Embryotoxic and Teratogenic Potentials of Salacia lehmbachii in Rats

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Abstract: Use of herbal remedies in pregnancy may be deleterious to both mother and fetus. Against this background, aqueous and ethanol root bark extracts of Salacia lehmbachii, widely used herbal remedies for treatment of malaria in some localities were evaluated for embryotoxicity and teratogenicity in albino rats. One hundred and five pregnant rats weighing 170-180g were assigned 7 groups (n=15), labeled Control, 1-6. Control rats had 2 mL of distilled water. Groups 1, 2 and 3 received 250, 500 and 750mg/kg body weight of aqueous extract respectively while groups 4, 5 and 6 had similar doses of ethanol extract. Administration was orally on days 1 to 6 of gestation for implantation studies and days 7 to 15 for teratogenicity. The rats were observed daily for lethality, abnormal behaviour and vaginal bleeding. Their weights, food and water intakes were recorded. Cesarean sections were performed on day 20 of gestation to remove their uterine horns and implantations, resorptions, live and dead fetuses recorded. The weight and crown rump lengths of live fetuses were obtained and the placentas weighed and examined along with the litters for anomalies. There were no signs of maternal toxicity, miscarriages and dead fetus. Number of live fetuses in treated rats were similar to control just like litter weights, crown rump lengths and placental weight. There were no external anomalies on the fetuses and placenta. The extracts at the doses used in the study are relatively safe for pregnant rats and developing fetuses.

Keywords: Embryotoxicity, pregnancy, teratogenicity, root bark, Salacia lehmbachii.

I. Introduction

The use of herbal remedies in primary health care continues to gain popularity especially in developing countries. Herbal medicines contain active ingredients of whole plants or parts thereof perceived to have therapeutic benefits [1]. Although these medicines are generally considered safe, in reality it is known that the active components may cause serious adverse effects especially as they are not subjected to rigorous scrutiny for safety as is the case with conventional medicines [2,3]. The risk of herb induced adverse effects is higher in people who are already prone to adverse reactions to conventional pharmacological preparations. Such persons include pregnant women, fetuses, children, lactating mothers and the elderly [4-7]. Pregnancy is associated with marked physiological alterations which perhaps make pregnant women more susceptible to certain diseases like malaria [8,9]. The use of medications during pregnancy may expose the developing fetus to risks like miscarriage, intrauterine growth retardation, malformations and mortality. In this respect, the historical saga of thalidomide, a sedative used in the treatment of hyperemesis gravidarum, but later found to be responsible for a string of birth defects readily comes to mind [10-13]. Many herbal remedies have been reported to impair fetal development following usage in pregnancy [14-22].

Salacia lehmbachii (Family, Celastraceae) is a popular medicinal plant used by locals in some parts of Southeastern Nigeria and Cameroun particularly the Bakassi forest reserves, for the treatment of febrile illnesses like malaria. Pregnant women are vulnerable to malaria which is endemic in these parts [23]. The plant is called ‘eba-enang-enang’ by the peoples of Akwa Ibom and Cross River States and ‘ara-mmnuu’by the Igbo, all of Nigeria. Earlier work from this department had shown the median lethal dose (LD₅₀) of the root bark extract of the plant in albino rats to be above 5000 mg/kg, while the chemical constituents were alkaloids, glycosides, flavonoids, tannins, saponins and polyphenols [24]. Biological effects of the plant include analgesia and antiinflammation [24], antipyresia [25], nephroprotection [26], anti-abortifacient action [27], antioxidant action [28], inhibition of male sex hormones [29] and antifertility in male rats [30]. The hepatotoxic and hematotoxic potentials of the plant have also been evaluated [31,32] and the antimalarial action of the plant has been established (Essien, AD, University of Calabar, Calabar, Nigeria, unpublished results). This study aimed to evaluate the embryotoxic and teratogenic potential of aqueous and ethanol extracts of the root bark of the plant in albino rats.

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II. Materials And Methods

2.1 Collection and identification of plant material
The roots of Salacia lehmbachii were purchased from a local market in Calabar, capital of Cross River State, Nigeria. The plant was authenticated by the department of Botany of the University of Calabar and a Specimen with Voucher number 688 deposited in the department for reference.

2.2 Preparation of the extract
The roots were washed with water to remove dirt and dried in their lengths in an electric oven, thermostatically controlled at 40°C, for 12 hours. The root bark powder of the plant was obtained and stored following an earlier described method [29]. The dry aqueous and ethanol extracts were derived from a two-staged Soxhlet extraction of the root bark powder following the methods of Essiet et al [31]. The solid extract which formed the yield was weighed and preserved in a clean and dry container until required for the experiments. The extracts were constituted in distilled water to give the doses required for the study.

2.3 Experimental animals
One hundred and five pregnant rats weighing 170-180g purchased from the Animal house, Department of Pharmacology, University of Calabar were used for this study. They were housed in well ventilated plastic cages, each containing fifteen rats. Each rat was properly identified by branding with dilute picric acid. The animals were acclimatized for seven days to normal laboratory conditions (relative humidity: 50±5%; temperature: 28±2°C and 12 hours of light-dark cycle) before the start of the experiment and maintained at the same conditions throughout the study period. They were fed with standard rat chow (Agro Feeds, Calabar) and water (Water board, Calabar) ad libitum. The guidelines on Care and Use of Laboratory animals were scrupulously observed [33].

2.4 Animal grouping and treatment
Sexually mature female rats were paired with their male counterparts and confined in a cage overnight for mating. The following day was designated day 1 of pregnancy if the dams had vaginal seminal plugs or spermatozoa were found in their vaginal smears on microscopic examination [34]. Pregnant rats were weighed and randomly assigned to 7 groups (n=15), labeled Control, 1-6. Control rats were administered 2 mL of distilled water, the vehicle for the extract. Groups 1, 2 and 3 received 250, 500 and 750mg/kg body weight of aqueous extract respectively while groups 4, 5 and 6 were treated with 250, 500 and 750mg/kg body weight of ethanol extract respectively. Administration was via a gastric cannula from day 1 to 6 of gestation for implantation studies[35] and day 7 to 15 of gestation for studies on organogenesis and possible teratogenicity [36,37].

To evaluate the extracts for maternal toxicity, the rats in each study group were observed daily for certain changes (lethality, abnormal behaviour, vaginal bleeding) and their weights taken on the 1st, 7th, 14th and 20th day of gestation using an electronic weighing balance. Food and water intake by the pregnant rats were monitored using the periodic manual recordings[38]. Laparotomy was performed under chloroform inhalation anaesthesia on the 20th day of gestation to remove the rat uteri [37]. The uterine horns were then opened up to count the numbers of fetuses (both live and dead), resorption sites and implantations. The ovaries were retrieved and the corpora lutea within them counted. Live fetuses were weighed, their crown-rump lengths measured after which they were examined for gross malformations. The placentas were equally weighed and examined for abnormalities.

The data obtained above were processed to obtain the following parameters namely [36-38]: i) Implantation index = total number of implantation sites / number of corpora lutea x 100, ii) Resorption index = total number of resorption sites / total number of implantation sites x100, iii) Preimplantation loss = number of corpora lutea – number of implantations / number of corpora lutea x100 and iv) Post implantation loss = number of implantations - number of live fetuses / number of implantations x100.

2.5 Statistical Analysis
The Statistical Package for Social Sciences (SPSS) version 20 was used to analyse data obtained from the study and values obtained from descriptive statistics expressed as means ± standard error of mean (SEM).

Turkey’s multiple comparison post hoc testing was carried out to compare the different study groups. Where applicable students t-test was used to compare differences between treated and control groups for significance at discreet dose levels. Differences were considered significant at P values of P< 0.05.
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III. Results
The rats gained comparable weight during the different periods of gestation assessed (TABLE 1). During the period of experimentation, there were no maternal deaths, no toxic clinical signs and the treated pregnant dams consumed as much food and water per day as control rats (TABLE 2). There was no abortion as the control and treated pregnant rats carried their pregnancies to term when they were sacrificed.

There were no resorption sites, resulting in a nil value of resorption index. Other implantation indices (implantation index, pre-implantation loss and post-implantation loss) were insignificantly (p>0.05) different in treated rats compared to control (TABLE 3). There was no dead fetus in both control and treated rats and the difference between the number of live fetuses delivered by treated rats and those delivered by control rats was not statistically significant (p>0.05). The fetal indices namely fetal weight, crown rump length and placental weight were dose dependently and insignificantly (p>0.05) increased in aqueous extract-treated rats compared to control. The above indices were reduced in rats treated with the ethanol extract and the reduction was insignificant (p<0.5) only with the highest dose of 750mg/kg (TABLE 4). There were no gross abnormalities in the fetuses and placentas of treated and control rats (TABLE 4).

<table>
<thead>
<tr>
<th>Study groups (n=15)</th>
<th>Maternal weight (g)/days of pregnancy</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Day 1</td>
</tr>
<tr>
<td>Control</td>
<td>175.44 ± 10.09</td>
</tr>
<tr>
<td>1 (250mg/kg of ASL)</td>
<td>177.12 ± 9.77</td>
</tr>
<tr>
<td>2 (500mg/kg of ASL)</td>
<td>172.72 ± 6.18</td>
</tr>
<tr>
<td>3 (750mg/kg of ASL)</td>
<td>169.22 ± 5.77</td>
</tr>
<tr>
<td>4 (500mg/kg of ESL)</td>
<td>179.12 ± 9.13</td>
</tr>
<tr>
<td>5 (500mg/kg of ESL)</td>
<td>176.22 ± 11.11</td>
</tr>
<tr>
<td>6 (750mg/kg of ESL)</td>
<td>173.14 ± 10.11</td>
</tr>
</tbody>
</table>

Values are means ± SEM. Percentage body weight changes are italicized and bracketed ( ).

ASL = Aqueous extract of Salacia lehmbachii root bark
ESL = Ethanol extract of Salacia lehmbachii root bark

<table>
<thead>
<tr>
<th>Study group (n=15)</th>
<th>Food intake (g/day/rat)</th>
<th>Water intake (ml/day/rat)</th>
<th>Physical sign of toxicity</th>
<th>Number of maternal deaths</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>17.76±1.11</td>
<td>25.10±3.02</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>1 (250mg/kg of ASL)</td>
<td>17.86±1.91</td>
<td>25.21±2.08</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>2 (500mg/kg of ASL)</td>
<td>18.69±2.23</td>
<td>25.19±1.00</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>3 (750mg/kg of ASL)</td>
<td>18.96±3.11</td>
<td>24.94±2.05</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>4 (250mg/kg of ESL)</td>
<td>17.77±1.06</td>
<td>25.09±2.11</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>5 (500mg/kg of ESL)</td>
<td>18.66±3.33</td>
<td>25.00±3.46</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>6 (750mg/kg of ESL)</td>
<td>19.01±1.09</td>
<td>25.24±1.87</td>
<td>0</td>
<td>0</td>
</tr>
</tbody>
</table>

Values are means ± SEM.

ASL = Aqueous extract of Salacia lehmbachii root bark
ESL = Ethanol extract of Salacia lehmbachii root bark

<table>
<thead>
<tr>
<th>Study group (n=15)</th>
<th>No of Implantation sites</th>
<th>No of resorption sites</th>
<th>No of corpora Lutea</th>
<th>Implantation index (%)</th>
<th>Resorption index (%)</th>
<th>Preimplantation loss (%)</th>
<th>Postimplantation loss (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>9.01±0.02</td>
<td>0.00±0.00</td>
<td>9.27±0.02</td>
<td>97.17±3.95</td>
<td>0.00±0.00</td>
<td>2.81±1.11</td>
<td>3.77±0.05</td>
</tr>
<tr>
<td>1 (250mg/kg of ASL)</td>
<td>9.02±0.01</td>
<td>0.00±0.00</td>
<td>9.28±0.02</td>
<td>97.20±4.01</td>
<td>0.00±0.00</td>
<td>2.80±1.18</td>
<td>3.76±0.08</td>
</tr>
<tr>
<td>2 (500mg/kg of ASL)</td>
<td>9.00±0.01</td>
<td>0.00±0.00</td>
<td>9.26±0.02</td>
<td>97.19±4.03</td>
<td>0.00±0.00</td>
<td>2.81±1.15</td>
<td>3.78±0.06</td>
</tr>
<tr>
<td>3 (750mg/kg of ASL)</td>
<td>9.01±0.02</td>
<td>0.00±0.00</td>
<td>9.27±0.02</td>
<td>97.20±3.97</td>
<td>0.00±0.00</td>
<td>2.81±1.22</td>
<td>3.77±0.05</td>
</tr>
<tr>
<td>4 (250mg/kg of ESL)</td>
<td>8.99±0.01</td>
<td>0.00±0.00</td>
<td>9.25±0.02</td>
<td>97.22±4.00</td>
<td>0.00±0.00</td>
<td>2.81±1.01</td>
<td>3.78±0.02</td>
</tr>
<tr>
<td>5 (500mg/kg of ESL)</td>
<td>9.00±0.01</td>
<td>0.00±0.00</td>
<td>9.26±0.02</td>
<td>97.19±3.83</td>
<td>0.00±0.00</td>
<td>2.81±0.89</td>
<td>3.78±0.06</td>
</tr>
<tr>
<td>6 (750mg/kg of ESL)</td>
<td>9.01±0.02</td>
<td>0.00±0.00</td>
<td>9.27±0.02</td>
<td>97.20±4.09</td>
<td>0.00±0.00</td>
<td>2.80±0.88</td>
<td>3.77±0.05</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SEM.  n = 15

ASL = Aqueous root bark extract of Salacia lehmbachii
ESL = Ethanol root bark extract of Salacia lehmbachii

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Table 4. Fetal indices in Control and Treated Rats

<table>
<thead>
<tr>
<th>Study group</th>
<th>No. of dead fetus</th>
<th>No. of live fetus</th>
<th>Fetal weight (g)</th>
<th>Crown rump length (cm)</th>
<th>Placental Weight (g)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control:</td>
<td>0</td>
<td>8.67±0.56</td>
<td>2.65±0.09</td>
<td>3.40±0.56</td>
<td>0.44±0.02</td>
</tr>
<tr>
<td>Control</td>
<td>0</td>
<td>8.67±0.56</td>
<td>2.63±0.09</td>
<td>3.40±0.56</td>
<td>0.44±0.02</td>
</tr>
<tr>
<td>1 (250mg/kg of ASL)</td>
<td>0</td>
<td>8.68±0.02</td>
<td>2.68±0.15</td>
<td>3.48±0.02</td>
<td>0.45±0.01</td>
</tr>
<tr>
<td>2 (500mg/kg of ASL)</td>
<td>0</td>
<td>8.66±0.01</td>
<td>2.79±0.23</td>
<td>3.52±0.01</td>
<td>0.48±0.01</td>
</tr>
<tr>
<td>3 (750mg/kg of ASL)</td>
<td>0</td>
<td>8.67±0.01</td>
<td>2.86±0.27</td>
<td>3.58±0.01</td>
<td>0.51±0.01</td>
</tr>
<tr>
<td>4 (250mg/kg of ESL)</td>
<td>0</td>
<td>8.65±0.42</td>
<td>2.64±0.16</td>
<td>3.41±0.03</td>
<td>0.44±0.01</td>
</tr>
<tr>
<td>5 (500mg/kg of ESL)</td>
<td>0</td>
<td>8.66±0.68</td>
<td>2.61±0.05</td>
<td>3.39±0.04</td>
<td>0.41±0.01</td>
</tr>
<tr>
<td>6 (750mg/kg of ESL)</td>
<td>0</td>
<td>8.67±0.53</td>
<td>2.28±0.03*</td>
<td>3.13±0.02*</td>
<td>0.26±0.02*</td>
</tr>
</tbody>
</table>

Values are expressed as mean ± SEM, n= 15
* Significant compared to control at p<0.05
ASL and ESL= Aqueous and ethanol root bark extracts of Salacia lehmbachii respectively

IV. Discussion

Fetal development is a highly organized process in which complex changes are coordinated sequentially in time, and changes at the molecular and cellular levels are integrated to enable manifestation of a particular phenotype in the whole organism. Maternal homeostasis contributes significantly to fetal development and is assessed using some maternal parameters (body weight changes, food and water intake) and clinical signs of toxicity (salivation, diarrhea, behavioural changes and vaginal bleeding) during pregnancy [35,39,40]. Usually, increase in maternal body weight accompanies pregnancy, so the observation of such increase in the dams in this study was an indication that they were actually pregnant. Ekhator and Shelu [41] while evaluating the abortifacent potentials of the unripe seed extract of Carica papaya used increase in maternal mean body weight of rats as a sign and progress of pregnancy. Since literature search shows no previous work on the effect of Salacia lehmbachii on pregnancy and fetal development, there are no results on the plant to compare with findings of this work. However, effects of Salacia reticulata, a specie in the genus, Salacia and some other herbs on pregnancy and fetal development have been evaluated [35, 40-46].

It has been established that a correlation exists between maternal toxicity and developmental toxicity [47]. The lack of maternal toxicity in this study as evidenced by absence of maternal deaths and clinical signs of toxicity as well as the similarity in food and water intake in all experimental groups suggests that the extracts do not cause developmental toxicity. Similar results were obtained from the root extract of S. Reticulata and extract of the aerial parts of Mentha piperita [42,43].

Implantation of the fertilized egg is a critical step of conception and the conception index correlates with the site of implantation and the number of corpora lutea [44]. The implantation index and pre-implantation loss rate evaluate blastocyst implantation in the uterus while the resorption index and post-implantation loss rate establish correlation between the number of implanted blastocysts and those that have not developed [35,41]. The absence of statistical difference in the implantation index, pre-implantation and post-implantation losses between treated and control rats as observed in this study is an indication that the numbers of blastocysts implanted were within normal range. The absence of abortions in the experimental groups agrees with the observed nil value of resorption index and suggests normal development of the implanted blastocysts. Normal implantation was also reported in pregnant rats treated with Carapa quianensis seed oil and aqueous stem extract of Bulbine natalensis Baker [48, 45]. On the contrary, there was impaired embryo development in rats treated with the root extract of S. reticulata because of post-implantation losses, and those given extracts of Asparagus racemosus and Solanum lycocarpum as a result of increased number of resorptions [42, 46, 44].

The weights and crown rump lengths of litters and placental weights are known indicators of fetal development [49]. Birth weight is thus a reflection of intrauterine fetal growth which is also determined by factors like the amount of nutrients from the mother and the transporting capacity of such nutrients to the fetus by the placenta. Inhibition of fetal growth in vivo may thus manifest as a reduction in mean fetal weight, crown-rump length, and placental weight [44]. As an earlier study had shown the ASL extract to stimulate erythrocytosis [32], the slight increase in fetal weight, crown rump length and placental weight observed in the rats treated with ASL may be speculated to be from a positive nutritional effect of the extract. Further studies will however be required to verify this position. A reduction in the fetal parameters as mentioned, at a high dose (750mg/kg) of ESL suggests a potential embryotoxic effect. Similar dose dependent embryotoxic potential was observed with the aqueous stem extract of Bulbine natalensis Baker [45]. Furthermore, the absence of physical abnormalities in the fetuses and placentas with both ASL and ESL indicates there is no teratogenicity from the extracts at the doses studied. Lack of teratogenic effects was also reported with the root extract of Salacia reticulata [42].

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V. Conclusion

Findings from this study show that the aqueous and ethanol extracts of root bark of *Salacia lehmbachii* at the doses used are not embryotoxic and do not have teratogenic action. However, usage of the ethanol extract at doses higher than 750mg/kg should be avoided in pregnancy as there was a reduction in fetal indices at that dose.

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