutans*, a key caries pathogen, thrives in sucrose-rich environments and is inhibited by fluoride.

Pulpectomy is preferred for primary teeth with endodontic infections<sup>2,3,4</sup>. Early loss of these teeth can cause malocclusion and eruption issues, so retaining them until exfoliation is crucial<sup>5,6</sup>.

Ideal root canal filling materials should be bacteriostatic, resorbable, non-irritating, radiopaque, and easy to insert/remove. No current material meets all criteria. Zinc oxide eugenol and calcium hydroxide are common but have limitations. Calcium hydroxide-iodoform paste (e.g., Vitapex) is safe and resorbable but may cause internal resorption<sup>7,8,9</sup>.

Chitosan is biocompatible and antimicrobial, disrupting bacterial membranes and supporting biomedical use<sup>10</sup>. Chitosan nanoparticles (CS NPs), by virtue of their charge and size, are expected to possess enhanced antibacterial activity. In addition, Chitosan possesses several characteristics such as being nontoxic toward mammalian cells, colour compatibility to tooth structure, cost effectiveness, availability, and ease of chemical modification<sup>11</sup>.

Silver nanoparticles (AgNPs), or nano-silver, are nanoparticles of silver of between 1 nm and 100 nm in size containing 20 to 15,000 silver atoms. They have been one of the most popular topics of study in recent decades because of their outstanding antimicrobial activity even at low concentrations including antibacterial, antifungal, antiviral, and anti-inflammatory effects<sup>12</sup>. Silver nanoparticles (AgNPs) penetrate cell walls, disrupt DNA, and are highly effective against *S. mutans* due to their small size<sup>13</sup>.

## **II. Material And Methods**

The study was done in the Department of Pediatric and Preventive Dentistry, College of Dental Sciences, Davangere in collaboration with Maratha Mandala Dental College, Belgaum. Microbial type culture collection (MTCC) Chandigarh from where the maintained strains of *Streptococcus mutans* (MTCC 890) and *Enterococcus faecalis* (MTCC 3159) were collected. Ethical clearance was obtained from institutional review board of the college for the study.

**Study Design:** An in vitro comparative study

**Study Location:** Department of pediatric and preventive dentistry College of dental sciences Davangere.

**Study Duration:** 6months

**Sample size:** 90 samples.

**Sample size calculation:** The sample size was estimated to be 15 per each group, based on a 95% confidence level, an observed standard deviation of 0.2, and a permissible error margin of 10%.

### **Subjects & selection method:**

Metapex(MetaBiomed), Chitosan (Vedayukt India pvt limited), Nanosilverfluoride (Vedayukt India pvt limited), Agar plate with Muller Hinton agar,

Group A (*Streptococcus mutans*)

A1-Metapex (control group)

A2 -Metapex with nanosilverfluoride (experimental group)

A3-Metapex with chitosan (experimental group)

Group B (*Enterococcus faecalis*)

B1-Metapex (control group)

B2-Metapex with nanosilverfluoride (experimental group)

B3-Metapex with chitosan (experimental group)

### **Procedure methodology**

About 150 mg of Metapex was diluted in 1 mL of DMSO (dimethyl sulphoxide). To which, 50 mg of silver fluoride nanoparticle (M+SF) and Chitosan nanoparticle (M+C) were added separately in each microcentrifuge tube. After vigorous mixing of both the samples using a vortex, the sample was taken as a test compound. Metapex mixed with DMSO was considered as control (M).

### **Preparation of *S. mutans* and *E.Faecalis* cultures:**

Preparation of broth and bacterial growth:

The antimicrobial activity of obturating materials used in primary teeth against *streptococcus mutans* and *Enterococcus faecalis* was evaluated in this study by agar diffusion method. The standard bacterial strain of

streptococcus mutans (MTCC890) and enterococcus faecalis (MTCC3159) obtained from microbial type culture collection (MTCC) Chandigarh. The purity of test strain was confirmed using the Gram's stain. Only 0.37 grams of Brain Heart Infusion (BHI) broth was to 10mL of distilled water and mixed by gently shaking the container. This mixture was sterilized by autoclaving for 15 minutes at 121°C and 15lb pressure and then allowed to cool at room temperature. The bacterial strains are inoculated in BHI broth and incubated at 37°C for 24 hours. Following incubation, the cultures were centrifuged at 3000 rpm for 10 minutes. The supernatant liquid was discarded and the precipitate containing microbial cells was separated from the base of test tube. The precipitate containing microbial cells was re-suspended in saline and turbidity of this culture suspension was adjusted until it is equivalent to the no.1 McFarland Standard (Approximately  $3 \times 10^8$  cells/mL).

Preparation of culture medium:

Mueller Hinton agar (15.2gm) was added to 400mL of distilled water and mixed by gently shaking the container. This mixture was sterilized by autoclaving for 15 minutes at 121°C and 15lb pressure. After 15 minutes, the liquid was cooled to room temperature. In a laminar flow chamber, this liquid medium was poured in 20 petri dishes of size 90mm and allowed to set. Inoculation of bacterial strain on culture media: Each agar plate with 20mL of Mueller Hinton agar was inoculated with 0.1mL microbial suspension using sterile swab. The 3 wells (4mm of depth X 6mm of diameter) were made in each of the agar plates, i.e., total 60 wells were prepared in 20 agar plates to test two different obturating materials and a control.

Placement of obturating materials and incubation:

Each freshly mixed experimental obturating material was placed in 10 wells of different petri dishes. These agar plates were then incubated at 37°C under anaerobic conditions in an incubator using Microaerophilic (candle jar system). An anaerobic indicator tablet (Anaerobic indicator tablets) was placed in the jar to monitor oxygen contamination of the environment.

Measuring the size of zone of inhibition:

A lack in bacterial colonization was observed for each obturating material. It was indicated by growth inhibitory zones (clearing of agar) around each obturating material. The most uniform diameter segment of the zone of inhibition was determined in millimeters by measuring the shortest distance between the outer margin of the well and initial microbial growth after 24 hours of incubation.

### Statistical analysis

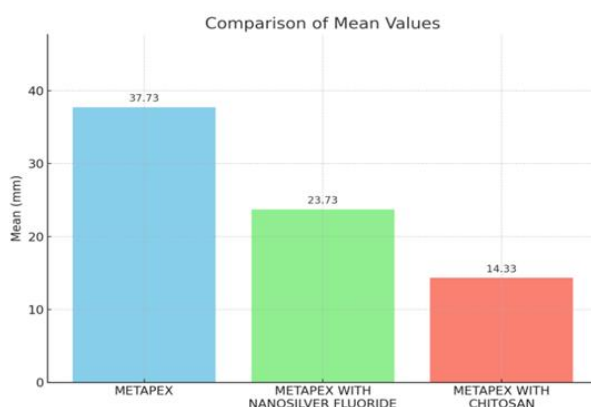
The data obtained was statistically analyzed by ANOVA, POST -HOC analysis

## III. Result

**TABLE 1: Explains mean value, standard deviation, standard error, 95% Confidence Interval for Mean Streptococcus mutans.**

Streptococcus mutans								
	N	Mean (mm)	Std. Deviation	Std. Error	95% Confidence Interval for Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
Metapex	15	37.73	7.658	1.977	33.49	41.97	28	55
Metapex With Nanosilver Fluoride	15	23.73	2.492	.643	22.35	25.11	20	28
Metapex With Chitosan	15	14.33	2.526	.652	12.93	15.73	10	18

**GRAPH 1: mean zone of inhibition for streptococcus mutans between groups**



The Metapex group exhibited the highest mean zone of inhibition, with a mean value of 37.73 mm and a standard deviation of 7.658 mm. The 95% Confidence Interval (CI) for this group ranged from 33.49 mm to 41.97 mm, with individual inhibition zones ranging between 28 mm and 55 mm.

The Metapex with Nanosilver Fluoride group showed a moderate inhibitory effect, with a mean zone of 23.73 mm and a standard deviation of 2.492 mm. The 95% CI for this group was 22.35 mm to 25.11 mm, with the minimum and maximum inhibition zones recorded as 20 mm and 28 mm, respectively.

In contrast, the Metapex with Chitosan group displayed the least antimicrobial activity, with a mean inhibition zone of only 14.33 mm and a standard deviation of 2.526 mm. The 95% CI ranged from 12.93 mm to 15.73 mm, with a minimum of 10 mm and a maximum of 18 mm.

**TABLE 2: Explains ANOVA of Streptococcus mutans zone of inhibition among three groups.**

Streptococcus mutans					
	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	4159.600	2	2079.800	87.597	<0.001*
Within Groups	997.200	42	23.743		
Total	5156.800	44			

\*<0.05 considered statistically significant

One-way Analysis of Variance (ANOVA) revealed a statistically significant difference in the *Streptococcus mutans* inhibition zones across the three groups. The between-group sum of squares was 4159.600 with 2 degrees of freedom, resulting in a mean square value of 2079.800. The F-value was 87.597 with a p-value of <0.001, indicating that the differences among group means were highly significant ( $p < 0.05$ ).

**Table 3: Explains Multiple Comparisons Of Streptococcus Mutans Zone Of Inhibition Using Post-Hoc Tukey.**

Dependent Variable: Streptococcus Mutans						
Tukey Hsd						
(I) Groups	(J) Groups	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
Metapex	Metapex With Nanosilver Flouride	14.000*	1.779	<0.001*	9.68	18.32
	Metapex With Chitosan	23.400*	1.779	<0.001*	19.08	27.72
Metapex With Nanosilver Flouride	Metapex	-14.000*	1.779	<0.001*	-18.32	-9.68
	Metapex With Chitosan	9.400*	1.779	<0.001*	5.08	13.72
Metapex With Chitosan	Metapex	-23.400*	1.779	<0.001*	-27.72	-19.08
	Metapex With Nanosilver Flouride	-9.400*	1.779	<0.001*	-13.72	-5.08

\*. The Mean Difference Is Significant At The 0.05 Level.

Post-hoc analysis using the Tukey HSD test further confirmed the statistical significance of the differences among all pairs of groups:

- Metapex (Group A1) v/s Metapex with Nanosilver Fluoride (Group A2): The mean difference was 14.000 mm, which was statistically significant ( $p < 0.001$ ), with a 95% CI ranging from 9.68 mm to 18.32 mm.
- Metapex (Group A1) v/s. Metapex with Chitosan (Group A3): The mean difference was 23.400 mm, also statistically significant ( $p < 0.001$ ), with a 95% CI from 19.08 mm to 27.72 mm.
- Metapex with Nanosilver Fluoride (Group A2) v/s. Metapex with Chitosan (Group A3): The mean difference was 9.400 mm, again statistically significant ( $p < 0.001$ ), with a 95% CI ranging from 5.08 mm to 13.72 mm.

All intergroup differences were statistically significant with p-values less than 0.001, suggesting that the choice of additive plays a critical role in determining the antibacterial effectiveness of Metapex.

**Table 4, Explains Mean Value, Standard Deviation, Standard Error, 95% Confidence Interval For Mean Of Enterococcus Faecalis**

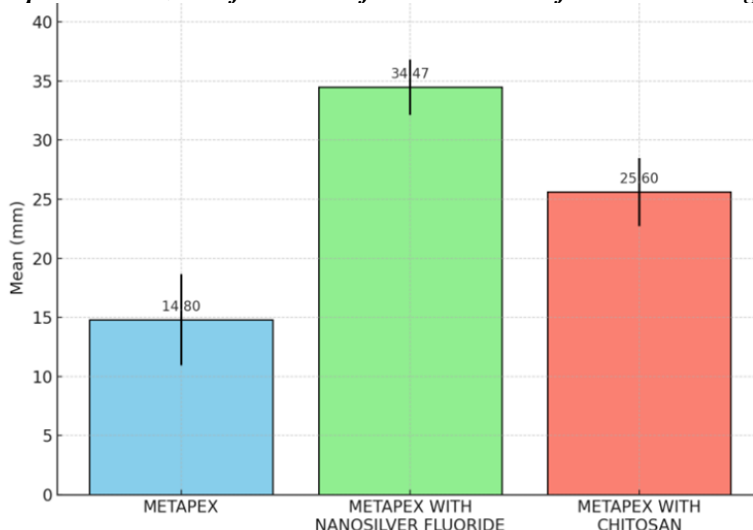
Enterococcus Faecalis								
	N	Mean (Mm)	Std. Deviation	Std. Error	95% Confidence Interval For Mean		Minimum	Maximum
					Lower Bound	Upper Bound		
Metapex	15	14.80	3.840	.991	12.67	16.93	5	20
Metapex With Nanosilver Flouride	15	34.47	2.326	.601	33.18	35.75	31	38
Metapex With Chitosan	15	25.60	2.874	.742	24.01	27.19	21	32

The Metapex with Nanosilver Fluoride group showed the highest mean zone of inhibition, with a mean of 34.47 mm and a standard deviation of 2.326 mm. The 95% Confidence Interval (CI) for the mean ranged from 33.18 mm to 35.75 mm, and the values ranged from 31 mm to 38 mm.

All intergroup comparisons were statistically significant ( $p < 0.001$ ), highlighting that the incorporation of **Nanosilver Fluoride** or **Chitosan** into Metapex significantly enhances its antimicrobial efficacy, with **Nanosilver Fluoride** providing the greatest enhancement.

- The Metapex with Chitosan group exhibited a moderate antibacterial effect, with a mean zone of inhibition of 25.60 mm, a standard deviation of 2.874 mm, and a 95% CI ranging from 24.01 mm to 27.19 mm. The minimum and maximum inhibition zones observed were 21 mm and 32 mm, respectively.
- In contrast, the Metapex group demonstrated the least inhibitory effect, with a mean inhibition zone of 14.80 mm and a standard deviation of 3.840 mm. The 95% CI ranged from 12.67 mm to 16.93 mm, with individual values ranging between 5 mm and 20 mm.

**Graph 2: Mean zone of inhibition for *Enterococcus faecalis* between groups**



**TABLE 5: Explains ANOVA of *Enterococcus faecalis* zone of inhibition among three groups.**

Enterococcus faecalis					
	Sum of Squares	df	Mean Square	F	Sig.
Between Groups	2910.178	2	1455.089	153.655	<0.001*
Within Groups	397.733	42	9.470		
Total	3307.911	44			

A one-way ANOVA revealed a highly statistically significant difference in the *Enterococcus faecalis* inhibition zones across the three groups. The between-group sum of squares was 2910.178 (df = 2), with a mean square value of 1455.089. The F-statistic was 153.655 and the associated p-value was <0.001, confirming that the observed differences among group means are statistically significant ( $p < 0.05$ ).

**TABLE 6: Explains multiple comparisons of *Enterococcus faecalis* zone of inhibition among three groups.**

Dependent Variable: Enterococcus Faecalis						
Tukey Hsd						
(I) Groups	(J) Groups	Mean Difference (I-J)	Std. Error	Sig.	95% Confidence Interval	
					Lower Bound	Upper Bound
Metapex	Metapex With Nanosilver Flouride	-19.667*	1.124	<0.001	-22.40	-16.94
	Metapex With Chitosan	-10.800*	1.124	<0.001	-13.53	-8.07
Metapex With Nanosilver Flouride	Metapex	19.667*	1.124	<0.001	16.94	22.40
	Metapex With Chitosan	8.867*	1.124	<0.001	6.14	11.60
Metapex With Chitosan	Metapex	10.800*	1.124	<0.001	8.07	13.53
	Metapex With Nanosilver Flouride	-8.867*	1.124	<0.001	-11.60	-6.14

\*. The Mean Difference Is Significant At The 0.05 Level.

Post-hoc analysis using the Tukey HSD test showed the following significant pairwise differences:

- Metapex (GROUP B1) v/s Metapex with Nanosilver Fluoride (GROUP B2): The mean difference was -19.667 mm, which was statistically significant ( $p < 0.001$ ), with a 95% CI from -22.40 mm to -16.94 mm.
- Metapex (GROUP B1) v/s. Metapex with Chitosan (GROUP B3): The mean difference was -10.800 mm, also statistically significant ( $p < 0.001$ ), with a 95% CI from -13.53 mm to -8.07 mm.
- Metapex with Nanosilver Fluoride (GROUP B2) v/s Metapex with Chitosan (GROUP B3): The mean difference was 8.867 mm, again statistically significant ( $p < 0.001$ ), with a 95% CI ranging from 6.14 mm to 11.60 mm.

#### IV. Discussion

Effective disinfection of primary root canals remains a major challenge due to complex anatomy and the persistence of microorganisms such as *Streptococcus mutans* and *Enterococcus faecalis*. *S. mutans* initiates caries, while *E. faecalis* is linked to persistent endodontic infections and periradicular pathologies owing to virulence factors and resistance to high pH and antimicrobial agents<sup>14,15</sup>.

This study compared the antibacterial efficacy of Metapex, Metapex + Nanosilverfluoride (NSF), and Metapex + Chitosan.

**Metapex:** A calcium hydroxide–iodoform paste, Metapex exerts antimicrobial activity through high pH but shows limited effect against *E. faecalis*<sup>16,17,18,19</sup>. Prior studies report moderate efficacy, with better action against *S. mutans* than *E. faecalis*<sup>20,21</sup>. Our findings corroborate these results, showing greater inhibition of *S. mutans* (mean 37.73 mm) compared to *E. faecalis* (14.80 mm).

**Metapex + NSF:** Silver nanoparticles provide potent antimicrobial action by disrupting bacterial membranes and biofilms<sup>22,23,24</sup>. NSF demonstrated significant efficacy against resistant pathogens like *E. faecalis*, consistent with previous studies. In our study, Metapex + NSF showed the highest antibacterial activity, likely due to nanoparticle penetration and sustained ion release.

**Metapex + Chitosan:** Chitosan disrupts microbial membranes and exhibits bioadhesive properties enhancing retention in canals. Studies confirm its effectiveness against *E. faecalis* and biofilms<sup>25,26,27</sup>. In combination with Metapex, chitosan enhanced antibacterial efficacy compared to Metapex alone, though slightly less than the NSF group.

**Comparative Analysis:** Overall, Metapex + NSF > Metapex + Chitosan > Metapex alone. The superior effect against *E. faecalis* is particularly relevant for preventing treatment failures. Incorporating novel agents like NSF and chitosan improves the antibacterial potential of traditional obturating materials.

**Strengths and Limitations:** Strengths include standardized methodology, reliable microbial strains, and evaluation of clinically significant pathogens. However, in vitro conditions cannot replicate oral environment factors such as saliva, host response, or polymicrobial infections. Agar diffusion, while simple, may not reflect sustained intracanal activity.

**Future Directions:** In vivo studies considering biofilms, host factors, and long-term outcomes are warranted. Further exploration of nanomaterial- or biopolymer-modified obturating agents may enhance pediatric endodontic success.

#### V. Conclusion

This in vitro study demonstrated that Metapex alone was most effective against *Streptococcus mutans*, while its efficacy against *Enterococcus faecalis* improved significantly when combined with Nanosilverfluoride. Among the tested groups, Metapex + Nanosilverfluoride showed the highest and broadest antibacterial activity, followed by Metapex + Chitosan, with Metapex alone being least effective against resistant pathogens. Incorporating novel agents such as Nanosilverfluoride or Chitosan into conventional obturating materials may enhance disinfection in pediatric endodontics. Further clinical studies are required to validate these findings and assess long-term biocompatibility.

#### References

- [1]. Navit S, Jaiswal N, Khan SA, Malhotra S, Sharma A, Jabeen S, Et Al. Antimicrobial Efficacy Of Contemporary Obturating Materials Used In Primary Teeth—An In-Vitro Study. J Clin Diagn Res. 2016 Sep 1;10(9):ZC09.
- [2]. Bodrumlu E, Semiz M. Antibacterial Activity Of A New Endodontic Sealer Against Enterococcus Faecalis. J Can Dent Assoc. 2006 Sep 1;72(7):637–9.

- [3]. Madiba M, Oluremi BB, Gulube Z, Oderinlo OO, Marimani M, Osamudiamen PM, Et Al. Anti-Streptococcus Mutans, Anti-Adherence And Anti-Acidogenic Activity Of Uvaria Chamae P. Beauv. J Ethnopharmacol. 2023 Jan 10;1(7):76–9.
- [4]. Sundqvist G, Figdor D, Persson S, Sjögren U. Microbiologic Analysis Of Teeth With Failed Endodontic Treatment And The Outcome Of Conservative Re-Treatment. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 1998 Jan 1;85(1):86–93.
- [5]. Shindova M. Root Canal Filling Materials In Primary Teeth—Review. Folia Med (Plovdiv). 2021 Oct 31;63(5):657–62.
- [6]. Shakti P, Singh A, Purohit BM, Purohit A, Taneja S. Effect Of Premature Loss Of Primary Teeth On Prevalence Of Malocclusion In Permanent Dentition: A Systematic Review And Meta-Analysis. Int Orthod. 2023 Dec 1;21(4):100816.
- [7]. Grossman LI, Oliet S, Del Rio CE. Endodontic Practice. 11th Ed. Philadelphia: Lea & Febiger; 1988.
- [8]. Sharma R, Garg R, Kaushik M. A Review On Obturating Materials For Deciduous Dentition. Int J Adv Res. 2022 Oct;10(10):387–93.
- [9]. Gupta S, Das G. Clinical And Radiographic Evaluation Of Zinc Oxide Eugenol And Metapex In Root Canal Treatment Of Primary Teeth. J Indian Soc Pedod Prev Dent. 2011 Jul 1;29(3):222–8.
- [10]. Sanap P, Hegde V, Ghunawat D, Patil M, Nagaonkar N, Jagtap V. Current Applications Of Chitosan Nanoparticles In Dentistry: A Review. Int J Appl Dent Sci. 2020;6(4):81–4.
- [11]. Radhakrishnan A, Panicker UG. Sustainable Chitosan-Based Biomaterials For The Future: A Review. Polym Bull. 2025 Feb;82(3):661–709.
- [12]. Jangid H, Singh S, Kashyap P, Singh A, Kumar G. Advancing Biomedical Applications: An In-Depth Analysis Of Silver Nanoparticles In Antimicrobial, Anticancer, And Wound Healing Roles. Front Pharmacol. 2024 Aug 8;15:1438227.
- [13]. Essawy MM, Al Achy SN, Talaat DM, El-Tekeya MM, Essa S, Nabil N, Et Al. Fluoridated Silver Nanocomposites For Caries Management: An In-Vitro Assessment Of The Cytological And Antibacterial Profiles. BMC Oral Health. 2025 Mar 9;25(1):363.
- [14]. Stuart CH, Schwartz SA, Beeson TJ, Owatz CB. Enterococcus Faecalis: Its Role In Root Canal Treatment Failure And Current Concepts In Retreatment. J Endod 2006 Feb;32(2):93–8.
- [15]. Loesche WJ. Role Of Streptococcus Mutans In Human Dental Decay. Microbiol Rev 1986 Dec;50(4):353–80
- [16]. Siqueira JF Jr, Lopes H. Mechanisms Of Antimicrobial Activity Of Calcium Hydroxide: A Critical Review. Int Endod J 1999 Sep;32(5):361–9. 49.
- [17]. Jain G, Sharma S, Rastogi S, Rajkumar B, Boruah LC. In-Vitro Evaluation Of Antimicrobial Efficacy Of Triple Antibiotic Paste, Metapex And Newly Introduced Iodine Based Asphalene Temp As Intracanal Medicament Against Enterococcus Faecalis. Int J Health Sci 2022;6(S1):568–73. 50.
- [18]. Dahlen G, Moller AJR. Microbiology Of Endodontic Infections. In: Slots J, Taubman MA, Editors. Contemporary Oral Microbiology And Immunology. St Louis: Mosby; 1992. P. 444–75. Page 74 Bibliography 51.
- [19]. Evans M, Davies JK, Sundqvist G, Figdor D. Mechanisms Involved In The Resistance Of Enterococcus Faecalis To Calcium Hydroxide. Int Endod J 2002 Mar;35(3):221–8.
- [20]. Ibrahim H, Khattab N. Assessment Of Antibacterial Efficacy Of Different Obturation Materials For Primary Teeth (An In Vitro Study). Egypt Dent J 2021 Jan 1;67(1):139 43. 39.
- [21]. Paranna S, Biradar J, Semwal M, Patil K, Suradkar S, Shinde S. Comparative Evaluation Of Postoperative Pain In Primary Teeth Obturated With Zinc Oxide Eugenol Versus Metapex: A Randomized Clinical Trial. Mymensingh Med J 2022 Oct 1;31(4):1148–52.
- [22]. Morones JR, Elechiguerra JL, Camacho A, Holt K, Kouri JB, Ramirez JT, Yacaman MJ. The Bactericidal Effect Of Silver Nanoparticles. Nanotechnology 2005 Aug 26;16(10):2346–53. 57.
- [23]. Nguyen S, Hiorth M. Advanced Drug Delivery Systems For Local Treatment Of The Oral Cavity. Ther Deliv 2015 May;6(6):595–608. 58.
- [24]. Sockanto SA, Marpaung LJ, Himmatshohwah, Djais A, Darwita RR. Efficacy Of Propolis Fluoride And Nano Silver Fluoride For Inhibition Of Streptococcus Mutans And Enterococcus Faecalis Biofilm Formation. Int J Appl Pharm 2017;9(Special Issue 2):51–4.
- [25]. Goy RC, Britto DD, Assis OB. A Review Of The Antimicrobial Activity Of Chitosan. Polímeros 2009;19(3):241–7. 60.
- [26]. Iqbal K, Alhomrany R, Berman LH, Chogle S. Enhancement Of Antimicrobial Effect Of Endodontic Sealers Using Nanoparticles: A Systematic Review. J Endod 2023 Oct;49(10):1238–48. 64. Pandey A, Bhushan J, Joshi RK, Uppal AS,
- [27]. Angrup A, Kansal S. Comparative Evaluation Of Antimicrobial Efficacy Of Chitosan Nanoparticles And Calcium Hydroxide Against Endodontic Biofilm Of Enterococcus Faecalis: An In Vitro Study. J Conserv Dent Endod 2024 Jul 1;27(7):750–4. 65.: An In Vitro Study. Acta Odontol Scand 2019 Jan 2;77(1):39–43.

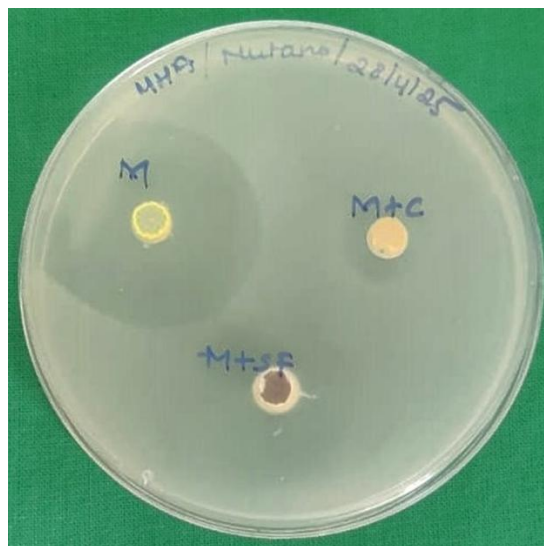


Figure 1: zone of inhibition on streptococcus mutans by Metapex, Metapex with nanosilverfluoride, Metapex with chitosan.

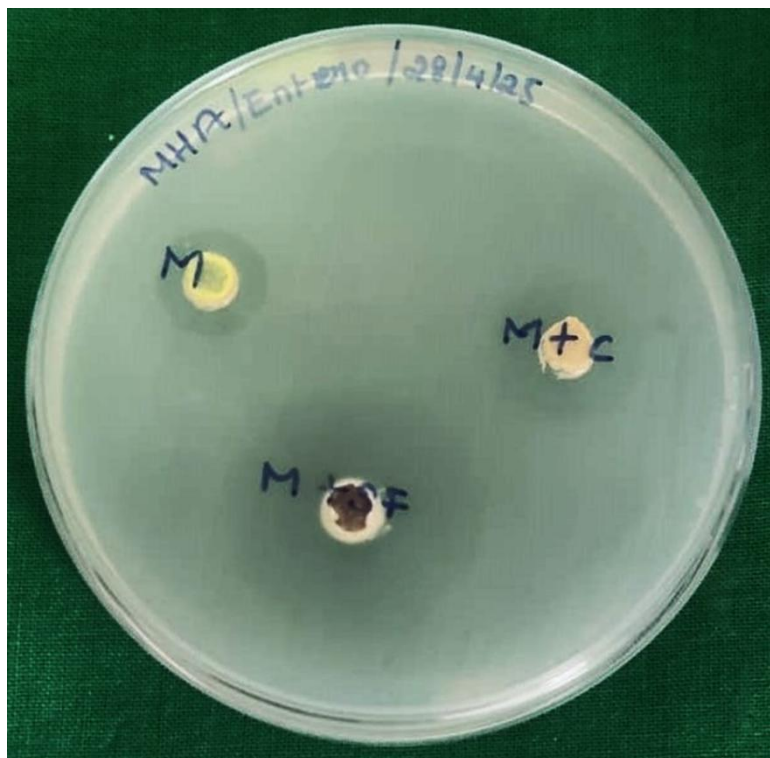


Figure 2: Zone of inhibition on Enterococcus Faecalis by Metapex, Metapex with nanosilverfluoride, Metapex with chitosan