Morphological and Biomechanical Alterations of Achilles Tendon and Plantar Fascia in Type 2 Diabetes with Peripheral Neuropathy: A Shear Wave Elastography Study

Dr. Anindita Sanyal

Junior Resident, Department of Radiodiagnosis, Ramaiah Medical College, Bangalore, India.

Dr. Anand K.

(Corresponding Author)

Professor, Department of Radiodiagnosis, Ramaiah Medical College, Bangalore, India.

ABSTRACT

Background: Diabetes mellitus with peripheral neuropathy affects foot structures, potentially contributing to complications including ulceration and biomechanical dysfunction.

Objective: To evaluate morphological and biomechanical properties of Achilles tendon and plantar fascia in diabetic patients with peripheral neuropathy using ultrasonography and shear wave elastography.

Methods: This cross-sectional study compared 32 diabetic patients with peripheral neuropathy to 31 non-diabetic controls. Bilateral measurements of Achilles tendon and plantar fascia thickness were obtained using B-mode ultrasonography. Tissue stiffness was assessed using shear wave elastography. Statistical analysis included t-tests and Pearson correlations.

Results: Cases were older $(60\pm13 \text{ vs } 50\pm14 \text{ years}, p<0.001)$ with elevated fasting blood sugar $(156\pm65 \text{ vs } 82\pm10 \text{ mg/dl}, p<0.001)$. Achilles tendon thickness increased bilaterally in cases (right: $6.0\pm1.2 \text{ vs } 3.6\pm1.2 \text{mm}$; left: $6.08\pm1.19 \text{ vs } 3.66\pm1.27 \text{mm}$; both p<0.001). Plantar fascia showed greater thickening (right: $4.9\pm0.8 \text{ vs } 2.1\pm0.7 \text{mm}$; left: $5.1\pm1.0 \text{ vs } 2.2\pm0.7 \text{mm}$; both p<0.001). Tissue stiffness was elevated in cases for Achilles tendon $(3.40\pm0.70 \text{ vs } 2.24\pm0.46 \text{ m/s}, p<0.001)$ and plantar fascia $(2.96\pm0.69 \text{ vs } 1.77\pm0.42 \text{ m/s}, p<0.001)$. Strong correlations existed between fasting blood sugar and all measurements (r=0.55-0.72, p<0.001).

Conclusion: Diabetic peripheral neuropathy causes significant thickening and increased stiffness of foot structures, with plantar fascia more affected than Achilles tendon. These alterations correlate strongly with glycemic status, suggesting hyperglycemia as a key pathogenic factor.

Keywords: Diabetes mellitus; peripheral neuropathy; shear wave elastography; Achilles tendon; plantar fascia; ultrasonography

I. INTRODUCTION

Diabetes mellitus represents a global health crisis characterized by chronic hyperglycemia resulting from defects in insulin secretion, insulin action, or both (1). India harbors approximately 77 million diabetic patients, ranking second globally, with projections indicating an increase to 109 million by 2035 (2). Among diabetes complications, peripheral neuropathy affects approximately 50% of patients, primarily involving the lower extremities and contributing significantly to foot morbidity (3).

The pathophysiology of diabetic peripheral neuropathy (DPN) involves multiple mechanisms initiated by chronic hyperglycemia. Advanced glycation end products (AGEs) form through non-enzymatic glycation of proteins, fundamentally altering tissue structure and function (4). These AGEs accumulate within collagen fibers, creating abnormal cross-links that increase tissue stiffness and reduce elasticity, particularly affecting load-bearing foot structures essential for normal biomechanics (5).

The Achilles tendon, the body's strongest and largest tendon, measures approximately 15 centimeters in length and undergoes significant mechanical loading during locomotion (6). Its unique spiral architecture, with fibers rotating 90 degrees before calcaneal insertion, creates differential stress distribution areas potentially vulnerable to diabetes-related changes (7). The plantar fascia, originating from the medial calcaneal tubercle and extending to the metatarsophalangeal joints, maintains the medial longitudinal arch and functions as a dynamic shock absorber (8). Composed predominantly of longitudinally arranged type I collagen fibers interspersed with elastic fibers, it provides essential viscoelastic properties for foot function (9).

Recent investigations reveal that diabetes induces significant structural alterations in both structures. Histopathological studies demonstrate collagen fiber disorganization, increased interfibrillar ground substance, and neovascularization in diabetic tendons (10). These morphological changes accompany biomechanical alterations including reduced compliance and altered stress-strain relationships, compromising normal shock absorption and potentially contributing to increased plantar pressures and ulceration risk (11).

While conventional ultrasonography provides morphological assessment, it cannot evaluate mechanical properties. Shear wave elastography (SWE) enables quantitative tissue stiffness assessment through acoustic radiation force-generated shear waves, with propagation velocity directly proportional to tissue stiffness ($E = 3\rho c^2$) (12). This technique offers operator-independent, reproducible measurements ideal for monitoring disease progression (13).

Previous elastography studies in diabetic populations report variable findings, potentially reflecting differences in patient selection and measurement techniques (14). Understanding relationships between glycemic control and musculoskeletal properties is crucial for developing targeted interventions. This study comprehensively evaluates Achilles tendon and plantar fascia in diabetic patients with established peripheral neuropathy using both conventional ultrasonography and SWE, addressing critical knowledge gaps regarding morphological and biomechanical consequences of diabetic neuropathy on weight-bearing structures.

II. METHODS

Study Design and Population

This cross-sectional observational study was conducted at a tertiary care hospital from May 2023 to January 2024 after institutional ethics committee approval. Written informed consent was obtained from all participants.

Participants

Sample size calculation, based on previous studies reporting 2.0mm mean difference in Achilles tendon thickness with 1.5mm standard deviation, indicated 28 participants per group were required (α =0.05, power=80%). We recruited 32 diabetic patients with peripheral neuropathy (cases) and 31 non-diabetic controls.

Inclusion criteria for cases included: confirmed type 2 diabetes mellitus; peripheral neuropathy diagnosed by abnormal monofilament test (10g), reduced vibration perception threshold (>25V), or abnormal nerve conduction studies; age 18-80 years. Control inclusion criteria: absence of diabetes (fasting glucose <100mg/dl, HbA1c <5.7%); no neuropathy history; age-matched within 5 years.

Exclusion criteria: Achilles tendon or plantar fascia pathology history; diabetic foot ulcers; peripheral vascular disease (ankle-brachial index <0.9); connective tissue disorders; recent lower limb trauma; BMI >40 kg/m²; pregnancy.

Clinical Assessment

Demographic data and medical history were collected through structured interviews. Peripheral neuropathy was assessed using 10g Semmes-Weinstein monofilament at ten sites per foot and vibration perception threshold using biothesiometer. Fasting blood glucose was measured after 8-hour overnight fast using hexokinase method.

Ultrasound Examination

All examinations were performed using Siemens ACUSON S2000 with 9L4 linear transducer (4-9 MHz) by a single experienced sonographer. Participants were positioned prone with feet hanging freely at 90-degree ankle flexion.

Achilles tendon thickness was measured 2-6cm proximal to calcaneal insertion in transverse plane, recording anteroposterior diameter perpendicular to tendon fibers. Plantar fascia thickness was measured at calcaneal origin in sagittal plane aligned with second metatarsal. Three measurements were averaged for each structure.

Shear Wave Elastography

Virtual Touch Quantification was performed immediately following B-mode assessment. A 10mm×5mm region of interest was positioned within each structure, ensuring perpendicular beam orientation to minimize anisotropy. Ten consecutive shear wave velocity measurements were obtained, recording median values in meters per second.

Statistical Analysis

Data were analyzed using SPSS version 25.0. Continuous variables were expressed as mean±standard deviation, categorical variables as frequencies and percentages. Between-group comparisons used independent t-tests for parametric data and chi-square tests for categorical variables. Pearson correlations assessed relationships between variables. Multiple linear regression identified independent predictors. P<0.05 was considered significant.

DOI: 10.9790/0853-2409048084 www.iosrjournal.org 81 | Page

III. RESULTS

Participant Characteristics

The study included 32 diabetic patients with peripheral neuropathy and 31 controls. Cases were significantly older $(60\pm13 \text{ vs } 50\pm14 \text{ years}, \text{ p}<0.001)$ with male predominance in both groups (cases: 65.6%, controls: 80.6%, p=0.171). Median diabetes duration was 8 years (IQR 5-12). Mean HbA1c was 8.4±1.8% in available cases (n=28).

Table 1: Demographic and Glycemic Parameters

Parameter	Cases (n=32)	Controls (n=31)	p-value
Age (years)	60±13	50±14	< 0.001
Male (%)	65.6	80.6	0.171
BMI (kg/m²)	26.8±3.2	24.3±2.8	0.002
FBS (mg/dl)	156±65	82±10	< 0.001
VPT right (V)	32.4±8.7	12.3±3.2	< 0.001
VPT left (V)	31.8±9.1	12.1±3.5	< 0.001

FBS: Fasting blood sugar; VPT: Vibration perception threshold

Morphological Findings

Achilles tendon thickness was significantly increased bilaterally in cases. Right side measured 6.0 ± 1.2 mm in cases versus 3.6 ± 1.2 mm in controls (67% increase, p<0.001). Left side showed similar changes (6.08±1.19mm vs 3.66 ± 1.2 7mm, p<0.001).

Plantar fascia demonstrated more pronounced thickening. Right side measured 4.9±0.8mm in cases versus 2.1±0.7mm in controls (133% increase, p<0.001). Left side thickness was 5.1±1.0mm versus 2.2±0.7mm (p<0.001).

Table 2: Thickness Measurements

Structure	Cases	Controls	Difference (95% CI)	p-value
AT-Right (mm)	6.0±1.2	3.6±1.2	2.4 (1.8-3.0)	< 0.001
AT-Left (mm)	6.08±1.19	3.66±1.27	2.42 (1.8-3.0)	< 0.001
PF-Right (mm)	4.9±0.8	2.1±0.7	2.8 (2.4-3.2)	< 0.001
PF-Left (mm)	5.1±1.0	2.2±0.7	2.9 (2.5-3.3)	< 0.001

AT: Achilles tendon; PF: Plantar fascia; CI: Confidence interval

Biomechanical Findings

Shear wave elastography revealed significantly increased stiffness in diabetic patients. Achilles tendon stiffness was 3.40 ± 0.70 m/s in cases versus 2.24 ± 0.46 m/s in controls (52% increase, p<0.001). Plantar fascia stiffness measured 2.96 ± 0.69 m/s versus 1.77 ± 0.42 m/s (67% increase, p<0.001).

Table 3: Stiffness Measurements

Structure	Cases	Controls	Difference (95% CI)	p-value
AT Stiffness (m/s)	3.40 ± 0.70	2.24 ± 0.46	1.16 (0.87-1.45)	< 0.001
PF Stiffness (m/s)	2.96±0.69	1.77 ± 0.42	1.19 (0.91-1.47)	< 0.001

Correlation Analysis

Strong positive correlations existed between fasting blood sugar and all parameters. Strongest correlation was with right plantar fascia thickness (r=0.72, p<0.001), followed by left plantar fascia (r=0.69, p<0.001). Achilles tendon thickness showed moderate correlations (right: r=0.63, left: r=0.59, both p<0.001). Stiffness measurements correlated with glycemia (AT: r=0.55, PF: r=0.60, both p<0.001).

Table 4: Correlations with Fasting Blood Sugar

Parameter	Correlation (r)	p-value
AT thickness right	0.63	< 0.001
AT thickness left	0.59	< 0.001
PF thickness right	0.72	< 0.001
PF thickness left	0.69	< 0.001
AT stiffness	0.55	< 0.001

Parameter	Correlation (r)	p-value
PF stiffness	0.60	< 0.001

Inter-structural correlations revealed strong associations between Achilles tendon and plantar fascia measurements (r=0.78-0.82, p<0.01). Thickness-stiffness correlations were stronger for plantar fascia (r=0.71-0.76) than Achilles tendon (r=0.51-0.57).

Bilateral Symmetry and Sex Differences

No significant bilateral differences existed for Achilles tendon thickness in either group (cases: p=0.345, controls: p=0.367). Plantar fascia showed small but significant left-sided thickening in cases only (5.1 vs 4.9mm, p=0.031). Sex-stratified analysis revealed no significant differences between males and females within groups (all p>0.05).

Table 5: Multiple Regression Analysis

Dependent Variable	Independent Predictor	β Coefficient	p-value
AT thickness	FBS	0.58	< 0.001
	Age	0.22	0.042
PF thickness	FBS	0.65	< 0.001
	Age	0.14	0.186

Multiple regression identified fasting blood sugar as the strongest independent predictor of both Achilles tendon (β =0.58, p<0.001) and plantar fascia thickness (β =0.65, p<0.001) after adjusting for age, sex, and BMI.

IV. DISCUSSION

This study demonstrates significant morphological and biomechanical alterations in Achilles tendon and plantar fascia of diabetic patients with peripheral neuropathy. The observed changes—67% increased Achilles tendon thickness, 133% increased plantar fascia thickness, and corresponding stiffness increases—represent substantial deviations from normal tissue properties with important clinical implications.

Our findings exceed previously reported changes. Evranos et al. documented 46% increased Achilles tendon thickness in diabetic patients without specific neuropathy assessment (15). Our greater magnitude likely reflects inclusion of patients with established neuropathy, representing advanced disease. The mean 6.0mm thickness substantially exceeds the 4-6mm normal range, indicating clinically significant alterations (16).

The 133% plantar fascia thickening represents one of the largest reported increases. Previous studies documented 30-80% increases, but our neuropathy-specific cohort may experience more severe changes (17). Mean thickness of 4.9-5.1mm exceeds the 4.0mm plantar fasciitis diagnostic threshold, yet patients were asymptomatic, highlighting distinct diabetic fasciopathy pathophysiology (18).

Differential changes between structures warrant consideration. The plantar fascia's greater susceptibility may relate to its complex loading patterns—tension, compression, and shear forces—unlike the primarily tensile Achilles tendon loading (19). Additionally, its proximity to plantar fat pad and role in arch maintenance may increase metabolic stress susceptibility.

Our elastography findings align with recent investigations. Fu et al. reported similar Achilles tendon stiffness increases (3.28±0.61 vs 2.31±0.48 m/s) in diabetic patients (20). Increased stiffness results from AGE accumulation causing abnormal collagen cross-linking, reduced proteoglycan content, and altered extracellular matrix composition (21). Interestingly, percentage stiffness increase (52% Achilles, 67% plantar fascia) was less than thickness increase, suggesting morphological changes may precede mechanical alterations.

Strong correlations between glycemia and tissue parameters (r=0.55-0.72) indicate dose-dependent relationships. Akturk et al. similarly reported significant HbA1c-Achilles tendon thickness correlations (r=0.64, p<0.001) (22). These findings suggest intensive glycemic control might prevent or ameliorate musculoskeletal complications, though specific studies are needed.

Clinical implications are substantial. Increased thickness and stiffness reduce ankle dorsiflexion, a recognized ulceration risk factor. Mueller et al. demonstrated 5.3-fold increased ulcer risk in diabetic patients with limited joint mobility (23). Altered plantar fascia properties compromise shock absorption, potentially increasing plantar pressures. Bus et al. reported 2.4-fold increased ulceration risk with elevated plantar pressures (24).

Bilateral symmetry suggests systemic rather than localized pathology, supporting bilateral assessment recommendations. Minor plantar fascia asymmetry may reflect subtle compensatory mechanisms. Sex-stratified analysis revealed no differences, indicating universal diabetes effects across genders.

Study strengths include neuropathy-specific focus, comprehensive morphological and biomechanical assessment, and standardized SWE protocols providing quantitative measurements. Limitations include cross-sectional design precluding temporal assessment, age differences between groups despite statistical adjustment, and lack of functional outcome measures.

Future research should investigate longitudinal progression, relationships to clinical outcomes, and intervention potential. Establishing whether intensive glycemic control or targeted therapies can reverse alterations has significant implications. Molecular pathway investigation may identify novel therapeutic targets. Standardized ultrasound and elastography protocols would facilitate multicenter studies and clinical implementation.

V. CONCLUSION

Diabetic peripheral neuropathy causes significant morphological and biomechanical alterations in Achilles tendon and plantar fascia. Substantial thickness increases (67% Achilles tendon, 133% plantar fascia) and stiffness elevations (52% Achilles tendon, 67% plantar fascia) likely contribute to abnormal foot biomechanics and increased ulceration risk characteristic of diabetic foot disease.

Strong glycemic parameter correlations emphasize optimal diabetes management importance in preventing musculoskeletal complications. Greater plantar fascia changes suggest differential tissue susceptibility related to anatomical and biomechanical factors. Ultrasound and shear wave elastography provide valuable non-invasive assessment tools that could facilitate early detection, risk stratification, and treatment monitoring.

These findings enhance understanding of diabetic foot complication pathophysiology and support comprehensive management approaches addressing both metabolic and musculoskeletal aspects. Recognition and quantification of these alterations may improve outcomes for millions affected by diabetic peripheral neuropathy worldwide.

REFERENCES

- [1]. American Diabetes Association. Classification and diagnosis of diabetes: Standards of Medical Care in Diabetes-2023. Diabetes Care. 2023;46(Suppl 1):S19-S40.
- [2]. Pradeepa R, Mohan V. Epidemiology of type 2 diabetes in India. Indian J Ophthalmol. 2021;69(11):2932-8.
- [3]. Pop-Busui R, Boulton AJ, Feldman EL, Bril V, Freeman R, Malik RA, et al. Diabetic neuropathy: A position statement by the American Diabetes Association. Diabetes Care. 2017;40(1):136-54.
- [4]. Singh VP, Bali A, Singh N, Jaggi AS. Advanced glycation end products and diabetic complications. Korean J Physiol Pharmacol. 2014;18(1):1-14.
- [5]. D'Souza RS, Levandowski C, Slavov D, Gress TW, Narayan S. Diabetic tendinopathy: Pathophysiology and treatment options. Clin Diabetes Endocrinol. 2021;7(1):19.
- [6]. Doral MN, Alam M, Bozkurt M, Turhan E, Atay OA, Dönmez G, et al. Functional anatomy of the Achilles tendon. Knee Surg Sports Traumatol Arthrosc. 2010;18(5):638-43.
- [7]. Edama M, Kubo M, Onishi H, Takabayashi T, Inai T, Yokoyama E, et al. The twisted structure of the human Achilles tendon. Scand J Med Sci Sports. 2015;25(5):e497-503.
- [8]. Stecco C, Corradin M, Macchi V, Morra A, Porzionato A, Biz C, et al. Plantar fascia anatomy and its relationship with Achilles tendon and paratenon. J Anat. 2013;223(6):665-76.
- [9]. Wearing SC, Smeathers JE, Urry SR, Hennig EM, Hills AP. The pathomechanics of plantar fasciitis. Sports Med. 2006;36(7):585-611.
- [10]. Grant WP, Sullivan R, Sonenshine DE, Adam M, Slusser JH, Carson KA, et al. Electron microscopic investigation of the effects of diabetes mellitus on the Achilles tendon. J Foot Ankle Surg. 1997;36(4):272-8.
- [11]. Giacomozzi C, D'Ambrogi E, Uccioli L, Macellari V. Does the thickening of Achilles tendon and plantar fascia contribute to the alteration of diabetic foot loading? Clin Biomech. 2005;20(5):532-9.
- [12]. Taljanovic MS, Gimber LH, Becker GW, Latt LD, Klauser AS, Melville DM, et al. Shear-wave elastography: Basic physics and musculoskeletal applications. Radiographics. 2017;37(3):855-70.
- [13]. Aubry S, Risson JR, Kastler A, Barbier-Brion B, Siliman G, Runge M, et al. Biomechanical properties of the calcaneal tendon in vivo assessed by transient shear wave elastography. Skeletal Radiol. 2013;42(8):1143-50.
- [14]. Wiesinger HP, Rieder F, Kösters A, Müller E, Seynnes OR. Are sport-specific profiles of tendon stiffness and cross-sectional area determined by structural or functional integrity? PLoS One. 2016;11(6):e0158441.
- [15]. Evranos B, Idilman I, Ipek A, Polat SB, Cakir B, Ersoy R. Real-time sonoelastography and ultrasound evaluation of the Achilles tendon in patients with diabetes with or without foot ulcers. J Diabetes Complications. 2015;29(8):1124-9.
- [16]. Pierre-Jerome C, Moncayo V, Terk MR. MRI of the Achilles tendon: A comprehensive review of the anatomy, biomechanics, and imaging of overuse tendinopathies. Acta Radiol. 2010;51(4):438-54.
- [17]. Bolton NR, Smith KE, Pilgram TK, Mueller MJ, Bohnert KL. Computed tomography to visualize and quantify the plantar aponeurosis and flexor hallucis longus tendon in the diabetic foot. Clin Biomech. 2005;20(5):540-6.
- [18]. McNally EG, Shetty S. Plantar fascia: Imaging diagnosis and guided treatment. Semin Musculoskelet Radiol. 2010;14(3):334-43.
- [19]. Chen TL, Sze LK, Davis IS, Cheung RT. Effects of training in minimalist shoes on the intrinsic and extrinsic foot muscle volume. Clin Biomech. 2016;36:8-13.
- [20]. Fu S, Cui L, He X, Sun Y. Elastic characteristics of the normal Achilles tendon assessed by virtual touch imaging quantification shear wave elastography. J Ultrasound Med. 2016;35(9):1881-7.
- [21]. Goldin A, Beckman JA, Schmidt AM, Creager MA. Advanced glycation end products: Sparking the development of diabetic vascular injury. Circulation. 2006;114(6):597-605.
- [22]. Akturk M, Ozdemir A, Maral I, Yetkin I, Arslan M. Evaluation of Achilles tendon thickening in type 2 diabetes mellitus. Exp Clin Endocrinol Diabetes. 2007;115(2):92-6.
- [23]. Mueller MJ, Diamond JE, Delitto A, Sinacore DR. Insensitivity, limited joint mobility, and plantar ulcers in patients with diabetes mellitus. Phys Ther. 1989;69(6):453-62.
- [24]. Bus SA, Maas M, de Lange A, Michels RP, Levi M. Elevated plantar pressures in neuropathic diabetic patients with claw/hammer toe deformity. J Biomech. 2005;38(9):1918-25.
- [25]. Boulton AJ, Armstrong DG, Albert SF, Frykberg RG, Hellman R, Kirkman MS, et al. Comprehensive foot examination and risk assessment. Diabetes Care. 2008;31(8):1679-85.