Biosynthesis and characterisation of zinc doped iron oxide nanoparticles from *pedalium murex* and its new avenues in pharmacological applications

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**Abstract:** Biosynthesis of metal oxide nanoparticles gains attention over the past decades due to its eco-friendly nature and provides nanoparticles with controlled size and shape. In the present work, zinc doped iron oxide nanoparticles was synthesized by co-precipitation method using *pedalium murex* leaf extract as reducing agent and capping agent. The resulting nano-particles were characterized by UV-Vis, FTIR techniques. XRD analysis shows that synthesized ZnIONPs are in hematite phase, rhombohedral in structure. The antimicrobial and anti diabetic activities of synthesized ZnIONPs are analysed by disc diffusion and pancreatic α-amy method and it shows that high level of inhibition.

**Keywords:** Anti-diabetic, Anti-microbial, Co-precipitation, Pedalium murex, ZnIONPs

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**I. Introduction**

Nanostructure iron oxides have attracted the attention of several researchers because of their technological utilizations such as magnetic storage media, catalysis, biomedicine, biotechnology, biosensors, etc due to their large surface area, robustness and availability [1-3]. Iron oxides can be divided into 4 types depending on their different molecular structures: α-Fe₂O₃, γ-Fe₂O₃, α-Fe₃O₄, and γ-Fe₃O₄ [4-5]. Hematite is the most common phase of iron oxide that occurs naturally in rocks and soil. It is thermodynamically the most stable iron oxide with n-type semiconductor properties. Hematite has a complex magnetic behavior that is highly dependent on temperature and particle size. Hematite has a Néel temperature of about 948 K [6] and also undergoes a low temperature transition just below room temperature called the Morin transition [7].

Doping of transition metal ions into Fe₂O₃ can improve the properties of nano crystalline materials by narrowing the energy-band gap and inhibiting electron-hole recombination [8]. In addition, several dopant species have been introduced into α-Fe₂O₃ in attempts to control materials properties, including Ti⁴⁺, Si⁴⁺, Nb⁵⁺, V⁵⁺, Al³⁺, Zn²⁺, and Pt⁶⁺ [9]. Hence doping with transition metal impurities is a widely applied process in material science that involves incorporation of atoms or ions of appropriate elements into host lattices to yield hybrid materials with desirable properties and function. Development of new magnetic semiconductors based on the metal doped hematite is a subject of increasing research interest.

Zinc belongs to the microelements group/category that can play a vital role in many important biochemical reactions and physiological processes, such as development and growth of cells [10]. Different types of synthesis techniques are used for the synthesis of zinc-doped iron oxide nanomaterials such as bottom-up approach, viz sol-gel technique, chemical precipitation technique and in top-down, ball milling etc. Recently, great efforts were made to use green and eco-friendly method for synthesis of nanosize materials. These efforts include the use of plant or fruit extracts as surfactant [11]. The plant extracts release a variety of metabolites including carbohydrates, polysaccharides, phenols, amino acids, and vitamins, which can act as capping agents, reducing agents, stabilizing and chelating agents. The use of plant extracts in the synthesis can influence the size, the shape, and the morphology of the nanoparticles. They generate nanoparticles with high dispersity, high stability, and narrow size distribution [12-13].

The present investigation deals with the structural and magnetic characterization of zinc substituted hematite nanoparticles synthesized by co-precipitation method and it also aims to detect the anti microbial, anti diabetic activity of these nanoparticles.

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II. Materials and Methods

Plant source- leaves of *Pedalium murex*
The plant sources were collected from Sholavandan , Madurai District.

<table>
<thead>
<tr>
<th>Kingdom</th>
<th>Plantae</th>
</tr>
</thead>
<tbody>
<tr>
<td>Division</td>
<td>Magnoliophyte</td>
</tr>
<tr>
<td>Class</td>
<td>Magnoliopsida</td>
</tr>
<tr>
<td>Order</td>
<td>Lamiales</td>
</tr>
<tr>
<td>Family</td>
<td>Pedaliaceae</td>
</tr>
<tr>
<td>Genus</td>
<td>Pedalium</td>
</tr>
<tr>
<td>Species</td>
<td>Pedalium murex Linn</td>
</tr>
</tbody>
</table>

2.1 Plant description
Traditionally, P. murex was utilized in various ways, either as a whole plant or individual plant parts or sometimes in different special preparations. The leaves are cooked and eaten as a vegetable. In the form of powder, it can be applied locally with butter are used for rheumatic pains [14]. Leaf decoction is used to treat diabetes [15]. Leaves of P. murex are used to treat ulcers, dysuria, Bone fracture, diarrhea and in splenic enlargement [16]. Leaves of Aloevera, P. murex and Bauhinia racemosa crushed together and mixed with water can be given to animals three times a day can relief food poisoning in cattle. An infusion or extract prepared from the different parts of the plant in cold water is used as demulcent, diuretic and also found to be best used in the treatment of disorders of urinary systems such as gonorrhea, dysuria, incontinence of urine and vice versa[17]. The plant is also used by the local people as analgesic and antipyretic activities [18-19].

![Image of Pedalium murex leaves]

Fig.2.1 Leaf of *Pedalium murex*

2.2 Chemicals required
Stock solutions of FeCl$_3$.7H$_2$O, FeSO$_4$.7H$_2$O and ZnCl$_2$.6H$_2$O, NH$_4$OH.

2.3 Preparation of plant extract
Pedalium murex plant were cut into pieces and boiled with 100ml of double distilled water in a 400ml beaker for 30 minutes. The solution is then filtered through Whatman no.1 filter paper. The pH value of the extract is noted. The pure filtered plant extract is stored in a container and refrigerated at 4°C for further experimental work [20].

2.4 Phytochemical analysis
The results for the phytochemical analysis are shown below

![Table 1. Phytochemical analysis of the Pedalium murex leaf]

<table>
<thead>
<tr>
<th>Phytochemicals</th>
<th>Result</th>
</tr>
</thead>
<tbody>
<tr>
<td>Steroid</td>
<td>++</td>
</tr>
<tr>
<td>Tannin</td>
<td>++</td>
</tr>
<tr>
<td>Flavonoid</td>
<td>++</td>
</tr>
<tr>
<td>Phlobatinin</td>
<td>-</td>
</tr>
<tr>
<td>Terpenoids</td>
<td>++</td>
</tr>
<tr>
<td>Cardial glycosides</td>
<td>++</td>
</tr>
<tr>
<td>Reducing sugar</td>
<td>--</td>
</tr>
</tbody>
</table>
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Fig. 2.4 (a) Tests for the presence of phytochemicals in the plant extract

2.5 Synthesis of zinc doped iron oxide nanoparticles

For the synthesis of these nanoparticles, co-precipitation method is employed. The standard stock solutions of FeCl$_3$.7H$_2$O, FeSO$_4$.7H$_2$O and ZnCl$_2$.6H$_2$O are added in the ratio of 2:1:1. Then the mixture is allowed to boil for 15 minutes in a magnetic stirrer with constant stirring at 460 ppm. After 15 minutes, the pure plant extract is added to the mixture and continued to boil. Then the base (NH$_4$OH) is added to the mixture. The pH level is noted after each addition. The whole solution mixture is then allowed to boil for half an hour. The Nano solution is stored for UV-Vis and FTIR spectroscopic analysis.

Then the solution is centrifuged at 8000 rpm and the colloidal solution is retained. The colloidal solution obtained is then transferred to the petri plate and dried in a hot air oven at 250°C for 3 hours. The dried sample is again calcinated using muffle furnace.

2.6 Disc diffusion method

The standardized inoculums are inoculated in the plates prepared earlier (aseptically) by dipping a sterile in the inoculums removing the excess of inoculums by passing, pressing and rotating the swab firmly against the side of the culture tube above the level of the liquid and finally streaking the swab all over the surface of the medium 3 times rotating the plates through an angle of 60° c after each application. Finally pass the swab around the edge of the agar surface. Leave the inoculums to dry at room temperature with the lid closed. The petri dish is divided into parts, in each part, sample disc such as ZnIONPs(100µg) (discs are soaked overnight in sample solution) and standard Ciprofloxacin & Flucanazole (10µg),are placed in the plate with the help of sterile forceps. Then petri dishes are placed in the refrigerator at 4°C or at room temperature for one hour for diffusion. Incubate at 37°C for 24 hours. Observe the zone of inhibition produced by different samples. Measure it using a scale and record the average of two diameters of each zone of inhibition.

2.7 Pancreatic α-amy assay

The α-Amy inhibitory activity of zinc doped iron oxide nanoparticles was performed, according to our former publication [21].Briefly-amy (0.5mg/ml) was incubated with and without synthetic compounds for 10 minutes at 25 °c. This experiment was performed in 20mM sodium phosphate buffer, pH 6.9, containing 6mM sodium chloride. After pre-incubation, the starch solution was added and the reaction mixture was incubated for 30 minutes at 25°C.In order to stop the enzymatic reaction, dinitrosalicylic acid was added as the colour reagent and then incubated in a boiling water bath for minutes. After cooling down to the room temperature ,the reaction mixture was then diluted by adding distilled water and the absorbance measured at 540 nm on a T90+ UV-VIS spectrophotometer instrument [PG Instrument Ltd.,UK].The measured absorbance was compared with that of the control experiment and the obtained result were considered as criteria for the percentage of α-amy inhibition[21].In this study, the pharmacological inhibitor ,Acarbose was used as a positive control and the experiment were repeated for at least three times .

2.8 Uv-visible spectroscopy

Samples [1mL] of the suspension were collected periodically to monitor the completion of bio-reduction of iron in aqueous solution, followed by dilution of the samples with 2 ml of deionized water and subsequent scan in UV-visible [vis] spectra, between wave lengths of 200 to 700 nm in a spectrophotometer [Beckman - Model No. DU - 50, Fullerton], having a resolution of 1 nm.
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2.9 FTIR spectroscopy

FTIR analysis of the dried ZNIONPs was carried out through the potassium bromide (KBr) pellet (FTIR grade) method in 1:100 ratio and spectrum was recorded using Jasco FT/IR-6300 Fourier transform infrared spectrometer equipped with JASCO IRT-7000 Intron Infrared Microscope using transmittance mode operating at a resolution of 4 cm⁻¹.

2.10 XRD analysis

Crystallographic information about the samples was obtained from X-ray diffractometer (PAnalytical, Philips PW 1830) in the range of 20°-70° with 2°/min scanning rate, operating at 40 kV and a current of 30 mA with Cu Kα radiation (λ = 1.5404 Å) was used. The colloidal suspension containing metal nanoparticles was dried on a small glass slab.

III. Results and discussion

3.1 UV-visible spectroscopy

The surface Plasmon resonances (SPR) of synthesized zinc doped iron oxide nanoparticles have been studied by UV-Vis spectroscopy. After the addition of Pedalium murex leaf extract into the aqueous solution of iron chloride and iron sulphate, the solution was filled in glass cuvette of path length 10mm and UV-Vis spectral analysis has been done in the range of 300 to 700 nm. DI water was used as blank.

UV-Vis absorption spectral study may be assisted in understanding electronic structure of the optical band gap of the material. Absorption in the near ultraviolet region arises from electronic transitions associated within the sample.

![Fig.3.1(a) UV-Visible spectrum of plant extract (Pedalium murex)](image)

![Fig. 3.1(b) UV-Visible spectrum for zinc doped iron oxide nanoparticles using pedalium murex.](image)

The optical properties of synthesized iron oxide nanoparticles using UV visible spectroscopy were studied and the recorded spectra are shown in figure 3.1(a) and 3.1(b). The optical absorption of zinc doped iron oxide a nanoparticle of which is measured in a scanning range of wavelength from 300 to 800 nm.

The absorption band around at 509.25 nm by the optical absorption results [22], it is possible to determine band gap energy for the prepared sample.

3.1.1 Band gap calculation

The absorbance peak is related to the band gap energy, and hence using maximum absorbed wavelength, peak wavelength can be converted into band gap energy [22]. This can be converted using Einstein-plank’s relation: \[ E = \frac{hc}{\lambda} \]

\[ E = \frac{6.626 \times 10^{-34} \times 3 \times 10^8}{509.25} \]
3.2 FTIR Spectroscopy

The FTIR spectrum shows the presence of absorption bands associated with Fe-O stretching vibrations. These peaks represent the following bonding in the sample confirms the reducing agent role in the formation of iron oxide nanoparticles. The peak observed at 3424 cm\(^{-1}\) corresponds to the \(\text{–OH, –NH}\) stretching vibrations. The bands observed at 1623 cm\(^{-1}\), 1384 cm\(^{-1}\) are due to the C–O stretching C=O stretching mode of carbonyl functional groups in alcohol, ethers, acids and esters. The strong peaks at 548 cm\(^{-1}\), 465 cm\(^{-1}\) corresponds to the inorganic stretching indicates the Fe\(_2\)O\(_3\). This suggests that biological molecule could possibly perform dual function of formation and stabilization of iron oxide nanoparticles in the aqueous medium. These results implied that tannins, saponins, flavonoids, steroids, amino acids play a major role as a capping agent or reducing agent. The intensity of absorption band at 548 cm\(^{-1}\) is stronger than at 465 cm\(^{-1}\). It gives evidence for the formation of zinc doped \(\alpha\text{-Fe}_2\text{O}_3\) [23] and this is in agreement with the XRD measurement.

**Fig. 3.2** FTIR spectrum of zinc doped iron oxide nanoparticles

3.3 XRD Analysis

The XRD data is used in interpretation of grain size of crystal using Debye–Scherrer formula:

\[
D = \frac{0.9\lambda}{B \cos \Theta}
\]

where \(D\) is the particle size
\(\lambda\) is the wavelength of the incident x-ray beam
\(\Theta\) is the Bragg’s diffraction angle
\(B\) is full width at half maxima (FWHM) of the zinc doped iron oxide peak

The calculated size of the Nano crystallites with the Scherrer formula was 40.49 nm. The intense XRD patterns clearly showed the crystalline nature of the nanoparticles. The distinct peaks observed at 24.3150, 33.2925, 35.4381, 49.7152, 54.0576, 56.7577, 62.4739 accounting for crystal planes at (0,1,2), (1,0,4), (1,1,0), (0,2,4), (1,1,6), (2,1,1), (2,1,4) respectively. These planes represent the synthesized iron oxide nanoparticles are in hematite phase (\(\alpha\text{-Fe}_2\text{O}_3\)), rhombohedral in structure (JCPDS file no 89-8104).

**Fig. 3.3** XRD pattern for the zinc doped iron oxide nanoparticles
3.4 Anti-bacterial activity
The anti-bacterial activity was tested by using disc diffusion method against two bacterial species. Among them, one is gram-positive and the other is gram negative such as Staphylococcus aureus and Escherichia coli [24]. Ciprofloxacin is kept as the standard sample and the zone of inhibition is noted.

![Fig.3.4 Inhibition zone of Escherichia coli (a) and Staphylococcus aureus (b)](image)

<table>
<thead>
<tr>
<th>Organisms</th>
<th>Zone of inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Standard Ciprofloxacin [10µg/disc]</td>
</tr>
<tr>
<td>Staphylococcus aureus</td>
<td>32</td>
</tr>
<tr>
<td>Escherichia coli</td>
<td>48</td>
</tr>
</tbody>
</table>

3.5 Anti-fungal activity
The synthesized nanoparticles were tested against two fungal species i.e. Aspergillus niger and Candida albicans by disc diffusion assay [25]. Flucanazole, a commercial antifungal drug is kept as the standard sample. The zone of inhibition is noted.

![Fig.3.5 Inhibition zone of Candida albicans (a) and Aspergillus niger (b)](image)

<table>
<thead>
<tr>
<th>Organisms</th>
<th>Zone of inhibition[mm]</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Standard Flucanazole [10µg/disc]</td>
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<td></td>
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<table>
<thead>
<tr>
<th></th>
<th>Aspergillus niger</th>
<th>Candida albicans</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>11</td>
<td>10</td>
</tr>
<tr>
<td></td>
<td>10</td>
<td>12</td>
</tr>
</tbody>
</table>

3.6 Anti diabetic activity

3.6.1 The inhibition activity of zinc doped iron oxide nanoparticles against α-amy

The anti-diabetic activity was tested by using pancreatic α-amy assay method. Inhibition of α-amy is considered a useful strategy for the treatment of disorders in carbohydrate uptake, such as diabetes and obesity as well as the dental caries and periodontal disease[2]. Acarbose, a commercial ant diabetic drug is kept as the standard sample and then inhibitory activity of zinc doped iron oxide nanoparticle is noted.

![Fig. 3.6.1 (a) Inhibition zone of Acarbose](image1)

![Fig 3.6.1 (b) inhibition zone of blank (plant extract)](image2)

![Fig. 3.6.1 (c) Inhibition zone of ZnIONPs](image3)

Table – 4 Anti diabetic activity of ZnIONPs

<table>
<thead>
<tr>
<th>Concentration</th>
<th>Blank</th>
<th>Standard [Acarbose]</th>
<th>ZnIONPs</th>
</tr>
</thead>
<tbody>
<tr>
<td>20</td>
<td>10.12</td>
<td>17.68</td>
<td>13.46</td>
</tr>
<tr>
<td>50</td>
<td>15.35</td>
<td>29.32</td>
<td>23.42</td>
</tr>
<tr>
<td>100</td>
<td>25.76</td>
<td>50.42</td>
<td>36.56</td>
</tr>
<tr>
<td>150</td>
<td>42.23</td>
<td>65.66</td>
<td>60.42</td>
</tr>
<tr>
<td>200</td>
<td>63.33</td>
<td>89.98</td>
<td>78.87</td>
</tr>
</tbody>
</table>

IV. Conclusion

The present work showed that zinc doped iron oxide nanoparticles were synthesized by co-precipitation method using Pedalium murex as reducing agent. In UV-Vis Analysis, the peak observed at 509.25nm shows the formation iron oxide nanoparticles and the calculated band gap energy is 2.43eV. FTIR analysis shows that the peaks observed at 548cm\(^{-1}\) and 465cm\(^{-1}\) concludes the formation of zinc doped iron oxide nanoparticles. XRD analysis shows that the size of synthesized nanoparticles is 40.49nm, the prepared iron oxide nanoparticles are in hematite (α-Fe\(_2\)O\(_3\)) phase. The synthesized zinc doped iron oxide shows good antimicrobial and anti diabetic activity.
References


