Preclinical Toxicity Studies of Oxalis Acetocella by Brine Shrimp Lethality Test

Pushpalal Shresta¹ Mohammed Vazir
¹ Post Graduate, Dept of Pharmacology, Hillside college of Pharmacy and Research center, Bangalore, India.
² HOD, Dept of Pharmacology, Hillside college of Pharmacy and Research center, Bangalore, India.

Corresponding Author: PushpalalShresta

Date of Submission: 22-02-2019   Date of acceptance: 08-03-2019

I. Introduction

Wounds are generally classified as, wounds without tissue loss and wounds with tissue loss, such as burn wounds, wounds caused as a result of trauma, abrasions or as secondary events in chronic ailments e.g.: venous stasis, diabetic ulcers or pressure sores and iatrogenic wounds such as skin graft donor sites and derma abrasions. Wound healing involves complex series of interactions between different cell types, cytokine mediators and the extra cellular matrix. The phases of normal wound healing include haemostasis, inflammation, proliferation, and remodeling. *Oxalis acetosella* is a rhizomatous plant from the genus Oxalis, common in most of Europe and parts of Asia. The binomial name is *Oxalis acetosella*, because of sour taste. The plant has reported for antioxidant, diuretic, antiscorbutic, refrigerant action and on decoction. It gives relief from fever, haemorrhage, urinary disorders, the juice of leaves is used as gargle and is remedy for ulcers in the mouth, nutritional disorder. The previous studies reported the antioxidants influences wound healing activity. Hence the present study is undertaken to evaluate the wound healing activity of *Oxalis acetosella*.

Wood Sorrel (*Oxalis Acetosella*) is an amazing wild edible that has leaves and flowers that are best eaten raw. Commonly called sour grass, and often confused with “yellow clover”, Wood Sorrel is a sour treat that will quench your thirst, while you enjoy it as a trail side nibble. The familiar acid of the Wood Sorrels is refreshing in warm weather and the leaves have long been popular with trampers and mountain climbers for their mildly tonic and refreshing properties. In small quantities the foliage is a wholesome addition to a salad, but, on account of the abundance of oxalic acid contained in the plant, it is unwise to eat the foliage in very large quantity.

II. Methodology

**Plant Survey & collection & Authenticifying Plant:**

The leaves of Oxalis acetocellawascollected from the local region, TamilNadu and authenticated by Taxonomist. The herbarium of the plant was been kept in our institution. The leaves were dried in the shade for 7days at room temperature (28°C).

**Extraction of Plant materials:**

The crisp plant materials were washed with running faucet water and shade dried. The examples were squashed to coarse powder by processor. These coarse powders (25g) were then exposed to defatted with oil ether and after that separated with ethanol till weariness by utilizing Soxhlet apparatus. The gathered concentrates were saved at that point taken up for further examinations. The DMSO (Dimethylsulfoxide) is go about as broken up solvents for these concentrates.

**Preliminary Qualitative Biochemical Tests for detection of chemical constituent:**

Preliminary phytochemicals analysis was carried out for ethanol extract as per standard methods.

**Detection of Alkaloids:**

The powder of ethanol extract was dissolved in dilute hydrochloricacidanted filtered. The filtrates were used to test the presence of alkaloids.

*Mayer’s test:*

Filtrate was treated with Mayer’s reagent. Formation of a yellow cream precipitate indicates the presence of alkaloids.

*a) Wagner’s test:*

Filtrate was treated with wagner’s reagent. Formation of reddish brown precipitate indicates the presence of alkaloids.
Preclinical Toxicity Studies of Oxalis Acetocella by Brine Shrimp Lethality Test

Detection of Flavonoids

Leadacetetatest: Ethanol extract powder was treated with few drops of leadacetate solution. Formation of yellow color precipitate indicates the presence of flavonoids.

a)Sulphuric acid test: Extract was treated with few drops of sulphuric acid. Formation of orange color indicates the presence of flavonoids.

Detection of Steroids

2ml of aceticanhydridewas added to 0.5g of the powder ethanol extract, with 2ml of H₂SO₄. The colour changed from violet to blue or green in sample indicate the presence of steroids.

Detection of Terpenoids

Salkowski’s test

0.2g of the extract of the whole plant sample was mixed with 2ml of chloroform and 3ml of concentrated sulphuric acid was carefully added to form a layer. A reddish brown coloration of the inner face indicates the presence of terpenoids.

Detection of Anthraquinones

Borntrager’s test

About 0.2g of the powered extract was boiled with 10% HCl for few minutes in a water bath. It was filtered and allowed to cool. Equal volume of CHCl was added to the filtrate. Few drops of 10% NH₄ were added to the mixture and heated. Formation of pink colour indicates the presence anthraquinones.

Detection of Phenols

Ferric chloride test: Extract was treated with few drops of ferric chloride solution. Formation of bluish black color indicates the presence of phenol.

b)Leadacetetetests: Extract was treated with few drops of lead acetate solution. Formation of yellow color precipitate indicates the presence of phenol.

Detection of Saponins:

About 0.2g of the ethanol powder extract was shaken with 5ml of distilled water. Formation of frothing (appearance of creamy miss of small bubbles) shows the presence of saponins.

Detection of Carbohydrates:

Extract was dissolved in 5ml distilled water and filtered. The filtrate was used to test the presence of carbohydrates.

Detection of Oils and Resins:

Test solution was applied on filter paper. It develops transparent appearance on the filter paper. It indicates the presence of oils and resins.

To evaluate the cytotoxic studies by leaf extract

Brine Shrimp Lethality Assay (BSLA)

(Artemiasalina) eggs were hatched in artificial sea water prepared from commercial sea salt 38g/L. A lamp was placed above the openside of the tank to attract the hatched shrimps close to the tank wall. After 24 hours, the shrimps matured as nauplii(Artemiasalina) and were ready for the assay. The brine shrimp lethality bioassay was carried out on the ethanol extract using the standard. Twenty milligrams of the extract was dissolved in 1ml of Prophylene glycol/Tween 80/water (4:1:4) to give a crude extract concentration of 20mg/ml. A crude fold serial dilution was carried out with salt water to obtain a test solution in the range of 0.1-10mg/ml. Each concentration was tested in triplicate. A test tube contained prophylene glycol/Tween80/water(4:1:4) in 5ml of salt water was used as the negative control. Ten milligrams of potassium dichromate (as positive control) was dissolved in prophylene glycol/Tween80/water (4:1:4) and serially diluted, to obtain test concentrations ranging from 0.01 to 5mg/ml. A suspension of larvae(0.1ml), containing about 10-15 larvae, was added into each test tube and incubated for 24 hours. The test tubes were then examined and the number of dead larvae in each bottle. The total number of shrimps in each bottle was counted and recorded. The death percentage and lethal concentration (LC₅₀) were determined using statistical analysis.
Preclinical Toxicity Studies of Oxalis Acetocella by Brine Shrimp Lethality Test

Equation:
Percentage of Death (%): (Total naupii - Alive naupii) x 100/% Total naupii

Plant extracts:
The percentage yield of the ethanolic leaf extract of Oxalis Acetocella was found to be 68.78% obtained by soxlet extraction process. Both powder as well as extract of Oxalis Acetocella were used in quantitative analysis and verified.

Qualitative phytochemical analysis of Oxalis Acetocella leaves.

**Table 1:** Phytochemical characteristics of ethanolic leaves extract & powder of Oxalis Acetocella leaves.

<table>
<thead>
<tr>
<th>Sl. No</th>
<th>Phytochemical Test</th>
<th>Ethanolic leaf extract of Oxalis Acetocella</th>
<th>Ethanolic leaf powder of Oxalis Acetocella</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Carbohydrates</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>2</td>
<td>Alkaloids</td>
<td></td>
<td></td>
</tr>
<tr>
<td>3</td>
<td>Saponins</td>
<td>+</td>
<td></td>
</tr>
<tr>
<td>4</td>
<td>Phenolics</td>
<td></td>
<td></td>
</tr>
<tr>
<td>5</td>
<td>Flavonoids</td>
<td></td>
<td></td>
</tr>
<tr>
<td>6</td>
<td>Steroids</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>7</td>
<td>Tannins</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Brine Shrimp Lethality Test:

**Table 2:** Effect of ethanolic leaves extract of Oxalis Acetocella on Brine shrimp lethality assay.

<table>
<thead>
<tr>
<th>Sl No</th>
<th>Time (hours)</th>
<th>Conc</th>
<th>No of Nauplii</th>
<th>Total no of Surviving nauplii</th>
<th>Total No of dead nauplii in (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>µg</td>
<td></td>
<td>O. acetocella Leaf extract</td>
<td>O. acetocella Leaf Powder</td>
</tr>
<tr>
<td>1</td>
<td>2</td>
<td>0.1%</td>
<td>25</td>
<td>18</td>
<td>07</td>
</tr>
<tr>
<td>2</td>
<td>4</td>
<td>0.2%</td>
<td>25</td>
<td>15</td>
<td>10</td>
</tr>
<tr>
<td>3</td>
<td>6</td>
<td>0.4%</td>
<td>25</td>
<td>10</td>
<td>15</td>
</tr>
<tr>
<td>4</td>
<td>8</td>
<td>0.8%</td>
<td>25</td>
<td>6</td>
<td>09</td>
</tr>
</tbody>
</table>

As shown in the Table 2, Ethanolic leaves extract of Oxalis Acetocella was subjected to Brine Shrimp lethality bioassay for possible cytotoxic action was carried out against at different concentrations of 1µg, 10µg, 100 µg and 1000 µg. The suspensions of larva were examined after 8 hours of the incubation period. At 100µg/ml concentration of leaves extract of Oxalis Acetocella was shown 50 percent of mortality rate and 1000µg/ml concentration of leaves extract of Oxalis Acetocella were showed 100 percent of mortality rate. In this study, ethanolic leaves extract of Oxalis Acetocella was found to be toxic to Brine Shrimp nauplii, with LC50 value of 100µg/ml.

References


DOI: 10.9790/3008-1401053234  www.iosrjournals.org  34 | Page