

## Detection of Cancer Stem Cells in the Most Common Head and Neck Non-Hodgkin's Lymphoma Using CD34: in Vitro Study

Hadya N. Ghedan<sup>1</sup>, Samia M. El-Azab<sup>2</sup>, Safa F. Abd El-Ghany<sup>3</sup>, Mona A. Saker<sup>4</sup>

<sup>1</sup>B.D.S, Faculty of Oral and Dental Medicine, Cairo University, Egypt

<sup>2</sup>Oral and maxillofacial pathology, Faculty of Oral and Dental Medicine, Cairo University, Egypt

<sup>3</sup>Oral & Maxillofacial Pathology, Faculty of Oral and Dental Medicine, Cairo University, Egypt

<sup>4</sup>Pathology department, National Cancer Institute, Cairo University, Egypt

Corresponding Author: Hadya N. Ghedan

### Abstract:

**Background.** Non-Hodgkin Lymphomas (NHL) are a heterogeneous group of malignancies of the immune system; diffuse large B-cell lymphoma and Burkitt's lymphoma are among its most common subtypes. Recently, it is widely believed that cancer stem cells play critical roles in the initiation, metastasis, and relapse of cancers. However, no sufficient data supported the presence of cancer stem cells in NHL of the head and neck.

**Purpose.** To detect cancer stem cells in diffuse large B-cell lymphoma and Burkitt's lymphoma using immunohistochemical expression CD34, a widely used marker for identification and detection of hematopoietic stem cells.

**Methodology.** The present work was performed on archival paraffin blocks of diffuse large B-cell lymphoma and Burkitt's lymphoma. The mean count and mean area of the positive CD34 cells were assessed by the image analyzer computer system using the software Leica Qwin 500.

**Results.** The results showed positive CD34 immunoreaction in 90% of the diffuse large B-cell lymphoma cases, and 70% of the Burkitt's lymphoma cases, denoting presence of cancer stem cells in both tumors. Diffuse large B-cell lymphoma showed significantly higher mean count and mean area of positive CD34 cells than those of Burkitt's lymphoma.

**Conclusion.** Cancer stem cells are present in both diffuse large B-cell lymphoma and Burkitt's lymphoma and may be a common origin of these lesions. Diffuse large B-cell lymphoma may be more aggressive than Burkitt's lymphoma. The level of cancer stem cells may be a valuable prognostic tool which is directly proportional with more aggressive behavior and poor prognosis. CD34 is a valuable cancer stem cell marker in diffuse large B-cell lymphoma and Burkitt's lymphoma and can be used as prognostic marker in these lesions.

**Key words:** CD34, Non-Hodgkin's Lymphoma (B-Cell lymphoma and Burkitt's lymphoma), Cancer Stem Cells

Date of Submission: 29-05-2019

Date of acceptance: 15-06-2019

### I. Introduction

Non-Hodgkin Lymphomas (NHLs) are heterogeneous group of lymphoproliferative disorders originating from B-lymphocytes, T-lymphocytes or Natural Killer (NK) lymphocytes.<sup>1, 2</sup> Diffuse large B-cell lymphoma (DLBCL) has been reported as the most common subtype of aggressive B-cell lymphoma worldwide, which accounted for 25–30% of adult NHL and showed considerable biologic and clinical heterogeneity. In Egypt, it has also been reported as the most common subtype constituting 49% of cases as reported in the National Cancer Institute (NCI) Cancer Pathology Registry.<sup>3,4</sup>

On the other hand, Burkitt's lymphoma (BL) was first described by Dermis Burkitt (1958) in African children with large jaw in relation to Epstein-Barr virus. It is a highly aggressive B-cell neoplasm which has been characterized by the translocation and deregulation of the c-myc gene on chromosome 8. It has been reported as a rapidly growing neoplasm requiring immediate diagnosis and treatment.<sup>5,6</sup>

Cancer stem cells (CSCs), also known as tumor-initiating cells, or tumor propagating cells are cancer cells that possess capability of proliferation, differentiation, and self-renewal. It was widely believed that CSCs played critical roles in the initiation, metastasis, and relapse of cancers.<sup>7,8</sup> Moreover; they may have possible roles in cancer therapy. However, specific recognition of these cells from tumour masses has represented the first challenge. CSCs have been detected and isolated using various methods, including overexpression of specific cell surface markers. Among these surface markers, CD34 has been widely used as a marker for identification and isolation of hematopoietic stem cells (HSCs).<sup>9,10,11</sup> It is a 105-120-kDa transmembrane glycoprotein which was first identified in 1984 on hematopoietic stem and progenitor cells.<sup>12,13</sup>

To date, no sufficient data supports the presence of CSCs in lymphoma, although they were detected in many cancers.<sup>14, 15</sup> Since, the lymphoma stem cell hypothesis has remained largely unexplored, it seems interesting to detect the presence of CSCs in lymphoma. This could contribute to better understanding of the role of CSCs in development, tumorigenesis, prognosis and treatment of oral NHL.

## **II. Material And Methods**

### **2.1. Tissue specimens**

A total of 30 specimens of NHL from cervical lymph nodes were retrieved from the archival paraffin blocks from files of Pathology Department, NCI, Cairo University. They comprised DLBCL (20 cases) and BL (10 cases).

### **2.2. Histopathological and IHC staining**

All the procedures took place at pathology lab, NCI, Cairo University.

Thin (4 microns) paraffin sections of each tissue specimen were stained with Haematoxylin and Eosin (H&E) to reconfirm the diagnosis and the most representative sections were selected. Paraffin sections were mounted on positively charged glass slides (Optiplus, Biogenex, USA) for immunostaining.

The initial immunohistochemical panel for lymphomas [CD-3, CD-20, CD10 and terminal deoxynucleotidyltransferase (TDT) antibodies, Ventana Medical Systems, USA] was used to confirm the diagnosis. CD-3 (T-cell marker, rabbit monoclonal antibody, catalogue#790-4341) and CD20 (B-cell marker, mouse monoclonal antibody, catalogue#760-2531) were used for both DLBCL and BL, to confirm the B cell origin and exclude the T-cell one. Furthermore, CD-10 (rabbit monoclonal antibody, catalogue#790-4506) and TDT (rabbit polyclonal antibody, catalogue#760-2670) were used on the BL cases to identify whether the B cells were mature or immature. Staining for each antibody was reported as positive or negative immunoreaction.

Then, the selected cases were stained by anti-CD34 mouse monoclonal antibody (catalogue#790-2927) and iVIEW detection kit (catalogue#760-091) (Ventana Medical Systems, USA), using the Bench Mark XT system (Ventana autostainer, USA).

### **2.3. Assessment of immunostaining**

All the cells that showed positive CD34 immunoreactivity were considered as CSCs. They were detected by brownish color in the cytoplasm and cell membrane. Immunostaining of CD34 was assessed in Research Unit of Oral and Maxillofacial Pathology Department, Faculty of Oral and Dental Medicine, Cairo University. Image analyzer computer system was used for counting of CD34 positive cells and measuring area of positive CD34 immunoreaction of tumor cells.

### **2.4. Counting of CD34 positive cells**

The number of CD34 positive cells on magnification (X400) was counted in 5 randomly chosen fields using manual counter across the entire sample by two investigators.

#### **2.4.1 Measuring area of positive reaction of tumor cells**

Areas of positive CD34 immunoreaction in tumour cells were chosen for evaluation, and the positive reaction in blood vessels was excluded.

#### **2.4.2. Statistical analysis**

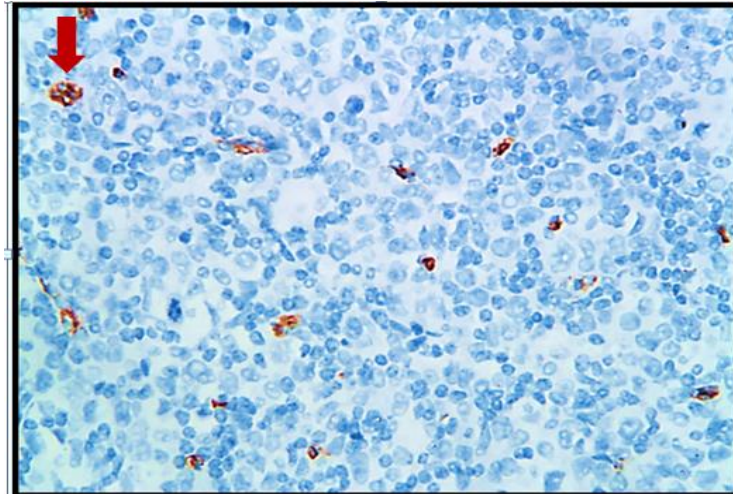
The data obtained from the image analyzer computer system were tabulated and statistically analyzed. Data was presented as mean  $\pm$  standard deviation (SD) values. Student's t test was used for comparing mean count and mean area of positive CD34 immunoreaction between both DLBCL and BL. The P values were considered as the following: not significant (P value > 0.05), significant (P value  $\leq$  0.05), or highly significant (P value  $\leq$  0.01).

## **III. Results**

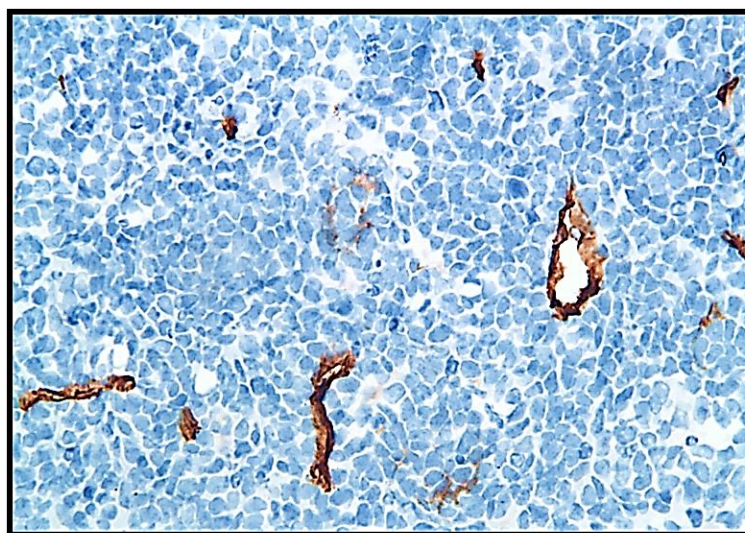
### **3.1. Immunohistochemical findings**

All the examined cases of DLBCL and BL (100%) showed positive CD20 immunoreaction and negative CD3 immunoreaction, denoting their B cell origin. The BL cases (100%) showed positive CD10 and negative TDT immunoreaction, denoting that the involved B-cells were mature.

Positive CD34 immunoreactivity was detected in 18 out of 20 examined DLBCL cases (90%), and 70% of BL cases (7/10). Cytoplasmic and membranous CD34 immunoreaction was restricted to CSCs and endothelial cells. The CSCs were either sporadic or arranged in small groups (Fig. 1 and 2).



**Fig. 1.** A photomicrograph of DLBCL showing strong membranous immunoreaction of CD34 in CSCs, which are either sporadic or in small groups (arrow), anti-CD34



**Fig. 2.** A photomicrograph of BL showing strong immunoreaction in blood vessels lined by endothelial cells, anti-CD34 antibody, ×400.

### 3.2. Statistical analysis

Both the mean count and the mean area of CD34 immunoreactivity were higher in DLBCL than BL, which reflects the increased number of CSCs in DLBCL than BL (Table 1) (Fig. 3 and 4).

**Table 1.** The meancount & mean area of CSCs in both DLBCL and BL Groups.

	Group	Mean ±SD	P value
Mean count	DLBCL	10.90±2.77	0.0119*
	BL	7.30±2.98	
Mean area	DLBCL	269.29±81.34	0.0087**
	BL	167.28±73.61	

\* P < 0.05 Significant

\*\* P < 0.01 highly Significant

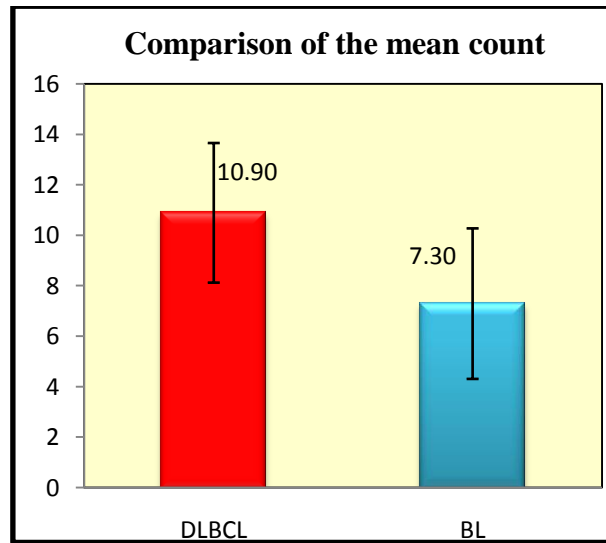


Fig (3): Bar chart showing the mean count of CSCs in both DLBCL and BL groups.

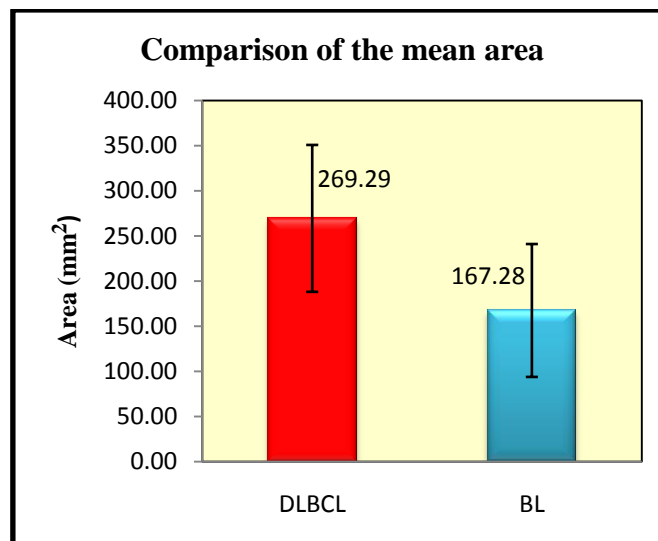


Fig (4): Bar chart showing mean area of CSCs in both DLBCL and BL groups.

#### IV. Discussion

Two models were proposed to explain tumor development: the conventional “stochastic model” and the “CSC model”. The former propounds that every cancer cell has the ability to be the founder of a new tumor. Recently, the “CSC model” has proposed that cancer evolution occurs from tissue progenitor or stem cells which have a “deregulated” self-renewal pathway. Several studies confirmed the potential role of CSCs in the pathogenesis of cancers. However, no sufficient data supported the presence of CSCs in NHL.<sup>16,17</sup>

Various methods have been used for detection and isolation of CSCs; however cell surface marker detection has shown a major advantage of obtaining a precise population compared to other methods. In the present study, CD34 immunostaining was precisely localized to a small population of cells which were considered as CSCs. This was in agreement with what has been reported by Sasseti et al. (2000) and Pisacane et al. (2007), who pointed out that expression of CD34 was maintained in the undifferentiated phenotype of progenitor cells and was not present on mature hematopoietic cells.<sup>18,19</sup>

In the present study, CSCs were detected successfully in the most common subtypes of NHL. These cells could play an important role in the development, tumorigenesis and prediction of prognosis of these lesions. Moreover, both DLBCL and BL might share a common origin which is CSCs derived from HSCs that are responsible for tumor growth and formation of both CSCs as well as the non tumorigenic progeny that form bulk of tumor mass.<sup>16</sup>

Presence of CSCs is in agreement with the results of Bonnet and Dick (1997), Cozzio et al. (2003) and Janikova et al. (2009) who confirmed presence of CSCs in hematopoietic malignancies.<sup>20,21,22</sup> Also, Nakashima et al. (2010), Shafer et al. (2010) and Ikeda et al. (2010) pointed out that HL had CSCs.<sup>23,24,25</sup>

Similarly, Vega et al. (2010) and Jung et al. (2011) supported the existence of lymphoma SCs in studied mantle cell lymphoma.<sup>26,27</sup> More recently, Kim (2011) and Lee et al. (2013) supported that B-cell NHL could be enriched with lymphoma SCs.<sup>28,29</sup>

In the same context, CSCs have now been isolated from several human malignancies, as lung tumors (Gibbs et al., 2005), bone sarcomas (Gutova et al., 2007) and pancreatic carcinomas (Hermann et al., 2007).<sup>30,31,32</sup> In contradiction to our results, Kim et al. (2013) failed to detect CSCs in canine lymphoma.<sup>33</sup>

The findings of the present study also shed the light on the prognostic value of CSCs as a sign of more aggressive behavior and poor prognosis. That's why we propose that DLBCL could have a more aggressive behavior than BL as it showed higher mean count and mean area for CD34 positive cells, denoting an increased number of CSCs. This could explain the different invasion potentials of these lesions and recommending different treatment modalities for each of them (Diaz-Cano, 2012).<sup>34</sup>

Also, Kim (2011) and Lee et al. (2013) reported the association of CSCs with malignant potential of B-cell lymphomas.<sup>28,29</sup> In the same context, Zhang et al. (2008), Beier et al. (2008), Jiang et al. (2009), Su et al. (2010), Liu et al. (2015) and Su et al. (2015) agreed that higher CSC levels and high expression of CSC markers were associated with advanced disease stage, high histologic grade, and poor clinical outcome.<sup>35, 36,37,38,39,40</sup>

On the contrary, some studies proposed that the increase in CSCs had a favorable effect on tumor behavior as Costa et al. (2012) who reported that low expression of CSC markers was associated with higher probability of disease progression and death.<sup>41</sup> Similarly, Kasem et al. (2012) found that solid pattern of adenoid cystic carcinoma was associated with low expression of CSCs marker.<sup>42</sup>

## V. Conclusions

CSCs are present in both DLBCL and BL and may be a common origin of these lesions. DLBCL may be more aggressive than BL. The level of CSCs may be a valuable prognostic tool which is directly proportional with more aggressive behavior and poor prognosis. CD34 is a valuable CSC marker in DLBCL and BL and can be used as prognostic marker in these lesions.

## References

- [1]. **Mokhtar, N. (2007):** Review article composite lymphoma. *J Egypt Natl Canc Inst.*, 19(3): 171–175.
- [2]. **Zelenetz, A.; Gordon, L.; Wierda, W.; Abramson, J.; Advani, R.; Andreadis, C. and Sundar, H. (2014):** Non-Hodgkin's lymphomas, version 2.2014. *J Natl Compr Canc Netw.*, 12(6): 916–946.
- [3]. **Mokhtar, N. and Khalid, H. (2002):** Lymphoma. The National Cancer Institute, Cairo University, pp 123-141, Egypt.
- [4]. **Seshadri, T.; Kuruvilla, J.; Crump, M., and Keating, A. (2008):** Salvage therapy for relapsed/refractory diffuse large B cell lymphoma. *Biol Blood Marrow Transplant*, 14(3): 259–267.
- [5]. **Coakley, D. (2006):** Denis Burkitt and his contribution to haematology/oncology. *Br J Haematol.*, 135(1): 17–25.
- [6]. **Bellan, C.; Stefano, L.; Giulia, D.; Rogena, E. and Lorenzo, L. (2009):** Burkitt lymphoma versus diffuse large B-cell lymphoma: a practical approach. *Hematol Oncol.*, 27(4): 182–185.
- [7]. **Bu, Y. and Cao, D. (2012):** The origin of cancer stem cells. *Front Biosci.*, 4(1), 819–830.
- [8]. **Karamboulas, C., and Ailles, L. (2013):** Developmental signaling pathways in cancer stem cells of solid tumors. *Biochim Biophys Acta*, 1830(2): 2481–2495.
- [9]. **Klonisch, T.; Wiehac, E.; Hombach-Klonisch, S.; Ande, S.; Wesselborg, S.; Schulze-Osthoff, K. and Los, M. (2008):** Cancer stem cell markers in common cancers- therapeutic implications. *Trends Mol Med*, 14(10): 450-460.
- [10]. **Kazi, R.; Sayed, S. and Dwivedi, R. (2010):** Cancer stem cells: an enigma in head and neck cancer. *J Cancer Res Ther*, 6(4): 411–413.
- [11]. **Mackie, A. and Losordo, D. (2011):** CD34-positive stem cells: in the treatment of heart and vascular disease in human beings. *Tex Heart Inst J.*, 38(5): 474–485.
- [12]. **Pusztaszeri, M.; Seelentag, W. and Bosman, F. (2006):** Immunohistochemical expression of endothelial markers CD31, CD34, von willebrand factor, and Fli-1 in normal human tissues. *J Histochem Cytochem.*, 54(4): 385–395.
- [13]. **Karsten, U. and Goletz, S. (2013):** What makes cancer stem cell markers different? *Springerplus.*, 2(1): 301–309.
- [14]. **Gross, E.; Quillet-Mary, A.; Ysebaert, L.; Laurent, G. and Fournie, J.-J. (2011):** Cancer stem cells of differentiated B-cell malignancies: models and consequences. *Cancers*, 3(2): 1566–1579.
- [15]. **Khot, K. and Deshmane, S. (2015):** Oral cancer stem cells: drivers of local recurrence and metastasis. *Clin Cancer Investig J.*, 4(6): 691-695.
- [16]. **Al-Hajj, M. and Clarke, M. (2004):** Self-renewal and solid tumor stem cells. *Oncogene*, 23(43): 7274–7282.
- [17]. **Morrison, S. and Spradling, A. (2008):** Stem cells and niches: mechanisms that promote stem cell maintenance throughout life. *Cell*, 132(4): 598–611.
- [18]. **Sasseti, C.; Van Zante, A. and Rosen, S. (2000):** Identification of endoglycan, a member of the CD34/podocalyxin family of sialomucins. *J Biol Chem.*, 275(12): 9001–9010.
- [19]. **Pisacane, A.-M.; Picciotto, F. and Risio, M. (2007):** CD31 and CD34 expression as immunohistochemical markers of endothelial transdifferentiation in human cutaneous melanoma. *Cell Oncol.*, 29(1): 59–66.
- [20]. **Bonnet, D. and Dick, J. (1997):** Human acute myeloid leukemia is organized as a hierarchy that originates from a primitive hematopoietic cell. *Nat Med.*, 3(7): 730–737.
- [21]. **Cozzio, A.; Passegué, E.; Ayton, P.; Karsunky, H.; Cleary, M. and Weissman, I. (2003):** Similar MLL-associated leukemias arising from self-renewing stem cells and short-lived myeloid progenitors. *Genes Dev.*, 17(24): 3029–3035.
- [22]. **Janikova, A.; Mayer, J.; Kren, L.; Smardova, J.; Dvorakova, D.; Neubauer, J. and Vasova, I. (2009):** The persistence of t(14;18)-bearing cells in lymph nodes of patients with follicular lymphoma in complete remission: the evidence for “a lymphoma stem cell”. *Leuk Lymphoma*, 50(7): 1102–1109.

- [23]. Nakashima, M.; Ishii, Y.; Watanabe, M.; Togano, T.; Umezawa, K.; Higashihara, M. and Horie, R. (2010): The side population, as a precursor of Hodgkin and Reed-Sternberg cells and a target for nuclear factor- $\kappa$ B inhibitors in Hodgkin's lymphoma. *Cancer Sci.*, 101(11): 2490–2496.
- [24]. Shafer, J.; Cruz, C.; Leen, A.; Ku, S.; Lu, A.; Rousseau, A. and Foster, A. (2010): Antigen-specific cytotoxic T lymphocytes can target chemoresistant side-population tumor cells in Hodgkin lymphoma. *Leuk Lymphoma*, 51(5): 870–880.
- [25]. Ikeda, J.; Mamat, S.; Tian, T.; Wang, Y.; Rahadiani, N.; Aozasa, K. and Morii, E. (2010): Tumorigenic potential of mononucleated small cells of Hodgkin lymphoma cell lines. *Am J Pathol.*, 177(6): 3081–3088.
- [26]. Vega, F.; Davuluri, Y.; Cho-Vega, J.; Singh, R.; Ma, S.; Wang, R.-Y. and Ford, R. (2010): Side population of a murine mantle cell lymphoma model contains tumour-initiating cells responsible for lymphoma maintenance and dissemination. *J Cell Mol Med.*, 14(6B): 1532–1545.
- [27]. Jung, H.; Chen, Z.; McCarty, N. and Carty, N. (2011): Stem-like tumor cells confer drug resistant properties to mantle cell lymphoma. *Leuk Lymphoma*, 52(6): 1066–1079.
- [28]. Kim, S. (2011): Lymphoma stem cells: a step toward a new therapeutic target. *Korean J Hematol.*, 46(4): 211–213.
- [29]. Lee, M.; Ju, H.-J.; Kim, B.; Ko, Y.; Kim, W. and Kim, S. (2013): Isolation of side population cells in B-cell non-Hodgkin's lymphomas. *Acta Haematol.*, 129(1): 10–27.
- [30]. Gibbs, C.; Kukekov, V.; Reith, J.; Tchigrinova, O.; Suslov, O.; Scott, E. and Steindler, D. (2005): Stem-like cells in bone sarcomas: implications for tumorigenesis. *Neoplasia*, 7(11): 967–976.
- [31]. Gutova, M.; Najbauer, J.; Gevorgyan, A.; Metz, M.; Weng, Y.; Shih, C.-C. and Aboody, K. (2007): Identification of uPAR-positive chemoresistant cells in small cell lung cancer. *PLoS One*, 2(2): e243.
- [32]. Hermann, P.; Huber, S.; Herrler, T.; Aicher, A.; Ellwart, J.; Guba, M. and Heeschen, C. (2007): Distinct populations of cancer stem cells determine tumor growth and metastatic activity in human pancreatic cancer. *Cell Stem Cell*, 1(3): 313–323.
- [33]. Kim, M.-C.; D'Costa, S.; Suter, S. and Kim, Y. (2013): Evaluation of a side population of canine lymphoma cells using Hoechst 33342 dye. *J Vet Sci.*, 14(4): 481–486.
- [34]. Diaz-Cano, S. (2012): Tumor heterogeneity: mechanisms and bases for a reliable application of molecular marker design. *Int J Mol Sci.*, 13(2): 1951–2011.
- [35]. Zhang, M.; Song, T.; Yang, L.; Chen, R.; Wu, L.; Yang, Z. and Fang, J. (2008): Nestin and CD133: valuable stem cell-specific markers for determining clinical outcome of glioma patients. *J Exp Clin Cancer Res.*, 27(1): 85–92.
- [36]. Beier, D.; Wischhusen, J.; Dietmaier, W.; Hau, P.; Proescholdt, M.; Brawanski, A. and Beier, C. (2008): CD133 expression and cancer stem cells predict prognosis in high-grade oligodendroglial tumors. *Brain Pathol.*, 18(3): 370–377.
- [37]. Jiang, F.; Qiu, Q.; Khanna, A.; Todd, N.; Deepak, J.; Xing, L. and Katz, R. (2009): Aldehyde dehydrogenase 1 is a tumor stem cell-associated marker in lung cancer. *Mol Cancer Res.*, 7(3): 330–338.
- [38]. Su, S.; Hong, F.; Liang, Y.; Zhou, J.; Liang, Y.; Chen, K. and Peng, L. (2015): Lgr5 methylation in cancer stem cell differentiation and prognosis-prediction in colorectal cancer. *PLoS One*, 10(11): e0143513.
- [39]. Liu, R.; Shen, Y.; Nan, K.; Mi, B.; Wu, T.; Guo, J. and Guo, H. (2015): Association between expression of cancer stem cell markers and poor differentiation of hepatocellular carcinoma: a meta-analysis (PRISMA). *Medicine*, 94(31): e1306.
- [40]. Su, Y.; Qiu, Q.; Zhang, X.; Jiang, Z.; Leng, Q.; Liu, Z. and Jiang, F. (2010): Aldehyde dehydrogenase 1 A1-positive cell population is enriched in tumor-initiating cells and associated with progression of bladder cancer. *Cancer Epidemiol Biomarkers Prev.*, 19(2): 327–337.
- [41]. Costa, W.; Rocha, R.; Cunha, I.; Fonseca, F.; Guimaraes, G. and Zequi, S. (2012): CD133 immunohistochemical expression predicts progression and cancer-related death in renal cell carcinoma. *World J Urol*, 30(4): 553–558.
- [42]. Kasem, R.; Awad, R. and Hegazy, R. (2012): Identification of normal and cancerous salivary stem cells using CD44. *EDJ*, 58(1): 337-343.

Hadya N. Ghedan. "Detection of Cancer Stem Cells in the Most Common Head and Neck Non-Hodgkin's Lymphoma Using CD34: in Vitro Study." *IOSR Journal of Dental and Medical Sciences (IOSR-JDMS)*, vol. 18, no. 6, 2019, pp 64-72.