

Plant Parasitic Nematodes Parasitism On The Nutritional Status Of Yam (*Dioscorea* Species) Tubers In Nigeria.

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Abstract

Yam; *Dioscorea* species of crops are multivariant staples in Nigeria and are plagued by plant parasitic nematodes (PPNs). Four species of *Dioscorea*; *D. rotundata* (white yam), *D. cayenensis* (yellow yam), *D. alata* (water yam), *D. domestica* were selected to determine the relationship between PPNs load and the nutritional status of the tubers. Proximate analyses of the tubers were carried out using standard scientific methods while the Bearmann's and Sieving methods were used to determine the PPNs load of the tubers. *D. cayenensis* (38.89%) had the highest nematode load while *D. domestica* (8.47%) was the least infected. PPNs load decreased as tuber depth increased (outermost layers; 77.4% and the innermost; 0%). *Scutellonema* species (48.0%) was the most prevalent PPN in all the yam tubers. Moisture content of the stored tubers influenced the mobility of PPNs in tubers. However, moisture content of *D. cayenensis* and *D. rotundata* declined as nematode load decreased while that of *D. alata* and *D. domestica* show no association between PPN load and moisture content of the yam tubers. PPNs parasitism influenced crude carbohydrate and crude protein concentrations of yam tubers. Fibre content of the tubers varied in relation to the specific tuber layers. *D. domestica* was resistant to PPNs in the study. The long shelf life of yam tubers made them reservoir hosts of numerous plant parasitic nematodes which promoted the dispersal of the worm pests. Pre-planting sterilization of tubers is recommended to curb the transmission of PPNs to new agroenvironments.

Keywords: *Dioscorea* spp., nematode load, crude protein, tuber depth and reservoir hosts and agroenvironment.

Date of Submission: 10-03-2021

Date of Acceptance: 25-03-2021

I. Introduction

There is severe food scarcity and insecurity in sub-Saharan Africa, greatly occasioned by the deterioration of farm produce both in the field and storage resulting from pest attacks (FAO, 2009) and poor storage facility. Majority of the crops are attacked in the field, while others are attacked in storage. One of the pests that occur in both the field and in storage is the plant parasitic nematode (Amusa and Mohammed, 2003; Mwaniki, 2003; Williams and Gleason, 2003; Ogaraku, A.O. and Usman, 2008; Nzeako *et al.*, 2013). Nematodes; the round worms are ubiquitous, diverse and highly adaptable to almost any niche in the aquatic and terrestrial ecosystems (Ferris, and Berkelman, 2003 and Nzeako *et al.*, 2015). Their ecological flexibility of nematodes makes them one of the most successful animals on earth, however, their variable ecological roles accentuate them as suitable indicators of intricate alterations in the ambient physicochemical status of the environment. Nematodes' diverse trophic affiliations in the ecosystem makes some species extreme obligate biotrophs that cause huge losses to crops in the field and storage (Coursey, 1967; Nweke, 1991; GCDDT, 2014, Nzeako *et al.*, 2016 and 2019). Plant parasitic nematodes (PPN) are specialist obligates of crops and constitute a minute segment of the ambiguous set of nematoparasites that attack living things. PPNs are responsible for about 2.3% of the world annual agro-yield loss of economic crops (Nayak *et al.*, 1987, Kushwaha and Polycarp, 2001; Abad and Williamson, 2008; Shehu *et al.*, 2010; Ibitoye and Attah, 2012). All plant taxa are infected by plant parasitic nematodes, however, some show great host specificity such as *Ditylenchus dipsaci*, *Anguinatritici* and *Scutellonema* spp., while some are great generalist parasites such as *Meloidogyne* spp., and *Pratylenchus* spp., (Nzeako and Imafidor, 2010). Nematode invasion and migration in the plant tissues alters the root architecture, causes significant reductions in nutrients and water uptakes and consequently hampers crop yields (Curtis *et al.*, 2007).

Amongst the PPNs of great economic importance in Nigeria is *Scutellonema* spp., that is specific to the *Dioscorea* spp., of crops (yam) (Sasser, 1980, Adesiyun, *et al.*, 1990). Generally, phytoparasitic nematodes impact negatively on the growth and development of *Dioscorea* spp., resulting to poor yields and marketability of tubers. Yam; *Dioscorea* spp., in all its varieties is a major staple in Nigeria and west African sub region

(Adegbite,2006; Nzeako,*et al.*, 2010.). Nigeria remains the highest producer of the crop in ton per hecter worldwide but not the highest beneficiary in terms of foreign exchange acruing from its export (Page, 2006, GCDT, 2014, NRCRI, 1998, FAO, 2007, 1998, Izekor and Olumese,2010, Nsikak *et al.*, 2013). Nigeria’s premier status in yam production isnot due to improved yield per hectre cultivated nor due to improved planting materials, rather, due to increased population of farmers embanking on yam production in the last five decades (Mwaniki, 2003; FAO, 2009, 2010, Shehu *et al.*, 2010; Ibitoye and Attah, 2012). Yam as a crop has huge socioeconomic significance in Nigeria as it constitutes one of the important items in traditional marriage ceremony,child birth, burial and religious rites of the populace (Nzeako, *et al.*, 2010, Ibitoye and Onimissim, 2013, GCDT, 2014).

The wide geographical tolerance or adaptation of the *Diocorea* spp.,of crops in sub Saharan African had made it a crop for all the regions including Nigeria, especially, at the subsistent level. Also, the economic returns from yam farming and its unique role in the management of the health state; hypertension had made it a dominant target crop for supplemental vocation for people from all works of life in Nigeria. Yam is not a staple for the poorin the population, since, its consumption is more in the high socioeconomic group (GCDT, 2014). It has been recognised as a suitable crop for poverty alleviation interventions because of its reletively long shelf-lifethat is beyound twelve months that enables it to be stocked during harvest periods when it is abundant and cheap (Onwueme, 1978,Nweke, 1991; Nzeako *et al.*, 2015). This characteristic makes yam a highly valued cash crop, usually stored for up to 8 months and sold by poor farmers when the supply is low.

The impact of PPNs infetation of yam in the field results to great decrease in tuber sizeand tissue integrity in storage. This is a major contributor to very low agricultural performance in terms of productivity within the Nigerian agroecosystem. Low agricultural productivity hampers the effort of the government to achieving food security and sustainability in the country especially, for a crop of high sociocultural and economic status like *Dioscorea* spp. To this end,it isimportant to determine the post harvest nutritional status of naturally infected tubers in a yam producing region of Rivers State, Nigeria. Again, it is speculated that PPNs load of yam tubers influences the overall crude protein content of individual yam tubers (Caillaud *et al.*, 2008; Wei and Brent, 2006). This propositionimplies that; the higher the PPNs load, the higher the protein content of the yam tuber orvizvisa. However, this assertion needs validation. This study is designed to determine the nutritional status of PPNs infected yam tubers from Rivers State, Nigeria.The study will also; determine the relationship between Tuber Nematode Load (TNL) in relation to crude carbohydrate, crude protein, dietary fibre, and moisture contents of four yam species; *D. rotundata*, *D. cayenensis*, *D. alata* and *D. dometerum*.This study would unravel the misconception associated with PPNs load in relation to the protein, carbohydrate, moisture and fibre contents of yam tubers in storage.

Table 1. The nutritional content of yam

S/N	Nutrient (%/g/mg)	Value	S/N	Nutrient (%/g/mg)	Value
1	Moisture (%)	81.00-165.0	8	Phosphorus (mg)	17-61.0
2	Calories (%)	71.00-135.00	9	Iron (mg)	8.0-12.0
3	Carbohydrate (g)	16-31.8	10	Potassium (mg)	214-397
4	Protein (g)	1.4-3.5	11	B-carotene (mg)	0.0-10.0
5	Fat (g)	0.2-0.4	12	Riboflavin (mg)	0.01-0.011
6	Fibre (g)	0.40-10.0	13	Niacin (mg)	0.01-0.04
7	Calcium (g)	12-69.	14	Ascorbic acid (mg)	4.00-18.0

Standard nutritional content of 1g of yam as stated by FAO

II. Materials and Methods

2.1. Study Area and Collection of Samples

The study was carried out in four communities; Beeri,Taaba, Kaani and Nyo-buru in Khana L.G.A of RiversState. The Local Government Area is located between4.700’N and 7.350’E, occupies an area of 560km2 andhas a population of 294.217 (National census Commission, 2006). The studyarea experiences an annual maximum temperature of32°C and a minimum of 27°C. The average annualrainfall of the area is 2500 mm with July andSeptember having the highest. The vegetation of thestudy area is the tropical rainforest type. Samples of *D. rotundata* (white yam), *D. cayenensis* (yellow yam), *D. alata* (water yam), *D. dometerum* (three-leaf yam or the African bitter yam) were randomly collected fromthe study area. These were placed in appropriately labeled polythene bags and transported to the laboratory for analyses.

2.1.1. Exclusion Criteria: This work considered only stored tubers of yam and seasonal variation was not considered.

2.2. Extraction of plant parasitic nematodes of the yam tissues

Individual seed yams were washed in running tap to remove soil and debris. These were cut into 3 parts (anterior, middle and posterior). On each of the designated parts; concentric circular parts were cut off to represent the outer, middle and core of the sectioned parts of the yam tuber. The cut circular sections were macerated and placed on the modified Bearman's sieve for the extraction of endophytic nematodes (Hooper, 1970, Baker *et al.*, 1985, Hooper *et al.*, 2007, Golden, 1985a and b). The extraction setup was left for 48 hours after which the macerates were discarded, 10% formalin was added to the aliquots and allowed to sediment, after which the supernatant of approximately 90% was decanted. The sediment of about 10% was poured into labelled universal bottles for microscopic examination. Nematode identification was done using Goodey and Goodey (1963).

2.3. Determination of Crude Protein Content

Protein analysis of the yam tuber was ascertained following Kjeldahl Method as stated by the Analytical Organization of Analytical Chemists, (AOAC, 2016) which involved digestion, neutralization and titration techniques. Determination of protein was based on total N content \times , a factor = 6.25.

2.4. Determination of Carbohydrate content: This was done using the Clegg Method (AOAC, 2000). Method is on difference: 100 - (weight in grams [protein + fat + water + ash + alcohol] in 100 g of food) Measured in mg/ml.

2.5. Determination of Moisture Content: This was done using the Air Oven method as described in AOAC (2016).

2.6. Lipid Content Determination- Soxhlet Extraction Method as stated in AOAC (2016).

2.7. Ash Content Determination- The Furnace Method as described in AOAC (2016).

2.8. Nematode Identification

Nematodes were identified according to Goodey and Goodey (1963) and Golden (1985).

III. Results

3.1 Table 2. Plant Parasitic Nematodes population in yam tubers

The plant parasitic nematode load of the yam tubers revealed the occurrence of five nematode species; *Meloidogyne* spp., *Scutellonema* spp., *Hoplioliamus* spp., *Pratylenchus* spp. and *Helicotylenchus* spp., at varying intensities in the various *Dioscorea* species sampled. There was variability in the endophytic nematode load and specificity amongst the yam varieties with *D. cayensis* harbouring the highest load of 69 (38.98%), *D. alata*; 47 (26.56%), *D. rotundata*; 46 (25.98%) and *D. domestorum*; having 15 (8.47%). Although, the occurrence of the five nematode species recorded in the study were consistent amongst all the *Dioscorea* spp. tubers; *Scutellonema* spp., had the highest parasite density while the least density was recorded for *Helicotylenchus* spp. *Dioscorea* varieties were in this study were suitable hosts to plant parasitic nematodes with varying levels of resistance and tolerance. *D. domestorum* was regarded as resistant to plant parasitic nematodes as it harboured the lowest density of endophytic worms in the study (Table 2) which could be due to its unsuitability to nematode reproductivity (Figure 1). However, the other three sampled yam varieties displayed variability in their suitability to plant parasitic nematodes population.

Table 2. Plant Parasitic Nematodes population in yam tubers

Nematode species	Yam Species (%)				Total (%)
	<i>D. cayensis</i>	<i>D. alata</i>	<i>D. domestorum</i>	<i>D. rotunda</i>	
<i>Meloidogyne</i> spp.	22(42.30)	15(28.84)	2(3.84)	13(25.00)	52(29.71)
<i>Scutellonema</i> spp.	31(36.90)	22(26.19)	9(10.71)	22(26.19)	84(48.00)
<i>Hoplioliamus</i> spp.	5(33.33)	3(20.00)	2(13.33)	5(33.33)	15(8.57)
<i>Pratylenchus</i> spp.	6(35.29)	5(29.41)	2(11.76)	4(23.52)	17(9.71)
<i>Helicotylenchus</i> spp.	5(55.55)	2(22.22)	0(0.00)	2(22.22)	9(5.14)
Total (%)	69(38.98)	47(26.56)	15 (8.47)	46(25.98)	177

2.9. Specific Tuber layer protein content

Mean crude protein concentration of the yam tubers in relation to yam tuber specific layer and nematode load showed a decrease in protein concentration as nematode load increased. This pattern of distribution of plant parasitic nematode was observed in all the yam tubers except *D. cayensis* where the outermost layer contained more protein than the inner layers. The concentration of crude protein along the three considered layers of the tubers (specific tuber layer protein content) varied within the tubers and across the yam

varieties. There was a surface to core variation in the concentration of protein in the tubers examined. Variability in crude protein levels of the various yam layers gradually increased from surface to the core of the tubers in three *Dioscorea* species except the *D.dometurum* (table 5). Nematode load decreased as depth increased with the outermost layer harbouring 77.4% of the overall nematode load and the innermost; the core bearing no worms in all the yam species. This trend influenced protein concentration in the tubers examined. In all yam tubers protein content was lowest at the outermost surface except in *D.dometurum* where the outermost layer bore the highest protein concentration. This implied that tissue degeneration occurred in the region with high parasitic load such that protein depreciated. However, host specific factors may have contributed to the relatively high protein load of *D. dometurum* in addition to plant parasitic nematode resistance attributed to the yam species. There was no significant difference in protein concentration of at specific tuber layers across the yam varieties ($p>0.05$) however, the study revealed that the heavily infected tubers harboured lesser carbohydrate than the relatively less infected tubers.

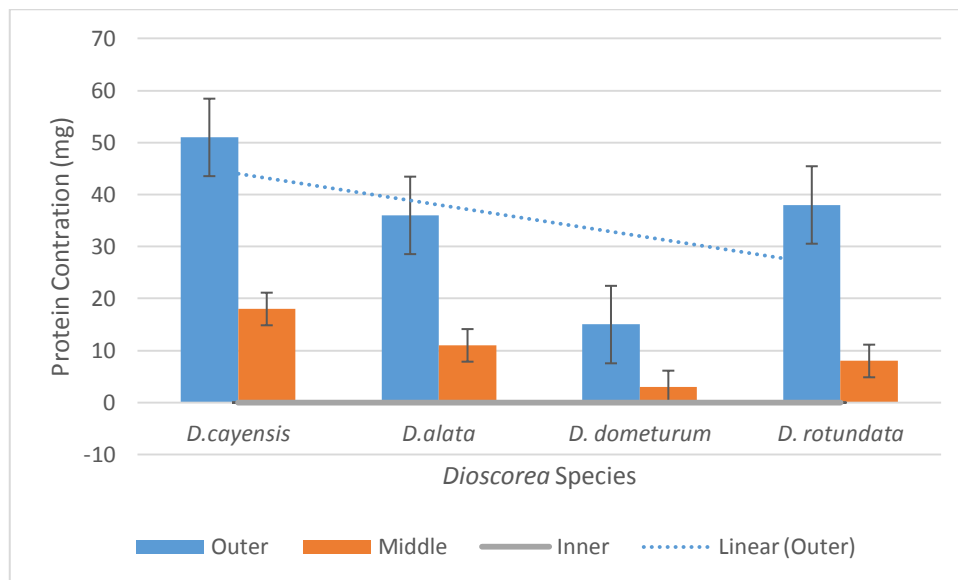


Figure 1., Specific Tuber Layer Nematode load

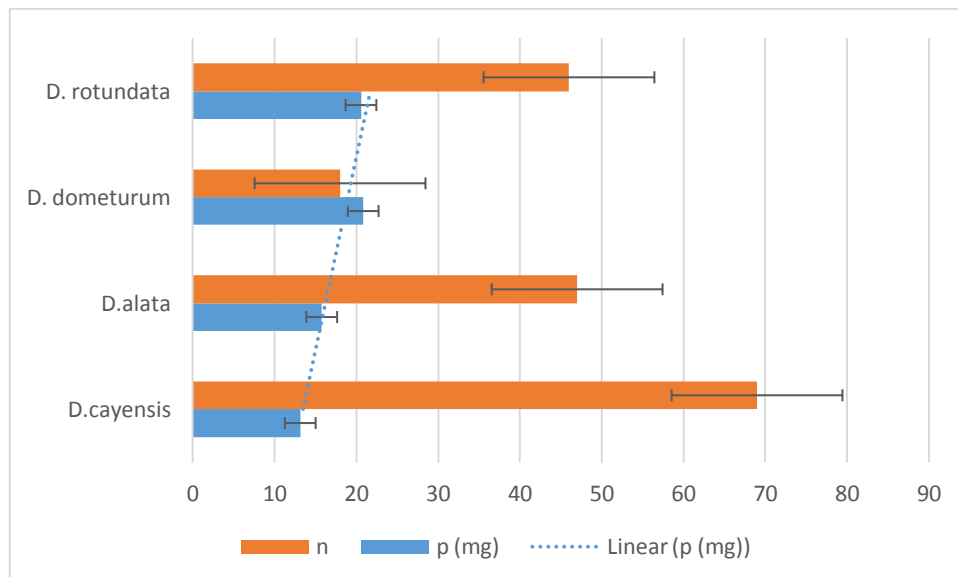


Figure 2. Gross protein and nematode load of yam tubers

Table 5. Comparison between protein content (mg) and layer specific nematode load

Layer of tuber	Species of yam							
	<i>D. cayensis</i>		<i>D. alata</i>		<i>D. dometurum</i>		<i>D. rotundata</i>	
	p (mg)	n	p (mg)	n	p(mg)	n	p(mg)	n
Outer	3.50±.04	51	3.94±.06	36	7.0±00	15	6.56±04	38
Middle	4.38±.02	18	5.69±.01	11	7.7±00	3	7.0±.01	8

Inner	5.25±.05	0	6.13±.07	0	6.13±.07	0	7.0±.00	0
Total (%)	13.13±.03	69	15.76±.04	47	20.83±.03	18	20.56	46

p =protein content of tuber, n=plant parasitic load of tuber



PLATE 1A-C: *Pratylenchus* spp., *Meloidogyne* spp., and *Meloidogyne* spp., adult

3.3. Tuber specific layer nematode load and moisture content

D. cayensis retained more water in the tissues than the other three sampled yam tubers, however, two different patterns of moisture retention was observed the specific tubers layers in relation to the nematode load of the yam tubers. In *D. cayensis* and *D. rotundata*, nematode load decreased as moisture content declined while, in *D. alata* and *D. dometorum* the middle layer of the tubers retained more moisture than the outermost and core (Table 4). The gross moisture content of the tubers indicated *D. dometorum* as being relatively less hydrated than the other yam species. The study also, showed that the outermost layers of the various tubers contained higher nematode load than the middle and core (Figure 4). There was a significant difference between the nematode load and moisture content amongst the yam species and within the specific layers of individual yam tubers ($p > 0.05$).

Table 4. Comparison between Tuber Layer Specific Nematode Load and moisture content

Layer of tuber	Species of Yam							
	<i>D. cayensis</i>		<i>D. alata</i>		<i>D. dometorum</i>		<i>D. rotundata</i>	
	n	m	n	m	n	m	n	m
Outer	51	66.53±.07	36	59.87±.03	15	79.76±.04	38	74.3±.07
Middle	18	62.28±.02	11	65.77±.03	3	80.79±.01	8	63.0±.01
Inner	0	60.25±.01	0	59.96±.04	0	78.22±.08	0	60.1±.03
Total (%)	69	189.06±.10	47	185.6±.03	18	158.77±.06	46	197.4±.04

n = Specific Tuber Layer Nematode Load, m=moisture content of tuber(%)

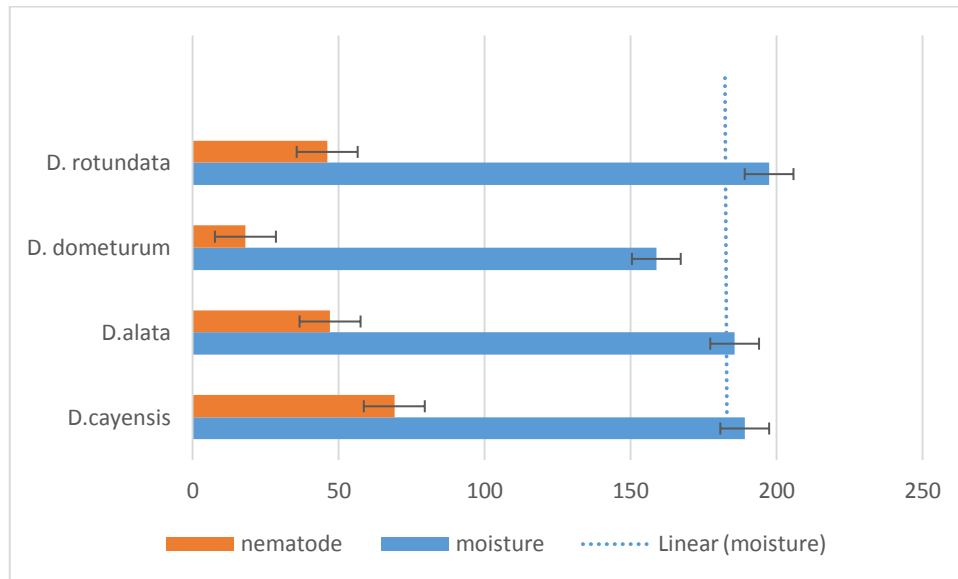


Figure 4. Gross nematode load and moisture content of the yam tubers

2.10. Comparison between tuber layer specific nematode load and carbohydrate content (mg/ml)

The crude carbohydrate composition varied within the specific layers of individual yam tubers and across the varieties of yams. *D. cayensis* and *D. rotundata* expressed drastic reductions in carbohydrate at the outermost layers of the yam which progressively increased as nematode load reduced. This trend was not observed in *D. alata* and *D. dometorum* where tuber carbohydrate content was not drastically depleted at the outermost layer. The carbohydrate integrity of the specific tuber layers showed the influence of nematode parasitism on carbohydrate composition of the tubers (Figure 5 and Table 6). The study revealed that the heavily infected tubers contained relatively lower concentration of carbohydrate than the relatively less infected tubers. There was a significant difference ($p < 0.05$) between the carbohydrate concentration of the specific tuber layers in all the yam varieties but none ($p > 0.05$) between the different *Dioscorea* species.

Table 6. Comparison between Tuber Layer Specific Nematode Load and carbohydrate content (mg/ml)

Layer of tuber	Species of yam							
	<i>D. cayensis</i>		<i>D. alata</i>		<i>D. dometorum</i>		<i>D. rotundata</i>	
	n	c	n	c	n	c	n	c
Outer	51	1.93±.07	36	30.03±.07	15	13.45±.05	38	1.73±.07
Middle	18	27.22±.08	11	25.34±.06	3	13.72±.08	8	27.22±.08
Inner	0	31.35±.04	0	30.30±.01	0	15.25±.05	0	31.35±.05
Total (%)	69	60.5±.00	47	85.67±.04	18	42.42±.05	46	60.3±.05

N=nematode in of yam, c= available carbohydrate content mg/ml)

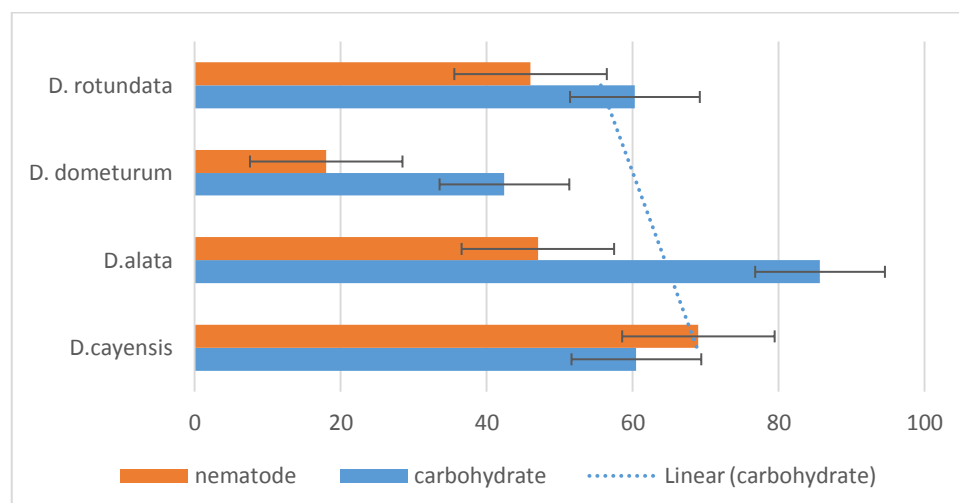


Figure 5., Gross nematode parasitism on total carbohydrate concentration

2.11. Comparison between tuber layer specific nematode load and fibre content

The fibre constitution of the specific layers of the yam tubers varied from the outermost to the innermost layer in all yam species which replicated the recorded trend of endophytic nematode load across the tuber layers, however, the trend was not consistent in all yam varieties. The fibre content of *D. rotundata* highest in comparison to the other sampled varieties. This trend could be due to host specific factors. There was a significant difference ($p < 0.05$) between the tuber specific layer fibre content of individual yam tubers but was not ($p > 0.05$) amongst the yam varieties.

Table 7. Comparison between Tuber Layer Specific Nematode Load and fibre content (mg)

Layer of tuber	Species of yam (%)							
	<i>D. cayensis</i>		<i>D. alata</i>		<i>D. dometorum</i>		<i>D. rotundata</i>	
	n	f	n	f	n	f	n	F
Outer	51	6.78	36	18.95	15	4.13	38	28.87
Middle	18	7.89	11	17.71	3	3.04	8	15.53
Inner	0	9.8	0	15.26	0	3.46	0	8.83
Total (%)	69	24.47	47	51.92	18	10.63	46	53.23

n=nematode in of yam, f= content (mg)

IV. Discussion

4.1 Plant Parasitic Nematodes population in yam tubers

Five species of plant parasitic nematodes; *Meloidogyne* spp., *Scutellonema* spp., *Hoplioliamus* spp., *Pratylenchus* spp., and *Helicotylenchus* spp., were reecovered from the various species of yam tubers used in the study. *Scutellonema* spp., had the highest overall occurrence amongst the yam species followed by *Meloidogyne* spp., and the least was *Helicotylenchus* spp. (Table 1., and figures 1-4). *Scutellonema* spp., was the most abundant and suitable nematode species of the sampled *Dioscorea* species tubers. *D. cayensis* harboured the highest nematode load followed by *D. alata*, while *D. dometorum* had the least infection (Chin *et al.*, 2018). The *Dioscorea* varieties were suitable hosts to some plant parasitic nematodes with varying levels of resistance and tolerance as stated by Abad and Williamson (2010). Nematode load of the specific yam tubers indicated the suitability of *Dioscorea* crop to plant parasitic nematode infectivity which manifested in the endophytic density of the parasites. *D. dometorum* was regarded as resistant to plant parasitic nematodes as it harboured the lowest density of endophytic worms in the study (Table 2) which could be due to its unsuitability to plant parasitic nematodes reproductivity. The relatively low parasite load of *D. dometorum* revealed the resistance of the yam species to plant parasitic nematodes. This attribute of *D. dometorum* could be used in the cultural management of plant parasitic nematodes in subsistence (Table 2, Figures 1-4). The intensity of PPNs decreased from the surface to the core of the tubers. The pattern of endophytic distribution of PPNs in the yam tubers reflected the penetration competence of the parasitic worms.

This study shows that *Dioscorea* spp., of crops maintained progressive parasitism *in vivo* because, field acquired infections remained viable as tubers aged in storage. Field infection of yam tubers implicates it as an important agent of nematode propagation and dispersal in the agroecosystem. Apart from this, nematode cycles are retained in the dormant tubers until planting season comes which confirms that nematodes are suitable to the *Dioscorea* spp., of crop assume quiescence during the dormant stage (storage) of the tubers (Nzeako *et al.*, 2019). The endophytic population of *Scutellonema* spp. was higher than that in the other varieties of *Dioscorea*

spp., because of its physiological characteristics of initiating sustainable feeding sites that are sufficient for the completion of PPNs lifecycle. Plants species usually respond to signal from nematodes to initiate and maintain feeding sites sufficient for completion of nematode lifecycles. *Diocorea* spp. tubers displayed relatively little or no manifestation of symptoms due to their high tolerance ability as demonstrated in *D. cayensis* and *D. rotundata* (Cook and Evans, 1987; Trudgill, 1991). Nemaode parastism of plant tissue generally induces the infected cells to develop proliferative nurturing cells; characterized by granulated cytoplasm with hypertrophied nuclei and nucleoli (Watsonn and Shorthouse, 1979). Several studies also, stated that plant parasitic nematode induced morphogenesis in plant tissues increases as nematode load increases (Watsonn and Shorthouse, 1979, Vovlas *et al.*, 2015, Palomares-Rius *et al.*, 2017). The ability to harbour high densities of parasitic nematodes in yam tubers during ststorage makes the use of local plantings materials (yam tubers) a high risk practice because they harbour high density of endophytic nematodes that are consequently transferred to the field during planting. This hinders any effort by farmers at controlling plant parasitic nematodes of yam using locally available seed yams.

4.2 Specific Tuber Layer Nematode Load and moisture content

Yam tuber loses water in storage resulting in the usual loss of weight and size of stored tubers. The loss of weight of tubers usually affects endogenous obligate nematodes. In this study the moisture content of the different layers of the yam (Specific tuber layers, STL) varied, however, two patterns of moisture dynamism was observed amongst the tuber layers. The pattern observed in *D. cayensis* and *D. rotundata* showed that the nematode load decreased as moisture content declined (Table 4 and Figure 4). In this pattern, there was a progressive top-bottom decline in moisture content which was also, replicated in endogenous nematode density of the STL. The second pattern of moisture dynamics manifested by *D. alata* and *D. dometorum* showed middle-tuber layer high moisture concentration which was not replicated in the nematode load pattern (Table 4.). Curtis *et al.* (2007) implicated plant parasitic nematodes invasion and endophytic migration in the alteration of the root architecture, significant reduction in nutrient uptake and crop yield. In this study, the tubers sampled manifested dormancy (Cook and Evans, 1987; Ogaraku and Usman, 2008). Consequently, the field acquired PPNs infections may have induced increased metabolism that prompted an oxidative boost (FAO, 2009) in the tubers. The study opines that the conventional loss of water by the yam tubers on storage affected the endophytic migration of PPNs species. However, the none-penetration of nematodes into the core of the tubers in all the yam varieties indicated the direction of moisture flow which was opposite to the direction of the migratory worms and migratory competence of the nematode juveniles. The conventional outward flow of moisture from the core to the periphery of the yam tubers influenced the concentration of PPNs at the peripheral layers of the tubers and the subsequent trapping of nematodes along their migratory paths in the tissues of the yam tubers.

4.3 Specific Tuber layer protein content

Mean crude protein concentration of the tubers in relation to tuber specific layers and nematode load respectively revealed a decrease in protein concentration as plant parasitic nematode load increased. This nematode density pattern was consistent in all the yam species except in the *D. dometorum*; where the outermost layer contained more crude protein than the inner layers. However, *D. dometorum* had the highest crude protein content amongst the sampled *Dioscorea* tubers followed by *D. rotundata*, *D. alata* and *D. cayensis* which partially reflected the PPNs status of the different the yam tubers. The relatively high PPNs load of *D. cayensis* and *D. alata* may be responsible for the lower crude protein content recorded in the yam species (Table, 5). This result suggests that nematodiasis of the *Diocorea* tubers influenced the protein concentration of the specific tuber layers where feeding sites were established (Plowright, and Kwoseh, 2000). The establishment of feeding sites by nematodes in plant tissues causes the release of several stylet borne proteinous secretions (Mitchum *et al.*, 2013, Jones, 1981, Williamsom and Kumar, 2006, Goellner *et al.*, 2001, Sobezak *et al.*, 2011). A lot of these proteins degenerate the tissue integrity of the plant host in a bid to manipulate the metabolism of the the infected cells especially, amongst the sedentary nematodes species (Barcala *et al.*, 2010, Damiani *et al.*, 2012, Williamson and Kumar, 2006, and Kaloshian *et al.*, 2011). The protein profile of the infected tissues varied from the uninfected sites in the study, due to drastic cellular changes associated with nematodiasis of the tubers (Gheysen and Fenoll, 2002). According to Mitchum *et al.* (2013), some of the cellular changes leading to establishment of feeding sites in plant hosts include; alterations to the nuclei, cytoskeleton, hormone status and metabolism of the selected cell(s), and extensive changes in cell wall architecture. These alterations are more prevalent in susceptible host's endophytic environments than in resistant ones (Gheysen and Fenoll, 2002; Hammes *et al.*, 2005, Jammes *et al.*, 2005; Ithal *et al.*, 2007, Szakastit *et al.*, 2009, Barcala *et al.*, 2010, Hofmann *et al.*, 2010). Considering the array of activities that accompany establishment of feeding sites by PPNs, Mitchum *et al.* (2013) opined that PPNs effectors should be used as molecular probes to understanding plant cell cycle, cell wall architecture and cellular metabolism, among other processes which this study agrees with. As insightful as this opinion is, the protein status of *D. dometorum* in this study appeared to be relatively

resistant to PPNs infection due to the host specific factors of the yam species accentuated in the progressive increase in the crude protein content of the tuber as nematode load diminished.

4.4 Tuber Layer Specific Nematode Load and Carbohydrate Content (mg/ml)

Carbohydrate composition varied within the specific layers of the yam tubers and across the varieties of yams, however, *D. cayensis* and *D. rotundata* manifested a top-down reduction in carbohydrate concentration (Table 6 and Figure 5). This trend was not observed in *D. alata* and *D. dometorum* where tuber carbohydrate concentrations showed relatively no top-down depletion. Nematode parasitism was implicated in the general alteration of plant tissues (Hammes *et al.*, 2005, Ithal *et al.*, 2007, Szakastit *et al.*, 2009). Such as degeneration of the carbohydrate integrity of the specific layers of tubers. This trend was not obvious in *D. dometorum* which could be attributed to the resistant nature of the yam species to PPNs build up in the tuber. The result shows that carbohydrate depreciation in the yam tubers was associated with high PPNs load.

4.5 Tuber Layer Specific Nematode Load and fibre content (g)

The fibre constitution of the specific layers of the yam tubers varied from the outermost to the innermost layer in all yam species. This trend was also replicated in the endophytic nematode load of the layers. The fibre composition of the yam tubers showed variable responses to nematode attack which could be due to host specific factors of the yam varieties.

V. Conclusion

Parasitism by PPNs on *Dioscorea* crops was influenced by both environmental and host specific factors in the study. PPNs infection on yam tuber was depth related because the overall PPNs load was concentrated at the peripheral regions of the tubers that are closest to the soil environment. The penetration and migratory competence of the nematodes were host specific and only aggressively mobile species such as *Pratylenchus* and *Scutellonema* were found within the deeper layers of the tubers. Total PPNs load of the *Dioscorea* spp. tubers did not contribute significantly ($P < 0.05$) to the overall crude protein status of the tubers. However, PPNs infection of the specific layers of the tubers influenced the decline in the crude protein content of the tubers significantly ($P < 0.05$) in relation to the uninfected portions of the tubers which was also, observed in the carbohydrate concentration. PPNs infection influence on the fibre integrity of the tubers was tuber depth related. Moisture content of the stored tubers influenced the mobility of nematode juveniles in the tubers. However, *D. dometorum* was relatively resistant to PPNs while *D. cayensis*, *D. alata* and *D. rotundata* were suitable to PPNs infection. The study opines that tuber resident nematodes were trapped in motion as tubers lost moisture conventionally. The study opines that the long shelf life of *Dioscorea* spp. tubers makes them reservoir hosts of numerous PPNs in the soil thereby making the *Dioscorea* species of crops reliable agents of dispersal of PPNs in the agroecosystem. Pre-planting sterilization of tubers is recommended to curb the transmission and dispersal of PPNs in the agroecosystem.

Conflict of Interest

The research hereby states that there is no conflict of interests in relation to any sections of this study by the authors.

Acknowledgement

Mr Barisuka Aboloboa of the Department of Animal and Environmental Biology who helped out in the nematological techniques and Mr Japhet Onwuegwu of the Department of Plant and Biotechnology for supervising the Biochemical Analysis are greatly appreciated.

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Caroline Kathure Gatobu, et. al. “The Influence of Smallholder Farming Characteristics on Household Food Security in West Pokot County, Kenya.” *IOSR Journal of Agriculture and Veterinary Science (IOSR-JAVS)*, 14(3), 2021, pp. 16-26.