# A Review Pesticidal Activity of Essential Oils against Sitophilus oryzae, Sitophilus granaries and Sitophilus Zeamais

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## Abstract

Infestation of cereals in storage by Sitophilus oryzae, Sitophilus granaries and Sitophilus Zeamais lead to great food loses in Sub-Saharan Africa. Some of the synthetic pesticides which are available are non-biodegradable and have adverse effect on the environment and humans. Previous studies show that plant extracts have the potential for use as pesticidal agents against the weevils. The use of plant extracts in pest management is preferred because they are readily available and chances of drug resistance are low. The aim of this study was to collate and review the fragmented information on essential oils from plants with pesticidal activity against S. zeamais, S. oryzae and S. granaries and present recommendations for future research. Peer-reviewed articles were retrieved from Scopus, Science Direct, SciFinder and Google Scholar. This study led to identification of 196 essential oils extracted form plant species belonging to 31 plant families. Essential oils from the Lamiaceae family are the most studied, followed by Myrtaceae, Asteraceae, Apiaceae, Lauraceae and Piperaceae families. Insecticidal activity studies of the essential oils were mostly tested against S. zeamais (115 essential oils) followed by S. oryzae (58 essential oils) and S. granaries (23 essential oils) which suggests that S. zeamais is the most rampant and the most dreaded species. Future studies aimed in isolating and characterizing the active compounds from the essential oil is necessary. It is also necessary to develop effective formulations for controlling the pests.

Keywords: Storage pest; Sitophilus; Essential oil; Toxicity

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# I. Introduction

Cereals are the most important source of food all over the world and approximately 75% of calories and 67% of protein consumed worldwide are provided by cereals. They provide 10000-15000kJ/Kg of energy, which is approximately 15-20 times more than fruits and vegetables. Cereals are also an important source of many essential vitamins, minerals and phytochemicals. The grains are easy to carry, package and transport from one place to another, and are used as an ingredient in many food products such as porridge, breakfast cereals, bread and cereal based beverages. Cereals can grow in adverse environmental and bad soil conditions, and the yield is not compromised due to harsh environmental conditions. The crops give the highest yield when compared to most other crops. However, a large proportion of cereals go to waste because of destruction by insect pest.

*Sitophilus* is a genus of weevils in the tribe Litosomini. As of 1993, there are about 14 species of *Sitophilus*<sup>1</sup>. Maize weevil (*Sitophilus zeamais* Motchu.), rice weevil (*Sitophilus oryzae* L.) and granary weevil (*Sitophilus granaries* L.) are among the most important pests that destroy cereals in storage. The pests affect stored grains including wheat, rice, sorghum, oats, barley, rye, buckwheat, peas and cottonseed. Both adults and larvae of the insects feed on the grains and infestation can start in the field but most damage occurs in storage. The extensive tunneling allows the pests to convert the harvest into flour within a very short time<sup>2</sup>. In addition, infestations of the insect result in development of *Aspergillus* which is carcinogic<sup>3-5</sup>. In most cases, small-scale farmers are forced to sell their produce immediately after harvest, thereby attract low prices and compromising food security. Synthetic insecticides are commercially available and the chemical method is the most effective. However, indiscriminate use of synthetic insecticides results pest resistance and undesirable effects on non-target organisms, human and environmental hazards<sup>6, 7</sup>. This call for a need to search for safe alternatives that is environmentally friendly.

Plants have been identified as a source of important metabolites that are used in defense against different pests and pathogenic microorganism<sup>8-20</sup>. Search for insecticidal compound from plants through *in-vivo* 

and *in-vitro* experiments has yielded important compounds including alkaloids, terpenoids, flavonoids, steroids and quinones<sup>21-27</sup>. Such compounds represent an important source of drugs in the process of developing new pharmacologically active compounds. The use of botanical for pests and disease control is preferred because they are environmentally friendly and non-toxic to non-targeted organisms<sup>4, 5, 28-38</sup>. In addition, chances of pests and pathogens developing resistance to botanical pesticides are highly unlikely. This paper provides a review on pesticidal activity of plant extracts with emphasis on plant essential oils exhibiting toxicity, repellent, antifeedant, oviposition deterrent and growth inhibition activities against *Sitophilus oryzae* (L.), *Sitophilus granaries* (L.) and *Sitophilus Zeamais* (Motsch.).

## II. Essential Oils in control of *Sitophilus species*

Essential oils from plants are considered to be an alternative means of controlling many harmful insects. Recent research has demonstrated their toxicity, antifeedant, adult emergence and oviposition inhibition and repellent activity against *Sitophilus species*<sup>23, 24, 39, 40</sup>. Although a number of review articles have appeared in the past on the various aspects of essential oils bioactivities<sup>27, 41, 42</sup>, the present paper emphasizes use of plant oils in control of *S. oryzae, S. granaries* and *S. zeamais* which are the most important storage pests in the genus *Sitophilus*.

Information access from published research in the last 15 years indicate that 196 essential oils from plants belonging to 31 different plant families were studied for their insecticidal activity against the three *Sitophilus* species weevils (Figure1). Essential oils from plants belonging to the Lamiaceae family are the most studied (50) followed by Myrtaceae (19), Asteraceae (17), Apiaceae and Lauraceae (16), Piperaceae (15), Rutaceae (13), Poaceae (8), Pinaceae, Schisandracea and Verbenaceae (4), Geraniaceae, Hypericaceae and Zingiberaceae (3), Anacardiaceae, Annonaceae, Atherospermataceae and Labiate (2), Alliaceae, Burseraceae, Canellaceae, Cupressaceae, Ericaceae, Fabaceae, Myristicaceae, Pedaliaceae, Ranunculaceae, Santalaceae, Simmondsiaceae and Solanaceae (1). Insecticidal activity against *S. zeamais* is the most studies (115 essential oils) followed by *S. oryzae* (58 essential oils) and *S. granariu* (23 essential oils) as shown in Figure 2. This probably indicates that *S. zeamais* is the most common of the three among the *Sitophilus* species. Most of essential oils studied against *S. zeamais* belong to the following families: Lamiaceae (12), Piperaceae (14), Lauraceae (3). Most of essential oils studied against *S. oryzae* belong to the following families: Lamiaceae, Poaceae and Verbenaceae (7), Poaceae and Rutaceae (4), and Lauraceae (3). For *S. granaries*, most essential oils studies (11) belong to Lamiaceae family.



Figure no 1: Distribution of plants species studied per family



Figure no 2: Distribution of number of plant essential oils by family per insect pest

# III. Activity of Essential Oils Against Sitophilus oryzae

From the information retrieved from published data, 58 essential oils were investigated against Sitophilus oryzae (Table 1). Essential oil from C. citratus, C. nardus, C. zeylanicum and A. calcarata gave LC<sub>50</sub> values of 35, 82, 70 and 367 mg/L respectively in fumigation test and 11.5, 18.7, 3.6 and 40µg/cm<sup>2</sup> respectively in contact toxicity test<sup>43</sup>. Carum copticum gave LC<sub>50</sub> value of 0.91µL/L<sup>44</sup> while Eucalyptus spp. had LC<sub>50</sub> values between 7-8.5  $\mu$ L/L <sup>45, 46</sup>. In the contact toxicity test, LC<sub>50</sub> values of 0.14, 0.08, 0.61, 0.42, 0.01, 0.22, 0.10 and 0.11mg/cm<sup>2</sup> for Achillea santolina L., Artemisia judaica L., Citrus reticulata Balanco, Schinus terebenthifolius Raddi, Mentha microphylla C. Koch., Lantana camara L., Majorana hortensis Moench and Eucalyptus camaldulensis Dehnh, respectively were recorded<sup>47</sup>. In the fumigant toxicity assay, LC<sub>50</sub> values recorded were 79.63, 100, 58.62, 56.48, 0.21, 29.47, 100 and 30.81 for *A. santolina A. judaica*, *C. reticulata*, *S. terebenthifolius*, *M. microphylla*, *L. camara*, *M. hortensis* and *E. camaldulensis*, respectively<sup>47</sup>. Acorus calamus gave  $LD_{50}$  value of 54.46µg/cm<sup>2</sup> against the weevil<sup>48</sup>. Origanum vulgare, Salvia fruticosa, S. officinalis, S. *pomifera, Thymbra capitata* and *Thymbra persicus* showed high fumigation toxicity toward *S. oryzae*, with  $LC_{50}$  values ranging between 1.5 and  $9\mu L/L^{49-51}$ . Application of *Mentha viridis* essential oil gave  $LC_{50}$  values of 239 and 158ppm after one and two weeks duration of exposure respectively<sup>52</sup>. Syzygium aromaticum and Cinnamomum zeylanicum gave 84 and 70% mortalities respectively at 4.0µl /50 ml air while Cinnamomum zeylanicum and Melaleuca alternifolia gave 90 % mortality at concentrations of 8.0 and 16.0µl /50 ml air respectively 24 hour after treatment<sup>53</sup>. Syzygium aromaticum gave LC<sub>50</sub> values of 17.3 and 15.3µl after 24 and 48h exposure duration respectively<sup>54</sup>. At a concentration of  $130\mu$ g/cm<sup>2</sup>, Coriandrum sativum and Eucalyptus obliqua oils caused 100% mortality of the insects 24h after treatment, whereas Pinus longifolia oil gave 90% mortality after 72h in the fumigation test. Coriandrum sativum Eucalyptus obliqua and Pinus longifolia oils gave 100, 94 and 80% mortality 72 in the contact toxicity assay. The oils gave  $LD_{50}$  values of 36.68, 52.77 and 77.30µg/cm<sup>2</sup> for C. sativum, E. obliqua and P. longifolia respectively<sup>55</sup>. Essential oil from Aegle marmelos was toxic to the weevil with  $LC_{50}$  values of 18.488 and 16.133 µl at 24 and 48h after treatment respectively<sup>54</sup>.

The LC<sub>50</sub> of contact and residue toxicities of essential oil from *Cymbopogon nardus* were 23.2% and 1.71% respectively against *S. oryzae*<sup>56</sup>. *Artemisia judaica, Callistemon viminalis and Origanum vulgare* gave LD<sub>50</sub> values of 0.08, 0.09, and 0.11 mg/cm<sup>2</sup> respectively while *Syzygium aromaticum* and *Lavandula officinalis* oils gave LD<sub>50</sub> values of 0.04 and 0.07mg/cm<sup>2</sup> respectively<sup>49,57</sup>. Fumigation of *S. oryzae* adults with *A. sativum* essential oil gave LC<sub>50</sub> values of 0.30 and 0.24µl/cm<sup>3</sup>air 24 and 48h after treatment respectively while LC<sub>50</sub> values of 0.17 and 0.13µl/cm<sup>2</sup> after 24 and 48h respectively in contact toxicity tests. Fumigation with *C. tamala* oil gave LC<sub>50</sub> values of 0.249 and 0.198µl/cm<sup>3</sup>air after 24 and 48h respectively while in contact toxicity test, LC<sub>50</sub> values of 0.241 and 0.218µL/cm<sup>2</sup> were observed after 24 and 48h respectively<sup>58,59</sup>. In a fumigant bioassay, *H. officinalis* had the highest toxicity against *S. oryzae* adults, followed by *R. officinalis* and *M. piperita* with LC<sub>50</sub> values of 78.16, 115.63 and 299.51µL/L air respectively<sup>60</sup>. Fumigation tests were performed to study toxicity of garlic, chili pepper, cardamom, peppermint, eucalyptus and tea tree essential oils on rice weevil at concentrations of 0.4, 0.6 and 0.8ml on 5cm diameter filter paper. Percentage mortalities were found to be in the order: peppermint (75.8%) > Tea tree (72.3%) > Garlic (65.5%) >Chili Pepper (62%) > Camphor (55.1%) > Cardamom oil (51.7%) <sup>61</sup>. The LD<sub>50</sub> values for aniseed, camphor, citronella, eucalyptus, geranium, lavender, lemon, rosemary, vetiver and wintergreen essential oils were found to be 96.53, 49.92, 67.42, 47.42, 67.29, 154.03, 40.38, 58.34, 109.36 and 59.57 respectively while LD<sub>90</sub> values were 178.38, 86.78, 110.29, 89.92,

103.47, 241.53, 83.12, 107.81, 162.11 and 115.95 respectively at 72 hours. *Laurus* nobilis oil gave LC<sub>50</sub> value of 8.0µL/L against the weevil<sup>50, 62</sup>. Essential oils of *Coriandrum sativum* seeds gave LC<sub>50</sub> value of 107.56 and 108.40µg/cm<sup>2</sup> against *S. oryzae* and *S. zeamais* respectively contact toxicity test while in fumigant toxicity test, LC<sub>50</sub> values of 79.88 and 80.29µg/cm<sup>2</sup> were recorded for *S. oryzae* and *S. zeamais* respectively<sup>63</sup>. Essential oil from *Eucalyptus globules*, *Cymbopogon citrates* and *Citrus maxima* caused 73.33, 81.67 and 65.0% mortality of adult weevil at 1% concentration, 21 days after treatment <sup>64</sup>. *Contact toxicity of Origanum majorana* oil against *S. oryzae* was 80.0 and 90.0% on 3<sup>rd</sup> and 7<sup>th</sup> day respectively after treatment with 4.0% w/w oil. The oil gave LC<sub>50</sub> values of 0.646 and 0.533 % w/w at 3 and 7 days after treatment respectively. In fumigant toxicity test, while the essential oil gave LC<sub>50</sub> of 130.1 and 72.8µL/L air at 3 and 7 days after treatment<sup>65</sup>. In a contact toxicity assay *Nepeta glomerulosa* (LC<sub>50</sub> = 124.318µL/L air) showed the highest toxicity against *S. oryzae* followed by *N. pogonosperma* (LC<sub>50</sub> =150.49µL/L air), *N. cataria* (LC<sub>50</sub> =152.630µL/L air) and *N. binaloudensis* essential oil (LC<sub>50</sub> = 366.80µL/L<sup>66</sup>. The LC<sub>50</sub> values for *T. copicum*, *L. camara*, *C. nardus and C. zeylanicum* were 07.24, 08.72, 10.63 and 06.19µL/L respectively at 72h after treatment<sup>67</sup>. In contact insecticidal test of *L. taraxacifolia* essential oil, LC<sub>50</sub> values ranged between 54.38 -10.10µL/mL between 24-120 h after treatment<sup>68</sup>.

Repellency of Allium sativum oil against S. oryzae was found to be 33.33, 54.16, 75.0 and 95.83% at concentrations of 0.2, 0.4, 0.8 and 1.6% respectively<sup>58</sup>. Repellency of *Cinnamonum tamala* essential oil was found to be 48.33, 75.83, 88.33, 97.50 and 100% at concentrations of 0.2, 0.4, 0.8, 1.6 and 3.2% oil respectively<sup>59</sup>. Repellency aniseed, camphor, citronella, eucalyptus, geranium, lavender, lemon, rosemary, vetiver and wintergreen essential oils were tested at concentrations of 10 and 50µL against adults of Sitophilus  $oryzae^{62}$ . The order of repellency of the plant oils at 10µL on 6 hours of exposure with EPI was: camphor (-0.90), wintergreen (-0.88), lavender (-0.70), citronella (-0.70), rosemary (-0.67), vetiver (-0.62), lemon (-0.57), eucalyptus (-0.55), geranium (-0.44) and aniseed (-0.04). At 50µL the order of repellency was camphor (-1.0), wintergreen (-0.89), citronella (-0.89), lemon (-0.89), lavender (-0.71), vetiver (-0.69), geranium (-0.65), rosemary (-0.57), eucalyptus (-0.52) and aniseed (-0.50)<sup>62</sup>. Sitophilus oryzae was repelled by M. piperita (95.0 %), R. officinalis (91.0 %) and H. officinalis (86.5 %) at oil concentration of 16  $\mu$ L/30 cm<sup>2</sup> while Repellent effect of marjoram oil ranged between 90-100% at a concentration of 4.0 % w/w within 72 hours after treatment <sup>60, 65</sup>. The order of repellency was of four plant oils tested against the weevil was: N. cataria (100%), N. pogonosperma (96.67%), N. glomerulosa (70%), and N. binaloudensis (100%), at 25µL/30 cm<sup>2</sup> after 5h<sup>66</sup>. In antifeedant assay, A. sativum essential oil significantly decreased consumption of flour disk by S. oryzae adults. Consumption of flour disk was reduced to 81.72, 50.91, 33.16 and 13.73 % of control when treated with A. sativum essential oil at concentrations of 3, 6, 9 and 12 µl/disk respectively<sup>58</sup>. Consumption of flour disk was reduced to 80.27, 46.59, 20.16 and 6.64% of control when treated with 3, 6, 9 and 12 µl/disk of C. tamala essential oil<sup>59</sup>. Fumigation of S. oryzae adults with A. sativum essential oil significantly reduced oviposition potential. Reduction in oviposition was 49.45 and 15.78% of the control when the weevil was fumigated with 40 and 80% of 24-h LC<sub>50</sub> respectively, 24h after treatment<sup>58</sup>. Application of *Mentha viridis* essential oil decreased the number of emerged S. oryzae adults by 58.7, 79.6 and 86.0% at concentrations of 100, 200 and 300 ppm respectively<sup>52</sup>. Essential oil of Origanum majorana caused 81.2% reduction of F1 progeny at 4.0 % w/w dosage <sup>65</sup>. Essential oil from *Eucalyptus globules*, *Cymbopogon citrates* and *Citrus maxima* gave 72.22, 77.78 and 61.11% reduction in progeny emergence at a concentration of 1% 50 days after treatment<sup>6</sup>

Table no 1: Plants species tested against Sitophilus oryzae

Family	Botanical name	Common name	Bioactivity	Reference
Alliaceae	Allium sativum	Garlic	Toxicity, repellency, feeding and oviposition inhibitory	58, 61
Anacardiaceae	Schinus terebenthifolius	Peppertree	Toxicity	47
Apiaceae	Carum copticum	Ajwain	Toxicity	44
Apiaceae	Coriandrum sativum L.	Coriander	Toxicity	55, 63, 69
Apiaceae	Cuminum cyminum	Cumin	Toxicity	70
Apiaceae	Pimpinella anisum	Aniseed	Toxicity and repellency	62
Apiaceae	Trachyspermmum copicum	Ajwain	Toxicity	67
Araceae	Acorus calamus	Sweet flag	Toxicity	48
Asteraceae	Achillea santolina	Santoline	Toxicity	47
Asteraceae	Artemisia judaica	Wormwood	Toxicity	47, 49
Asteraceae	Launaea taraxacifolia (Willd.)	Lettuce	Toxicity	68
Ericaceae	Gaultheria fragrantissima	Wintergreen	Toxicity and repellency	62
Geraniaceae	Pelargonium graveolens	Geranium	Toxicity and repellency	62
Labiatae	Majorana hortensis	Majorana	Toxicity	47
Labiatae	Rosmarinus officinalis L.	Rosemary	Toxicity and repellency	60, 62, 71
Lamiaceae	Hyssopus officinalis L.	Hyssop	Toxicity and repellency	60
Lamiaceae	Lavandula officinalis	Lavender	Toxicity and repellency	57, 62
Lamiaceae	Mentha microphylla	Spearmint	Toxicity	47
Lamiaceae	Mentha piperita L.	Peppermint	Toxicity and repellency	60

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Lamiaceae	Mentha pulegium	Pennyroyal	Toxicity and, repellency	61
Lamiaceae	Mentha viridis	Pudding grass	Toxicity, growth inhibition	52
Lamiaceae	Nepeta binaloudensis		Toxicity and repellency	66
Lamiaceae	Nepeta cataria	Catnip	Toxicity and repellency	66
Lamiaceae	Nepeta glomerulosa Boiss.		Toxicity and repellency	66
Lamiaceae	Nepeta pogonosperma		Toxicity and repellency	66
Lamiaceae	Origanum majorana	Sweet majorana	Toxicity, growth inhibition and	65
			repellency	
Lamiaceae	Origanum vulgare	Wild majorana	Toxicity	49, 50
Lamiaceae	Salvia fruticosa	Greek sage	Toxicity	49, 50
Lamiaceae	Salvia officinalis L.	Sage	Toxicity	49, 50
Lamiaceae	Salvia pomifera	Apple sage	Toxicity	49, 50
Lamiaceae	Thymbra capitata	Cone head thyme	Toxicity	49, 50
Lamiaceae	Thymbra persicus	Persian thyme	Toxicity	49, 50
Lamiaceae	Thymus vulgaris	Thyme	Toxicity	53
Lauraceae	Cinnamomum tamala	Cassia	Repellent, feeding and oviposition	59
			inhibition	
Lauraceae	Cinnamomum zeylanicum	Cinnamon	Toxicity, repellence	42,53, 62, 67
Lauraceae	Laurus nobilis	Bay tree	Toxicity	50
Myrtaceae	Callistemon viminalis	Weeping bottle-	Toxicity	49
		brush	-	
Myrtaceae	Eucalyptus camaldulensis	River red gum	Toxicity	47
Myrtaceae	Eucalyptus globules	Eucalyptus	Toxicity and emergence inhibition	53, 61,62,64
Myrtaceae	Eucalyptus obliqua L'Her	Stringybark	Toxicity	55
Myrtaceae	Eucalyptus spp		Toxicity	45,46
Myrtaceae	Melaleuca alternifolia	Tea tree	Toxicity	53, 61
Myrtaceae	Syzygium aromaticum	Cloves	Toxicity	53, 54, 57
Pinaceae	Pinus longifolia L.	Pine	Toxicity	55
Piperaceae	Piper nigrum	Black pepper	Toxicity	70
Poaceae	Cymbopogon citratus	Lemon grass	Toxicity and emergence inhibition	43, 64
Poaceae	Cymbopogon flexuosus	Cochin grass	Toxicity	53
Poaceae	Cymbopogon nardus	Citronella	Toxicity and repellency	43, 56, 62, 67
Poaceae	Vetiveria zizanioides	Vetiver	Toxicity and repellency	62
Rutaceae	Aegle marmelos	Golden apple	Toxicity	54
Rutaceae	Citrus aurantium	Lemon	Toxicity and repellency	62
Rutaceae	Citrus maxima	Citrus, pummelo	Toxicity, grain damage, weight	64
			loss and progeny emergence	
Rutaceae	Citrus reticulata	Tangerine	Toxicity	47
Simmondsiaceae	Simmondsia chinensis	Jojoba	Toxicity	53
Solanaceae	Capsicum annuum	Chili Pepper	Toxicity and repellency	61
Verbenaceae	Lantana camara	Lantana	Toxicity	47,67
Zingiberaceae	Alpinia calcarata	Snap ginger	Toxicity, repellence	43
Zingiberaceae	Elettaria cardamomum	True Cardamom	Toxicity and repellency	61

# IV. Activity Of Essential Oils Against Sitophilus Zeamais

Information retrieved from published data indicates that 145 essential oils were investigated against Sitophilus zeamais (Table 2). Essential oil of Eucalyptus saligna gave LD<sub>50</sub> value of 0.36µL/cm<sup>2</sup> in contact toxicity on filter paper disc and LD<sub>50</sub> value of 38.05µL/40g of maize grain<sup>72</sup>. Toxicity effect of essential oils from Tagetes minuta L., Mentha longifolia L., Rosmarinus officinalis L., Helichrysum odoratissimum L. and Pelargonium graveolens L. were investigated against maize weevil, S. zeamais. In contact test, T. minuta and M. *longifolia* oils caused 100% mortality within two days after treatment at concentrations of 0.375 and 0.50 $\mu$ L/g of grain respectively<sup>73</sup>. A mortality rate of up to 100% of adult S. zeamais was reported when the insects were fumigated with Tagetes patula oil<sup>74</sup>. Essential oils of Artemisia lavandulaefolia and A. sieversiana gave  $LC_{50}$ values of 11.2 and 15.0mg/ respectively)<sup>75</sup>. LD<sub>50</sub> values of 0.20, 0.21, 0.26, 0.40, 0.29 and  $0.22\mu$ L/cm<sup>2</sup> were reported for O. basilicum, O. americanum, O. tenuiflorum, C. hystrix, C. aurantifolia and E. caryophyllus respectively, on toxicity assay on filter paper. However, the LD<sub>50</sub> values were 0.43, 1.58, 2.33, 1.92, 2.29 and 1.91µL/cm<sup>2</sup> for O. basilicum, O. americanum, O. tenuiflorum, C. hystrix, C. aurantifolia and E. caryophyllus on rice grains respectively<sup>76</sup>. Litsea salicifolia gave LD<sub>50</sub> value of 0.079µL/insect and LC<sub>50</sub> value of 4.4µL/L air, in contact and fumigant toxicity assays respectively<sup>77</sup>. *Illicium fragesii* and *I. simonsii* gave LC<sub>50</sub> value of 11.36 and 14.95mg/L respectively while *Ostericum sieboldii* (LC<sub>50</sub> = 20.92 mg/L)<sup>78-80</sup>. Essential oil extract of *Ocotea odorifera* gave 86% mortality of the insects at a concentration of  $0.32\mu$ Lcm<sup>-2</sup> 24h after treatment with LD<sub>50</sub> value of  $0.09\mu$ l/cm<sup>2</sup> (81) while *Aster ageratoides* oil gave LD<sub>50</sub> value of 27.16 $\mu$ gcm<sup>-2 81, 82</sup>. *Blumea balsamifera* essential oil gave LC<sub>50</sub> of 10.71mg/L air against S. zeamais adults in fumigation toxicity test. The insecticidal compound from the oil were identified to be 1,8-cineole, 4-terpeneol and  $\alpha$ -terpeneol with LC<sub>50</sub> values of 2.96,4.79 and 7.45mg/L air respectively<sup>83</sup>. Ostericum grosseserratum oil exhibited contact and fumigant toxicity against adult S. zeamais with LC<sub>50</sub> value of 17.97 µg/adult and LC<sub>50</sub> value of 13.70mg/L air <sup>84</sup>.

Essential oil from Laurelia sempervirens caused 100% mortality at a concentration of 10 ml/kg grain in the contact toxicity test while it gave 72% mortality at 175 µL/L air<sup>85</sup>. Warburgia ugandensis essential oil caused 100% mortality of the weevil at 0.2 ml/ 20g of grains at 21 days after treatment<sup>39</sup>. Essential oil of D. carota, C. atlantica, C. zeylanicum, C. semperbirens, L. angustifolia, P. cablin, M. pulegium and M. alternifolia caused 26.67, 8.33, 10.00, 8.33, 25.0, 38.33, 96.67 and 63.33% mortality of S. zeamais at 24h after treatment at a concentration of  $10\mu$ L/L air in fumigant toxicity test. The LD<sub>50</sub> values for *M. pulegium* and *M. alternifolia* were 1.97 and 8.80 $\mu$ L/L respectively<sup>86</sup>. Essential oil of *M.* alternifolia also caused 82.22% mortality at a concentration of 11.97 mg/L air 24h after treatment. The LC<sub>50</sub> values were 8.42, 7.70, and 6.78mg/L air, after 24, 48, and 72h of oil treatment respectively. The most potent compounds in the oil were terpinen-4-ol and  $\alpha$ terpineol with a LC<sub>50</sub> value of 3.12 and 5.87mg/L air, respectively<sup>87</sup>. Essential oils from Aphyllocladus decussatus Hieron, Aloysia polystachya Griseb, Minthostachys verticillata Griseb Epling and Tagetes minuta L , were tested for toxicity activity. The percentage mortalities were 25, 32, 36, 82 and 66 at a concentration of 150 (µl/L air) for A. decussates, A. polystachya (1), A. polystachya (2), M. verticillata and T. minuta espectively. The LC<sub>50</sub> values were 212.12, 230.74, 218.65 and 116.61 $\mu$ L air for A. decussates, A. polystachya (1), A. polystachya (2) and M. verticillata respectively. The toxicity of pure compounds was as follows: pulegone (LC<sub>50</sub>: 11.8µl/L air), Rcarvone (LC<sub>50</sub>: 17.5µl/L air), S-carvone (LC<sub>50</sub>: 28.1µl/L air), ocimenone (LC<sub>50</sub>: 42.3µl/L air), α-thujone (LC<sub>50</sub>: 65.5µl/L air) and menthone (LC<sub>50</sub>: 85.4 µl/L air) <sup>88</sup>. Citrus limon gave LC<sub>50</sub> value of 9.89µL/Lair against the weevil<sup>49</sup>.

The *M. soochowensis* essential oil displayed contact toxicity against *S. zeamais* adults (LC<sub>50</sub> = 25.45µg/adult). It also showed pronounced fumigant toxicity against the insects with  $LC_{50} = 12.19$  mg/L air<sup>89</sup>. Essential oil of Coriandrum sativum fruit gave a LD50 and LD95 of 145.49 and 10124.20µL/L air respectively at 24h and bioactive compound was found to be linalool with LC50 value of 172.37 µL/L air. Percentage mortality of S. oryzae was 70% at 625µL/L air after 24h exposure to essential oil of coriander<sup>69</sup>. The essential oils pine (Pinus palustris), lemon grass (Cymbopogon citratus), peppermint (Mentha piperita), citronella grass (Cymbopogon nardus), sweet acacia (Acacia farnesiana), cinnamon (Cinnamomum verum), sweet orange (Citrus sinensis), basil (Ocimum basilicum L), clove (Syzygium aromaticum), and star anise (Illicium verum) were tested for toxicity activity at different concentrations. Sweet Acacia gave 100% mortality at 10µL after 24h while basil oil and star anise have 100% mortality after 36h<sup>90</sup>. The insecticidal effect of essential oil from leaves of *Porophyllum linaria* on maize weevil was evaluated at different doses. The mortality (%) obtained on the 10<sup>th</sup> and 15th day was 43 and 82% respectively at 800 ppm, while with at ppm 30% mortality was obtained on 15th day. The  $LC_{50}$  and  $LC_{90}$  values obtained after exposure for 15 days were 329.01 and 1058.86ppm respectively<sup>91</sup>. Essential oils from Cymbopogon winterianus, Eucalyptus globulus, Eucalyptus staigeriana, Foeniculum vulgare, Ocimum basilicum, Ocimum gratissimum and Piper hispidinervum were investigated for their toxicity to S. zeamais at five concentrations. The contact and ingestion toxicities of the essential oils to adult S. zeamais decreased in the following order: P. hispidinervum > F. vulgare > O. basilicum > E. globulus > O. gratissimum > E. staigeriana > C. winterianus with  $LC_{50}$  of 5.12, 26.78, 26.90, 37.88, 47.47, 61.73 and 78.89µL/40g of maize grains, respectively. In the fumigant toxicity the LC<sub>50</sub> values were 2.1, 13.9, 15.8 and 19.4 $\mu$ L/40g of maize grains in grains treated with P. hispidinervum, O. basilicum, F. vulgare and E. globulus essential oils respectively<sup>92</sup>. Essential oils from Anethum graveolens, Petroselinum crispum, Foeniculum vulgare and *Cuminum cyminum* were tested at 25, 50, 75, 100, 156, 300, 525 and 600 mg/L air, and at 0, 30, 50, 100 and 150 µg per insect in fumigant and contact toxicity assays respectively. In the fumigant toxicity assay, LC50 values were found to be 157.1, 229.4, 442.8 and 535.8mg/L air for A. graveolens, C. cyminum, F. vulgare and P. crispum respectively. In the contact toxicity assay, LD<sub>50</sub> values were 111.3, 120.4 and 128.2mg/L air for A. graveolens, C. cyminum and P. crispum respectively<sup>93</sup>.

Toxicity of 28 essential oils was tested against the adults of *S. zeamais.* Their LD<sub>50</sub> in mg/cm<sup>2</sup> in contact toxicity assay were as follows cinnamon (0.04), tea tree (0.15), marjoram (0.18), peppermint (0.24), lavender, bulgarian (0.31), ylang ylang (0.32), geranium (0.36), lemongrass (0.37), patchouli (0.40), spearmint (0.40), clary sage (0.42), clove bud (0.47), *E. radiate* (0.61), rosemary (0.67), basil (0.77), *E. globulus* (0.89), citronella (1.01), cypress (1.23), orange sweet (1.40), bergamot (1.70) and sandal wood (1.87)<sup>94</sup>. Essential oil from *Elaeodendron schweinfurthianum* caused 89.1% mortality at 0.2ml/20g of grain<sup>95</sup>. The essential oils extracted from *R. officinalis, L. stoechas, S. viminea, M. septentrionalis, Eucalyptus* sp., and *L. alba* exhibited high mortality rates (>90%). The percentage mortality of the essential oils was as follows: *M. septentrionalis* (%M = 97), *S. viminea* (%M = 94), *L. stoechas* (%M = 93), *X. discreta* (%M = 73), *P. el-metanum* (%M = 70), *A. cumanensis* (%M = 40) and *P. nubigenum* (%M = 40). *L. stoechas* and *L. alba* gave LC<sub>50</sub> values of 303.4 and 254.1µL/L air respectively<sup>96</sup>.

*Tagetes minuta* and *M. longifolia* essential oils gave percentage repellency of more than 90% against maize weevil. The repellency *R. officinalis*, *H. odoratissimum* and *P. graveolens* essential oils were 51.1, 49.4 and 51.7%, respectively. Essential oil from *M. longifolia* exhibited over 70% mortality at  $32\mu$ L/L air in the fumigation tests<sup>73</sup>. *Lippia origanoides*, *E. citriodora*, and *T. lucida* essential oils, at concentrations of 0.063 -

 $0.503 \ \mu L/cm^2$  gave 92, 91, and 79% repellencies, respectively<sup>74, 97</sup>. The oils from *C. bergamia* and *L. hybrida* were effective against *S. zeamais*, with average repellencies of 56.3 and 50%, respectively, in a dilution of 0.1% of the oils in ether<sup>98</sup>. The repellency for *O. basilicum*, *O. americanum*, *O. tenuiflorum*, *C. hystrix*, *C. aurantifolia* and *E. caryophyllus* oils were 92.5, 75.0, 100.0, 67.5, 90.0 and 80.0% at  $0.21\mu L/cm^2$  respectively on filter paper<sup>76</sup>. Essential oils from *Eucalyptus benthamii*, *E. dunnii*, *E. globulus*, *E. viminalis* and *E. saligna* also exhibited repellency against *S. zeamais*<sup>99</sup>. Essential oil of *Daucus carota*, *Cedrus atlantica*, *Cinnamonium zeylanicum*, *Cupressus semperbirens*, *Lavendular angustifolia*, *Pogostemin cablin*, *Mentha pulegium* and *Melaleuca alternifolia* repelled 61.8, 80.0, 83.3, 81.1, 66.7, 71.0, 97.1 and 87.1% of *S. zeamais* at a concentration of 10 $\mu$ L /filter paper<sup>86</sup>.

Cymbopogon winterianus, Eucalyptus globulus, Eucalyptus staigeriana, Foeniculum vulgare, Ocimum basilicum., Ocimum gratissimum and Piper hispidinervum essential oils were investigated for their repellency to *S. zeamais* at five concentrations. The mean repellency percentages were: *E. staigeriana* (96.25%), *O. basilicum* (91.19%), *O. gratissimum* (90%), *C. winterianus* (81.82%), *E. globulus* (79.62%), *F. vulgare* (77.07%), and *P. hispidinervum* (49.37%)<sup>92</sup>. Essential oils from Anethum graveolens, Petroselinum crispum, Foeniculum vulgare and Cuminum cyminum were tested at concentrations of 16, 47, 78 and 156µg/cm<sup>2</sup>. In the area preference bioassays, essential oil extracted from *F. vulgare* was the most repellent ( $RD_{50} = 24\mu g/cm^2$ ) followed by *A. graveolens* ( $RD_{50} = 37.9\mu g/cm^2$ ), *C. cyminum* ( $RD_{50} = 45.2\mu g/cm^2$ ) and *P. crispum* ( $RD_{50} = 166.0\mu g/cm^2$ ). In the two-choice bioassays, the most repellent was *P. crispum* oil followed by *C. cyminum* oil<sup>93</sup>. The repellency of *I. verum* ranged between 43.3 and 70% at 2h, and between 20 and 46.7% at 24h after treatment. The essential oils of *X. discreta*, *Eucalyptus* sp., *P. el-methanum*, *P. pertomentellum* and *C. citratus* exhibited the greatest repellent effect at 24h<sup>96</sup>.

Essential oil from *aurelia sempervirens* reduced F1 emergence to 10.1% at a concentration of 5ml/kg grain at 5 weeks after treatment<sup>85</sup>. The number of adults that emerged from corn grains infested with eggs exposed to *P. hispidinervum* oil was lower than the control. The median lethal time to kill 50% ( $LT_{50}$ ) of the eggs was 16.7h at a concentration of 1.25µL/L, and 16.52h at a concentration of 1.87µL/L<sup>92</sup>.

Family	Botanical name	Common name	Bioactivity	Reference
Anacardiaceae	Schinus molle	False pepper	Toxicity and repellency	96
Annonaceae	Cananga odorata	Ylang ylang	Toxicity, repellency	94
Annonaceae	Xilopia discreta		Toxicity and repellency	96
Apiaceae	Anethum graveolens	Dill	Toxicity, repellency	93
Apiaceae	Coriandrum sativum L.	Coriander	Toxicity	55,63, 69
Apiaceae	Cuminum cyminum	Cumin	Toxicity, repellency	93
Apiaceae	Daucus carota	Carrot seed	Toxicity and repellency	86
Apiaceae	Foeniculum vulgare Mill	Fennel	Toxicity, growth inhibition	92-94, 96
			and repellency	
Apiaceae	Ostericum grosseserratum		Toxicity	84
Apiaceae	Ostericum sieboldii		Toxicity	80
Apiaceae	Petroselinum crispum	Parsley	Toxicity, repellency	93
Asteraceae	Ambrosia cumanesis Kunth	Altamisa	Toxicity and repellency	96
Asteraceae	Aphyllocladus decussatus Hieron		Toxicity	88
Asteraceae	Artemisia drancunculus	Estragon	Toxicity and repellency	96
Asteraceae	Artemisia lavandulaefolia		Toxicity	75
Asteraceae	Artemisia sieversiana		Toxicity	75
Asteraceae	Aster ageratoides	Starshine	Toxicity	82
Asteraceae	Blumea balsamifera	Sambong	Toxicity	83
Asteraceae	Porophyllum linaria	Pipicha	Toxicity	91
Asteraceae	Tagetes minuta L.	Southern marigold	Toxicity	73, 88
Asteraceae	Tagetes patula	French marigold	Toxicity	74
Asteraceae	Tagetes lucida	Mexican marigold		97
Atherospermatacea	Laurelia sempervirens (Ruiz &	Nutmeg	Toxicity and emergence	85
<u>e</u>	Pav.) Tul.		inhibition	
Burseraceae	Boswellia carterii	Frankincense	Toxicity, repellency	94
Canellaceae	Warburgia ugandensis	Greenheart	Toxicity, repellency, growth	39
			inhibition	
Cupressaceae	Cupressus sempervirens	Common cypress	Toxicity, repellence, growth inhibition	72, 86, 94, 95
Fabaceae	Acacia farnesiana	Sweet acacia	Toxicity	90
Geraniaceae	Pelargonium graveolens	Geranium	Toxicity, repellency	94
Hypericaceae	Hypericum juniperinum Kunth		Toxicity and repellency	96
Hypericaceae	Hypericum mexicanum		Toxicity and repellency	96
Hypericaceae	Hypericum myricariifolium Hieron		Toxicity and repellency	96
Lamiaceae	Dracocephalum moldavica	Dragon head	Toxicity	100

Table no 2: Plants species tested against Sitophilus zeamais

		T		04.04
Lamiaceae	Lavandula angustifolia	Lavender	Toxicity and repellency	86, 94
Lamiaceae	Lavandula hybrida	Lavender		98
Lamiaceae	Lavandula stoechas	French Lavender	Toxicity and repellency	96
Lamiaceae	Mentha piperita	Peppermint	Toxicity, repellency	90.94
Lamiacoao	Montha pulacium	Dennyroval	Toxicity and repallency	96, 21
Lamiaceae	Mentha pulegium	Concentration for the second s		04
Lamiaceae	Mentha spicata	Spearmint	Toxicity, repellency	94
Lamiaceae	Minthostachys septentrionalis		Toxicity and repellency	96
Lamiaceae	Minthostachys verticillata	Peperina	Toxicity	88
Lamiaceae	Mosla soochowensis Matsuda		Toxicity	89
Lamiaceae	Ocimum amaricanum	Holy basil	Toxicity and repellency	76
Lamiaceae		Source the sil	Toxicity and rependicy	70 00 00
Lamaceae	Ocimum basuicum L.	Sweet basil,	and repellency	76, 90, 92,
Lomiococo	Osimum angliasimum I	Clove besil	Torisity growth inhibition	94, 90
Lannaceae	Ocimum granssimum L	Clove basii	Toxicity, growth inhibition	92
			and repellency	
Lamiaceae	Ocimum suave		Toxicity, repellency, growth	95
			inhibition	
Lamiaceae	Ocimum tenuiflorum	Hairy basil	Toxicity and repellency	76
Lamiaceae	Origanum majorana	Sweet marjoram	Toxicity, repellency	94
Lamiaceae	Pogostamin cablin	Patchouli	Toxicity and repellency	86
Lamiaceae	Providence and the	Patel and		04
Lamiaceae	Pogostemon cabiin	Patchouli	Toxicity, repellency	94
Lamiaceae	Rosmarinus officinalis	Rosemary	Toxicity, repellency	94, 96
Lamiaceae	Salvia sclarea	Clary Sage	Toxicity, repellency	94
Lamiaceae	Satureja viminea	Tree mint	Toxicity and repellency	94
Lauraceae	Aniba robusta		Toxicity and repellency	96
Lauraceae	Aniha nuchum minor		Toxicity and repellency	96
Lauraceae	Particul puchur y-minor			<i>7</i> 0
Lauraceae	Beilschmiedia costaricensis		I oxicity and repellency	96
Lauraceae	Cinnamomum cassia	Cinnamon	Toxicity, repellency	94
Lauraceae	Cinnamomum triplinerve		Toxicity and repellency	96
Lauraceae	Cinnamomum verum J.S. Presl	Cinnamon	Toxicity	90
Lauraceae	Cinnamonium zavlanioum	Cinnamon	Toxicity and repellency	86
Lauraceae	Litaan anligifalin	Cliniamon	Toxicity and repenency	77
Lauraceae	Litsea salicifolia		Toxicity	11
Lauraceae	Nectandra acutifolia		Toxicity and repellency	96
Lauraceae	Ocotea longifolia		Toxicity and repellency	96
Lauraceae	Ocotea odorifera	Cinnamon	Toxicity and repellency	81
Lauraceae	Ocotea sp		Toxicity and repellency	96
Muriatianana	Vinola carinata		Toxicity and repellency	06
Myristicaceae		D d l		90
Myrtaceae	Eucalyptus benthamii	Bentham's gum	Repellency	99
Myrtaceae	Eucalyptus citriodora	Lemon-scented gum		97
Myrtaceae	Eucalyptus dunnii Maiden	Eucalyptus	Repellency	99
Myrtaceae	Eucalyptus globulus Labill	Dunn's white gum	Toxicity, growth inhibition	92, 94, 99
5	51 0	6	and repellency	
Murtaceae	Eucabintus radiata	Fucalyptus	Toxicity repellency	04
Martaccac	Eucaryphus radiana Eucaryphus radiana Smith	Discours	Toxicity, reperiency	72.00
Myrtaceae	Eucalyptus saligna Smith	Blue gum	Toxicity, repellency,	72,99
			emergence inhibition	
Myrtaceae	Eucalyptus sp		Toxicity and repellency	96
Myrtaceae	Eucalyptus staigeriana F. Muell.	Lemon-scented iron	Toxicity, growth inhibition	92
	ex F.M. Bailey	bark	and repellency	
Myrtaceae	<i>Eucalyntus viminalis</i> I abill	Manna gum	Repellency	99
Murtaceae	Eucoria agmontrollus	Clove	Toxicity and repallency	76
Maurta a a	Malalana altan 'C 1'			10
wiyrtaceae	metaleuca alternifolia	1 ea tree	1 oxicity and repellency	80,87,94
Myrtaceae	Syzygium aromaticum	Clove	Toxicity and repellency	90, 94, 96
Pinaceae	Cedrus atlantica	Cedarwood	Toxicity and repellency	86
Pinaceae	Pinus Palustris	Longleaf pine	Toxicity	90
Pinaceae	Pinus spp	Pine	Toxicity repellency	94
Diperaceas	Panaromia co	1 1110	Toxicity and repailance	96
Dimension	Pinese and a sp.		Toxicity and repetiency	90
Piperaceae	riper aequale		1 oxicity and repellency	96
Piperaceae	Piper aduncum	Spiked pepper	Toxicity and repellency	96
Piperaceae	Piper bogotense		Toxicity and repellency	96
Piperaceae	Piper asperiusculum		Toxicity and repellency	96
Piperaceae	Piper el-metanum		Toxicity and repellency	96
Diperaceas	Piner arianadar		Toxicity and repellency	96
r iperaceae			Toxicity and repetiency	90
Piperaceae	Piper hispidinervum C. DC	Long pepper	1 oxicity, growth inhibition	92
			and repellency	
Piperaceae	Piper holtonii		Toxicity and repellency	96
Piperaceae	Piper imperiale		Toxicity and repellency	96
Piperaceae	Piper marginatum	Marigold nenner	Toxicity and repellency	96
Diperaceas	Pinor nubia onum	iningoia poppoi	Toxicity and repellency	96
Dimension				<u> </u>
Piperaceae	riper periomentellum		i oxicity and repellency	90
Piperaceae	Piper pesaresanum		Toxicity and repellency	96
Poaceae	Cymbopogon citratus	Lemon grass	Toxicity and repellency	90, 94, 96
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Poaceae	Cymbopogon winterianus Jowitt	Java citronella	Toxicity, growth inhibition and repellency	92
Rutaceae	Citrus aurantifolia	Lime	Toxicity and repellency	76
Rutaceae	Citrus bergamia			98
Rutaceae	Citrus bigaradia	Bergamot	Toxicity, repellency	94
Rutaceae	Citrus hystrix	Kaffir lime	Toxicity and repellency	76
Rutaceae	Citrus limon	Lemon	Toxicity, repellency	49,94
Rutaceae	Citrus reticulata	Mandarin	Toxicity, repellency	94
Rutaceae	Citrus sinensis	Orange sweet	Toxicity and repellency	90, 94, 96
Rutaceae	Coleonema album	Confetti Bush	Toxicity and repellency	96
Santalaceae	Santalum album	Sandal wood	Toxicity, repellency	94
Schisandracea	Illicium simonsii		Toxicity	78
Schisandracea	Illicium verum Hook	Star anise	Toxicity and repellency	90, 96
Schisandracea	Illicium fragesii		Toxicity	79
Schisandracea	Kadsura heteroclita		Toxicity	101
Verbenaceae	Aloysia polystachya Griseb	Donkey tea	Toxicity	88
Verbenaceae	Lippia alba	Mat grass	Toxicity and repellency	96
Verbenaceae	Lippia javanica	Fever tree	Toxicity and repellency	102
Verbenaceae	Lippia origanoides Kunth		Toxicity and repellency	96, 97
Zingiberaceae	Curcuma longa	Turmeric	Toxicity and repellency	96

## V. Activity of Essential Oils Against Sitophilus granaries

Information retrieved from published data indicates that 23 essential oils were investigated against *Sitophilus granaries* (Table 3). *Origanum acutidens* and *Mentha pulegium* oils were reported to exhibit insecticidal activity against S. granaries<sup>103, 104</sup>. *Azilia eryngioides* essential oil gave  $LT_{50}$  values of 21.04, 17.44, 14.80 and 10.38h at concentrations of 37.03, 74.07, 111.11 and 148.14µL/L air respectively. The oil gave  $LC_{50}$  value of 20.05 µL/L air against the insect<sup>105</sup>. In the fumigant toxicity test, essential oil from flower of *L. angustifolia* gave  $LC_{50}$  and  $LC_{90}$  values of 1.5 and 4.mg/L respectively in the absence of wheat grains while the oil gave  $LC_{50}$  and  $LC_{90}$  values of 10.9 and 47.6mg/L respectively in the presence of the substrate. In the contact toxicity test,  $LC_{50}$  and  $LC_{90}$  values were recorded as 83.8 and 379.7µg/adult respectively at 24h after treatment, while  $LC_{50}$  and  $LC_{90}$  values were 58.3 and 208.3µg/adult respectively at 48h<sup>106</sup>. Essential oil extracts from Coriander, Mint and Sesame gave 90.0, 89.0 and 87.3% cumulative mortality of the insects 7 days after treatment <sup>107</sup>. Fumigant effect of essential oil of *Mentha piperita*, *Citrus cienensis* and *Nigella sativa* were investigated against *S. granarius*. All the three essential oils caused 100% mortality of the weevil at 24h after treatment at a concentration of 7%<sup>108</sup>. Essential oil from *Thymus pallescens* gave  $LC_{50}$  and  $LC_{90}$  values of 9.3 and 34.6µL/mL respectively in the contact test while in the fumigation assy *T. pallescens* oil gave  $LC_{50}$  and  $LC_{90}$  of 12.6 and 9.7µL/mL in contact and fumigation toxicity assays respectively<sup>109</sup>. Rosemary oil exhibited 58.41% fumigant toxicity against *S. granarius* 24h after treatment<sup>110</sup>.

Essential oil from flower of *Lavandula angustifolia* were evaluated for repellence activity against *S. granaries* at concentrations of 0.74, 1.49, 2.97, 5.94, 11.88, 23.76 and 47.52mg/L. Mean repellence values were higher than 80% (V repellent class) starting from the 0.441mg/cm<sup>2</sup> dose<sup>106</sup>. Essential oil from *T. pallescens* showed a strong repellent effect (83.4-100%) against the weevil<sup>109</sup>. *Rosmarinus officinalis, Laurus nobilis, Echinacea purpurea, Origanum majorana, Ocimum basilicum and Foeniculum vulgare* essential oil showed repellent effect under laboratory conditions. The essential oils caused 84.27-98.88% decrease in the progeny production of F1 generation<sup>110</sup>.

Family	Botanical name	Common name	Bioactivity	Reference
Apiaceae	Azilia eryngioides		Toxicity	105
Apiaceae	Coriandrum sativum L.	Coriander	Toxicity, oviposition and growth inhibition	107
Apiaceae	Foeniculum vulgare	Fennel	Toxicity, reduction in F1 progeny	110
			development, repellency	
Asteraceae	Echinacea purpurea	Echinacea	Toxicity, reduction in F1 progeny	110
			development, repellency	
Asteraceae	Helichrysum odoratissimum	Kooigoed	Toxicity, repellency	73
Asteraceae	Tagetes minuta L.	Marigold	Toxicity, repellency	73
Geraniaceae	Pelargonium graveolens	Geranium	Toxicity, repellency	73
Lauraceae	Laurus nobilis	Daphne	Toxicity, emergence inhibition, repellency	110
Lamiaceae	Lavandula angustifolia Miller	Lavender	Toxicity, repellency, anti-feedant	106, 111
Lamiaceae	Mentha balsamea Willd.	Peppermint	Toxicity	111
Lamiaceae	Mentha longifolia	Horse mint	Toxicity, repellency	73
Lamiaceae	Mentha piperita L.	Peppermint	Toxicity	108
Lamiaceae	Mentha pulegium	Pennyroyal	Toxicity	103
Lamiaceae	Ocimum basilicum	Basil	Toxicity, emergence inhibition, repellency	110

Table no 3: Plants species tested against Sitophilus granarius

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A	Review	Pesticidal	Activity of	of Essential	Oils against	Sitophilus ory	zae,
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Lamiaceae	Origanum acutidens		Toxicity	103
Lamiaceae	Origanum majorana	Marjoram	Toxicity, emergence inhibition, repellency	110
Lamiaceae	Rosmarinus officinalis	Rosemary	Toxicity, emergence inhibition, repellency	73, 110, 111
Lamiaceae	Satureja viminea	Mint	Toxicity, oviposition and growth inhibition	107
Lamiaceae	Thymus pallescens Noe.		Toxicity, repellency	109
Pedaliaceae	Sesamum indicum	Sesame	Toxicity, oviposition and growth inhibition	107
Poaceae	Cymbopogon citratus Stapf	Lemon grass	Toxicity, repellency	109
Ranunculacea	Nigella sativa L.	Black cumin,	Toxicity	108
e				
Rutaceae	Citrus cienensis (L.)	Sweet orange	Toxicity	108

## VI. Conclusion

This study shows that essential oils from plants have a potential in managing insect pests belonging to genus *Sitophilus*. The information retrieved has also indicated that most studies carried out were mostly geared towards identification of biopesticidal agent against *S. zeamais* more than *S. oryzae* and *S. granaries*. This could be an indication that *S. zeamais* is the most rampant and the most dreaded species. It is also evident that most studies ended with testing of the crude essential oils. The information about compounds responsible for the insecticidal activity of the oils is scanty despite the fact that insecticidal compounds from nature have preferred because they are environmentally safe. Future studies aimed in isolating and characterizing the active compounds from the essential oil is necessary. It is also necessary to develop effective formulations for controlling the pests.

#### References

- [1]. Plarre R. An attempt to reconstruct the natural and cultural history of the granary weevil, Sitophilus granarius (Coleoptera: Curculionidae). Eur J Entomol. 2010;107:1-11.
- [2]. Islam T, Iqbal J, Abdullah K, Khan E. Evaluation of some plant extracts against maize weevil, *Sitophilus zeamais* (coleoptera: curculionidae) under laboratory conditions. Pakistan Journal of Agricultural Sciences. 2017;54(4):737-741.
- [3]. Carrieri F, Raimo F, Pentangelo L. Fusarium proliferatum and Fusarium tricinctum as causal agents of pink root of onion bulbs and the effect of soil solarization combined with compost amendment in controlling their infections in field. Crop Prot. 2013;43:31-37.
- [4]. Ndirangu E, Opiyo SA, Ng'ang'a M. Chemical composition and repellency of Nigella sativa L. seed essential oil against Anopheles gambiae sensu stricto. Trends in Phytochemical Research. 2020a;4(2):77-84.
- [5]. Ndirangu E, Opiyo SA, Ng'ang'a M. Repellent Properties of Compounds and Blends from Nigella sativa Seeds against Anopheles gambiae. Basic Sciences of Medicine. 2020b;9(1):1-7.
- [6]. White N, Leesch J. Chemical control. In Subramanyam B, Hagstrum D. Integrated management of insects in stored products.1995;pp. 287-330. New York: Marcel Dekker.
- [7]. El-Wakeil N, Gaafar N, Sallam A, Volkmar C. Side effects of insecticides on natural enemies and possibility of their integration in plant protection strategies. In S. Trdan, Insecticides-Developments of Safer and More Effective Technologies. 2013;pp.3-56. Croatia, Intech press.
- [8]. Opiyo SA. Phytochemical, anti-bacterial and anti-fungal studies of *Conyza floribunda*. MSc thesis. 2006; Maseno University, Kenya
- [9]. Manguro L, Opiyo SA, Herdtweck E, Lemmen P. Triterpenes of *Commiphora holtziana* Oleogum resin. Canadian Journal of Chemistry. 2009;87(8):1173-1179.
- [10]. Manguro L, Opiyo SA, Asefa A, Dagne E, Muchori W. Chemical constituents of essential oils from three Eucalyptus species acclimatized in Ethiopia and Kenya. Journal of Essential Oil-Bearing Plants. 2010a; 13(5):561-567. https://doi.org/10.1080/0972060X.2010.10643863.
- [11]. Opiyo SA, Manguro LOA, Okinda-Owuor P, Ateka EM, Lemmen P. 7-alpha Acetylugandensolide and antimicrobial properties of *Warburgia ugandensis* extracts and isolates against sweet potato pathogens. Phytochemistry Letters. 2011a;4:161-165. https://doi.org/10.1016/j.phytol.2011.02.007.
- [12]. Opiyo SA, Manguro L, Owuor P, Ochieng C, Ateka E, Lemmen P. Antimicrobial compounds from *Terminalia brownii* against sweet potato pathogens. Natural Products. 2011b;1:116-120. DOI: 10.2174/2210316311101020116.
- [13]. **Opiyo** SA, Evaluation of *Warburgia ugandensis* extracts and compounds for crop protection against *Prostephanus truncates*. Advances in Analytical Chemistry. 2020b;10(2):15-19.
- [14]. Opiyo SA, Manguro L, Owuor P, Ateka E. Triterpenes from *Elaeodendron schweinfurthianum* and their antimicrobial activities against crop pathogens. American Journal of Chemistry. 2017;7(3):97-104. DOI: 10.5923/j.chemistry.20170703.03.
- [15]. Ochieng C, Ishola I, Opiyo SA, Manguro L, Owuor P, Wong K. Phytoecdysteroids from the stem bark of *Vitex doniana* and their anti-inflammatory effects. Planta Medica. 2013;79:52-59. DOI: 10.1055/s-0032-1327880.
- [16]. Ochung AA, Manguro LAO, Owuor PO, Jondiko IO, Nyunja RA, Akala H, Mwinzi P, Opiyo SA. Bioactive carbazole alkaloids from *Alysicarpus ovalifolius* (Schumach). Korean Society for Applied Biological Chemistry. 2015;58(6):839-846. https://doi.org/10.1007/s13765-015-0100-4.
- [17]. Shasany A, Lal R, Patra N, Darokar M, Garg A, Kumar S, et al. Phenotypic and RAPD diversity among *Cymbopogon winterianus* Jowitt accessions in relation to *Cymbopogon nardus* Rendle. Genetic Resources and Crop Evolution. 2000; 47: 553-559.
- [18]. Ochung AA, Owuor PO, Manguro LAO, Ishola OI, Nyunja RA, Ochieng CO, Opiyo SA. Analgesics from *Lonchocarpus eriocalyx* Harms. Trends in Phytochemical Research. 2018;2(4): 253-260.
- [19]. Jeruto P, Arama P, Anyango B, Nyunja R, Taracha C, Opiyo SA. Morphometric study of *Senna didymobotrya* (Fresen.) H. S. Irwin and Barneby in Kenya. Journal of Natural Sciences Research. 2017;7(6):54-69.
- [20]. Njoroge P, Opiyo SA. Antimicrobial activity of root bark extracts of *Rhus natalensis* and *Rhus ruspolii*. Basic Sciences of Medicine. 2019a;8(2):23-28. doi: 10.5923/j.medicine.20190802.01.
- [21]. Opiyo SA. Evaluation of efficacy of selected plant extracts in the management of fungal and bacterial diseases which affect sweet potato. Unpublished PhD thesis. Maseno University, Kenya 2011.

- [22]. Opiyo SA. A review of <sup>13</sup>C NMR spectra of drimane sesquiterpenes. Trends in Phytochemical Research. 2019;3(3):147-180.
- [23]. Makenzi A, Manguro L, Owuor P, Opiyo S. Chemical constituents of Ocimum Kilimandscharicum Guerke acclimatized in Kakamega Forest, Kenya. Bulletin of the Chemical Society of Ethiopia. 2019a;33(3):527-539. DOI: https://dx.doi.org/10.4314/bcse.v33i3.13.
- [24]. Makenzi A, Manguro L, Owuor P, Opiyo SA. Flavonol glycosides with insecticidal activity from methanol extract of Annona mucosa Jacq. leave. Trends in Phytochemical Research. 2019b;3(4):287-296.
- [25]. Opiyo SA. Repellent effects of Ocimum suave extracts and compounds against Prostephanus truncatus Horn. American Journal of Chemistry. 2021a;11(2):23-27, DOI: 10.5923/j.chemistry.20211102.01.
- [26]. Opiyo SA. Insecticidal Drimane Sesquiterpenes from Warburgia ugandensis against Maize Pests American. American Journal of Chemistry. 2021b;11(4):59-65. DOI: 10.5923/j.chemistry.20211104.01.
- [27]. Opiyo SA. A review of insecticidal plant extracts and compounds for stored maize protection. IOSR Journal of Applied Chemistry. 2021c;14(10):23-37. DOI: 10.9790/5736-1410012337.
- [28]. Manguro L, Ogur J, Opiyo SA. Antimicrobial constituents of *Conyza floribunda*. Webmed Central Pharmacology. 2010b;1(9);WMC00842.
- [29]. Ochieng CO, Opiyo SA, Mureka EW, Ishola IO. Cyclooxygenase inhibitory compounds from Gymnosporia heterophylla aerial parts. Fitoterapia. 2017;119:168-174. DOI: 10.1016/j.fitote.2017.04.015.
- [30]. Njoroge P, Opiyo SA. Some antibacterial and antifungal compounds from root bark of *Rhus natalensis*. American Journal of Chemistry. 2019b;9(5):150-158. doi: 10.5923/j.chemistry.20190905.03.
- [31]. Opiyo SA. Insecticidal Activity of Ocimum suave Willd Extracts and Compounds against Sitophilus zeamais Motschulsky. Basic Sciences of Medicine2020a;9(2):32-37 DOI: 10.5923/j.medicine.20200902.03.
- [32]. Kuria K, Opiyo SA. Characterization of immunogenic soluble crude proteins from *Biomphalaria Pfeifferi* against *Schistosoma mansoni*. Journal of Natural Sciences Research. 2020;10(12):28-34. DOI: 10.7176/JNSR/10-12-03.
- [33]. Opiyo SA, Ogur J, Manguro L, Itietze L, Schuster H. A new sterol diglycoside from *Conyza floribunda*. S. Afr. J. Chem. 2009;62:163-167.
- [34]. Opiyo SA, Manguro L, Ogur J, Wagai S. Bioactive constituents of *Conyza floribunda*. Research Journal of Pharmacology. 2010a;4(3):55-59.
- [35]. Opiyo SA, Ateka E, Owuor P, Manguro L, Miano D. Development of a multiplex PCR technique for simultaneous detection of sweet potato feathery mottle virus and sweet potato chlorotic stunt virus. Journal of Plant Pathology. 2010b;92(2):363-366.
- [36]. Opiyo SA, Ateka E, Owuor P, Manguro L, Karuri H. Survey of sweet potato viruses in western Kenya and detection of cucumber mosaic virus. Journal of Plant Pathology. 2010c;92(3):795-799.
- [37]. Opiyo S, Njoroge P, Ndirangu G, Kuria, K. A review of biological activities and phytochemistry of *Rhus species*. American Journal of Chemistry. 2021a;11(2):28-36. DOI: 10.5923/j.chemistry.20211102.02.
- [38]. Opiyo SA, Muna K, Njoroge P, Ndirangu G. Analgesic Activity of *Conyza floribunda* Extracts in Swiss Albino Mice. Journal of Natural Sciences Research. 2021b;12(12):1-6.
- [39]. Opiyo SA, Manguro L, Akinyi D, Ochung A, Ochieng C. Biopesticidal extractives and compounds from *Warburgia ugandensis* against maize weevil (*Sitophilus zeamais*). Natural Products. 2015;5(4):236-243. DOI:10.2174/2210315505666150916000539.
- [40]. Prates L, Faroni L, Heleno F, Queiroz M, Sousa A, Assis SM. Eugenol diffusion coefficient and its potential to control Sitophilus zeamais in rice. Scientific Reports. 2019;9:11161-11170.
- [41]. Tripathi A, Upadhyay S, Bhuiyan M, Bhattacharya P. A review on prospects of essential oils as biopesticide in insect-pest management. Journal of Pharmacognosy and Phytotherapy. 2009;1:52-63.
- [42]. Campolo O, Giunti G, Russo A, Palmeri V, Zappala L. Essential Oils in Stored Product Insect Pest Control. Journal of Food Quality. 2018. https://doi.org/10.1155/2018/6906105.
- [43]. Paranagama P, Abeysekera K, Nugaliyadde L, Abeywickrama K. Repellency and toxicity of four essential oils to *Sitophilus oryzae* L. (Coleoptera:Curculionidae). J. Natn.Sci.Foundation Sri Lanka. 2004;32:127-138.
- [44]. Sahaf B, Moharramipour S, Meshkatalsadat M. Chemical constituents and fumigant toxicity of essential oil from *Carum copticum* against two stored product beetles. Insect Science. 2007;14(3):213-218.
- [45]. Kambouzia J, Negahban M, Moharramipour S. Fumigant toxicity of *Eucalyptus leucoxylon* against stored product insects. American-Eurasian Journal of Sustainable Agriculture. 2009;3(2):229-233.
- [46]. Negahban M, Moharramipour S. Fumigant toxicity of Eucalyptus intertexta, Eucalyptus sargentii and Eucalyptus camaldulensis against stored-product beetles. Journal of Applied Entomology. 2007;131(4): 256-261.
- [47]. Mohamed M, Abdelgaleil S. Chemical composition and insecticidal potential of essential oils from Egyptian plants against Sitophilus oryzae (L.) (Coleoptera: Curculionidae) and Tribolium castaneum (Herbst) (Coleoptera: Tenebrionidae). Appl. Entomol. Zool. 2008;43 (4):599-607.
- [48]. Talukder D, Khanam L. Toxicity of four plant based products against three stored product pests. Journal of Bio-Science. 2009;17:149-153.
- [49]. Abdelgaleil S, Mohamed M, Shawir M, Abou-Taleb H. Chemical composition, insecticidal and biochemical effects of essential oils of different plant species from Northern Egypt on the rice weevil, *Sitophilus oryzae* L. Journal of Pest Science. 2016;89:219-229.
- [50]. Koutsaviti A, Antonopoulou V, Vlassi A. et al. Chemical composition and fumigant activity of essential oils from six plant families against *Sitophilus oryzae* (Col: Curculionidae). Journal of Pest Science. 2017; 83(1):1-14.
- [51]. Saroukolai AT, Moharramipour S, Meshkatalsadat MH. Insecticidal properties of *Thymus persicus* essential oil against ribolium castaneum and *Sitophilus oryzae*. Journal of Pest Science. 2010;83(1):3-8.
- [52]. Derbalah A, Ahmed S. Oil and powder of spearmint as an alternative to *Sitophilus oryzae* chemical control of wheat grains. Journal of Plant Protection Research. 2011;51(2):145-150.
- [53]. Ahmed ME. Fumigant toxicity of seven essential oils against the cowpea weevil, *Callosobruchus maculatus* (F.) and the rice weevil, *Sitophilus oryzae* (L.). Egypt. Acad. J. biolog. Sci. 2010;2(1):1-6.
- [54]. Mishra B, Tripathi S, Tripathi C. Bioactivity of two plant derived essential oils against the rice weevils Sitophilus oryzae (L.) (Coleoptera: Curculionidae). Proc. Natl. Acad. Sci., India, Sect. B Biol. Sci. 2013. DOI 10.1007/s40011-012-0123-0.
- [55]. Rani P. Fumigant and contact toxic potential of essential oils from plant extracts against stored product pests. J. Biopest. 2012;5(2):120-128.
- [56]. Kardinan A, Maris P, Rizal M. Preliminary study of insecticidal effect of citronella grass essential oil (*Cymbopogon nardus*) against post harvest pest *Sitophilus oryzae*. IOP Conf. Series: Earth and Environmental Science. 2021;743(1):012015
- [57]. El-Bakry A, Abdel-Aziz N, Sammour E, Abdelgaleil S. Insecticidal activity of natural plant essential oils against some stored product insects and their side effects on wheat seed germination. Egyptian Journal of Biological Pest Control. 2016;26:83-88.

- [58]. Chaubey M. Fumigant and contact toxicity of Allium sativum (Alliaceae) essential oil against Sitophilus oryzae L. (Coleoptera: Dryophthoridae). Entomol. Appl. Sci. Lett. 2016a;3(2):43-48.
- [59]. Chaubey M. Insecticidal activities of *Cinnamomum tamala* (Lauraceae) essential oil against *Sitophilus oryzae* L. (Coleoptera: Curculionidae). Int. J. Entomol. Res. 2016b;4 (3):91-98.
- [60]. Khani M, Marouf A, Amini S, Yazdani D, Farashiani M, Ahvazi M, et al. Efficacy of three herbal essential oils against rice weevil, Sitophilus oryzae (Coleoptera: Curculionidae). Journal of Essential Oil Bearing Plants. 2017;20(4):937-950.
- [61]. Abdel-Fattah N, Doaa M. Fumigant and repellent effects of some natural oils against Sitophilus oryzae (L.) and Callosobruchus maculatus (F.). Egypt. J. Agric. Res. 2017;95(1):123-131.
- [62]. Jayakumar M, Arivoli S, Raveen R, Tennyson S. Repellent activity and fumigant toxicity of a few plant oils against the adult rice weevil *Sitophilus oryzae* Linnaeus (Coleoptera: Curculionidae). Journal of Entomology and Zoology Studies. 2017;5(2):324-335.
- [63]. Choi S, Lee H. Insecticidal activities of Russia coriander oils and these constituents against Sitophilus oryzae and Sitophilus zeamais. J Appl Biol Chem. 2018;61(3):239-243.
- [64]. Singh G, Chaudhary K, Rana R. Efficacy of plant derived essential oils against *Sitophilus oryzae* (L.) in stored wheat grains. Journal of Plant Development Sciences. 2018;10(11):633-636.
- [65]. Hosny A, Hasan N, Zayed G, Frawila H. Bioactivity of marjoram oil and powder against the rice weevil *Sitophilus oryzae*. Research & Reviews: Journal of Botanical Sciences. 2018;7(2):14-21.
- [66]. Amini S, Nohooji M, Khani M, Labbafi M, Khalighi-Sigaroodi F. Biological activity of some essential oil constituents in four Nepeta L. species against Sitophilus oryzae L. Biodiversitas. 2019;20:338-343.
- [67]. Kathirvelu C, Maline A. Fumigant effect of essential oils against pulse beetle (*Callosobruchus chinensis* L.) and rice weevil (*Sitophilus oryzae* L.) in stored products. Indian Journal of Science and Technology. 2020;13(25):2575-2581.
- [68]. Owolabi M, Ogundajo A, Alafia A, Ajelara K.O, Setzer W. Composition of the essential oil and insecticidal activity of Launaea taraxacifolia (Willd.) Amin ex C. Jeffrey Growing in Nigeria. Foods. 2020;9:914-921.
- [69]. Eljazi J, Bachrouch O, Salem N, Msaada K, Aouini J, Hammami M, et al. Chemical composition and insecticidal activity of essential oil from coriander fruit against *Tribolium castaenum*, *Sitophilus oryzae*, and *Lasioderma serricorne*. International Journal of Food Properties. 2018;20:sup3, S2833-S2845.
- [70]. Chaubey M. Fumigant toxicity of essential oils against rice weevil *Sitophilus oryzae* L. (Coleoptera: Curculionidae). Journal of Biological Sciences. 2011;11(6):411-216.
- [71]. Kiran S, Prakash B. Toxicity and biochemical efficacy of chemically characterized *Rosmarinus officinalis* essential oil against *Sitophilus oryzae* and *Oryzaephilus surinamensis*. Ind. Crops. Prod. 2015;74:817-823.
- [72]. Tapondjou A, Adler C, Fontem D, Bouda H, Reichmuth C. Bioactivivities of cymol and essential oils of *Cupressus sempervirens* and *Eucalyptus saligna* against *Sitophilus zeamais* Motchulsky and *Tribolium confusum* du Val. Journal of Stored Products Research. 2005;41(1):91-102.
- [73]. Odeyemi O, Masika P, Afolayan A. Evaluation of the activities of five essential oils against the stored maize weevil. Natural Product Communications. 2008;3(7):1097-1102.
- [74]. Restello R, Menegatt C, Mossi, A. Efeito do óleo essencial de *Tagetes patula* L. (Asteraceae) sobre *Sitophilus zeamais* Motschulsky (Coleoptera: Curculionidae). Revista Brasileira de Entomologia. 2009;53: 304-307.
- [75]. Liu Z, Liu Q, Chu S, Jiang G. Insecticidal activity and chemical composition of the essential oils of Artemisia lavandulaefolia and Artemisia sieversiana from China. Chem Biodiv. 2010;7:2040-2045.
- [76]. Kerdchoechuen O, Laohakunjit N, Singkornard S. Essential oils from six herbal plants for biocontrol of the maize weevil. HortScience. 2010;45(4):592-598.
- [77]. Ko K, Juntarajumnong W, Chandrapatya A. Insecticidal activities of essential oils from fruits of *Litsea salicifolia* Roxb. ex Wall. against *Sitophilus zeamais motschulsky* and *Tribolium castaneum* (Herbst). Pakistan Journal of Zoology. 2010;42(5):551557.
- [78]. Chu S, Liu S, Jiang G, Liu Z. Composition and toxicity of essential oil of *Illicium simonsii* Maxim (Illiciaceae) fruit against the maize weevils. Rec Nat Prod. 2010;4:205-210.
- [79]. Wang C, Liu P, Yang K, Zeng Y, Liu Z, Du S, et al. Chemical composition and toxicities of essential oil of *Illicium fragesii* fruits against *Sitophilus zeamais*. Afr J Biotechnol. 2011;10:18179-18184.
- [80]. Liu Z, Chu S, Jiang G. Insecticidal activity and composition of essential oil of Ostericum sieboldii (Apiaceae) against Sitophilus zeamais and Tribolium castaneum. Rec Nat Prod. 2011;5:74-81.
- [81]. Mossi A, Zanella C, Kubiak G, Lerin L, Cansian R, Frandoloso F, et al. Essential oil of Ocotea odorifera: An alternative against Sitophilus zeamais. Renewable Agriculture and Food Systems. 2014;29:161-166.
- [82]. Chu S, Liu S, Liu Q, Jiang G, Liu Z. Chemical composition and insecticidal activities of the essential oil of the flowering aerial parts of *Aster ageratoides*. Journal of Serbian Chemical Society. 2013a;78(2):209-216.
- [83]. Chu S, Du S, Lui Z. Fumigant compounds from essential oil of Chinese Blumea balsamifera leaves against the maize weevil (Sitophilus zeamais). Journal of Chemistry. 2013b;2013:1-7.
- [84]. Chu S, Lui Q, Du S, Liu Z. Chemical composition and insecticidal activity of the essential oil of the aerial parts of Ostericum grosseserratum (Maxim) Kitag (Umbelliferae). Trop J Pharm Res. 2013c;12(1):99-103.
- [85]. Torres C, Silva G, Tapia M, Rodríguez J, Figueroa I, Lagunes A, et al. Insecticidal activity of *Laurelia sempervirens* (Ruiz & Pav.) Tul. essential oil against *Sitophilus zeamais* Motschulsky. Chil J Agric Res. 2014;74:421-426.
- [86]. Kim Y, Kim K, Lee J, Lee H, Lee S. Fumigant toxicity and the repellent effect of essential oils against Sitophilus zeamais Motschulsky. Korean J. Appl. Entomol. 2012;51(4):389-395.
- [87]. Liao M, Xiao J, Zhou L, Liu Y, Wu X, Hua R, et al. Insecticidal activity of *Melaleuca alternifolia* essential oil and RNA-Seq analysis of *Sitophilus zeamais* transcriptome in response to oil fumigation. PLoS ONE. 2016;11(12): e0167748. doi:10.1371/journal.pone.0167748.
- [88]. Herrera J, Zunino M, Massuh Y, Pizzollito R. Dambolena J. Fumigant toxicity of five essential oils rich in ketones against Sitophilus zeamais (Motschulsky). Agriscientia. 2014;31(1):35-41.
- [89]. Chen X, Chen R, Luo Z. Chemical composition and insecticidal properties of essential oil from aerial parts of *Mosla soochowensis* against two grain storage insects. Trop J Pharm Res. 2017;16(4):905-910.
- [90]. Pangnakorn U, Chuenchooklin S. Evaluation of essential oils against Sitophilus zeamais (Motshulsdy) (Coleoptera: Curculionidae). Adv Med Plant Res. 2018; 6(2):19-25.
- [91]. Cruz J, Cruz A, Alvarado E, Sánchez E, Valenzuela N, Nicolás B, et al. Efficiency of the essential oil of *Porophyllum linaria* (Asteraceae) a Mexican endemic plant against *Sitophilus zeamais* (Coleoptera: Curculionidae). Journal of Insect Science. 2019 doi: 10.1093/jisesa/iez079.
- [92]. Araújo A, Oliveira J, Franca S. Toxicity and repellency of essential oils in the management of *Sitophilus zeamais*. R. Bras. Eng. Agríc. Ambiental. 2019;23(5):372-377.

- [93]. Rosa J, Oliveira L, Sousa R, Escobar C.F. Bioactivity of some Apiaceae essential oils and their constituents against *Sitophilus zeamais* (Coleoptera: Curculionidae). Bulletin of Entomological Research. 2019;110: 406-416.
- [94]. Yang Y, Isman M, Tak J. Insecticidal activity of 28 essential oils and a commercial product containing *Cinnamomum cassia* Bark essential oil against *Sitophilus zeamais* Motschulsky. Insects. 2020;11:474-488.
- [95]. Opiyo SA. Insecticidal activity of *Elaeodendron schweinfurthianum* extracts and compounds against *Sitophilus zeamais* Motschulsky. American Journal of Chemistry. 2020c;10(3):39-44. doi: 10.5923/j.chemistry.20201003.01.
- [96]. Patiño BW, Nagles GL, Bustos CJ, Delgado AW, Herrera DE, Suárez L, et al. Effects of essential oils from 24 plant species on Sitophilus zeamais Motsch (Coleoptera, Curculionidae). Insects. 2021;12:532-550.
- [97]. Nerio LS, Oliveiro VJ, Stashenko EE. Repellent activity of essential oils from seven aromatic plants grown in Colombia against Sitophilus zeamais Motschulsky (Coleoptera). Journal of Stored Products Research. 2009;45:212-214.
- [98]. Cosimi S, Rossi E, Cioni PL, Canale A. Bioactivity and qualitative analysis of some essential oils from Mediterranean plants against stored-product pests: Evaluation of repellency against *Sitophilus zeamais* Motschulsky, *Cryptolestes ferrugineus* (Stephens) and *Tenebrio molitor* (L.). Journal of Stored Products Research. 2009;45:125-132.
- [99]. Mossi AJ, Astolfi V, Kubiak G, Lerin L, Zanella C, Toniazzo G, et al. Insecticidal and repellency activity of essential oil of *Eucalyptus* sp. against *Sitophilus zeamais* Motschulsky (Coleoptera, Curculionidae). Journal of the Science of Food and Agriculture. 2010;91:273-277.
- [100]. Chu S, Liu S, Liu Z, Du, S. Composition and toxicity of Chinese Dracocephalum moldavica (Labiatae) essential oil against two grain storage insects. Journal of Medicinal Plant Research. 2011;5(18): 4621-4626.
- [101]. Li H, Bai, C, Chu S, Zhou L, Du S, Liu Z, et al. Chemical composition and toxicities of the essential oil derived from *Kadsura* heteroclita stems against Sitophilus zeamais and Meloidogyne incognita. J Med Plants Res. 2011;5:4943-4948.
- [102]. Kamanula J, Belmain, S, Hall D, Farman D, Goyder D, Mvumi B, et al. Chemical variation and insecticidal activity of *Lippia javanica* (Burm. F.) Spreng essential oil against *Sitophilus zeamais* Motschulsky. Ind. Crops Prod. 2017;110:75-82.
- [103]. Caglar O, Calmasur O, Aslan I, Kaya O. Insecticidal effect of essential oil of *Origanum acutidens* against several stored product pests. Fresenius Environmental Bulletin. 2007;16(11):1395-1400.
- [104]. Mahmoudvand M, Abbasipour H, Basij M, Hosseinpour M, Rastegar F, Nasiri M. Fumigant toxicity of some essential oils on adults of some stored-product pests. Chilean Journal of Agricultural Research 2011;71(1):83-89.
- [105]. Ebadollahi A, Mahboubi M. Insecticidal activity of the essential oil isolated from *Azilia eryngioides* (Pau) Hedge Et Lamond against two beetle pests. Chilean Journal of Agricultural Research. 2011;71(3):406-411.
- [106]. Germinara G, Stefano M, Acutis L, Pati S, Delfine S, Cristofaro A, et al. Bioactivities of Lavandula angustifolia essential oil against the stored grain pest Sitophilus granaries. Bulletin of Insectology. 2017;70 (1):129-138.
- [107]. Sabbour M. Toxicity of the three oils extract against *Sitophilus granaries* under laboratory and store conditions. Int. Res. J. Biol. Sci. 2020;2(1):14-18.
- [108]. Erdogan P, Mustafa Z. Fumigant activity of some essential oil against wheat weevil, *Sitophilus granarius* L. (coleoptera: curculionidae). J Bacteriol Mycol. 2021;9(2):57-60.
- [109]. Moutassem D, Bellik Y, Sannef M. Toxicity and repellent activities of *Thymus pallescens* and *Cymbopogon citratus* essential oils against *Sitophylus granaries*. Plant Protection Science. 2021;57(4): 297-309.
- [110]. Teke M, Mutlu C. Insecticidal and behavioral effects of some plant essential oils against *Sitophilus granarius* L. and *Tribolium castaneum* (Herbst). Journal of Plant Disease and Protection. 2021;128:109-119.
- [111]. Laznik Z, Matej VM, Trdan S. Efficacy of four essential oils against Sitophilus granarius (L.) adults after short-term exposure. Afr. J. Agric. Res. 2012;7(21):3175-3181.

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