

A Study on Prevalence and Antibiotic Susceptibility Pattern of Vancomycin Intermediate and Resistant Staphylococcus Aureus in Clinical Specimen in a Tertiary Care Hospital and Detection of their MIC Values by E-test.

Dr. Prasanna Gupta¹, Dr. Rahul Bhargava²

1 (Professor and Head Microbiology department, National Institute of Medical Science, NIMS, India)

2 (Microbiology, National Institute of Medical Science, NIMS, India)

Corresponding Author: Dr. rahul bhargava

Abstract: With the increasing incidence of Methicillin Resistant Staphylococcus Aureus (MRSA), Vancomycin Intermediate Staphylococcus Aureus (VISA) & Vancomycin Resistant Staphylococcus Aureus (VRSA) strains now a days. The study was conducted to find out the magnitude of vancomycin resistance in both MRSA and MSSA and antibiotic susceptibility pattern of those isolates in a tertiary care hospital, Jaipur between January 2017 to June 2018. In this cross sectional study, 287 Staphylococcus aureus were isolated and identified from various clinical specimens collected from different departments of the hospital. Among these 287 strains, 146 were found to be MRSA and 141 MSSA. Subsequently, the antimicrobial susceptibility test was performed by Kirby Bauer disc diffusion method as per CLSI guidelines. All strains found resistant to vancomycin by disc diffusion method were again grown on BHI-VSA (Brain heart infusion-Vancomycin screen agar) and also recruited to E-Test for confirmation of resistance. Minimum Inhibitory Concentration (MIC) of ≤ 2 were considered as VSSA, 4-8 as VISA. MIC of ≥ 16 as VRSA. Vancomycin resistance was seen in 9 isolates of S. aureus by disc diffusion method. Among them neither was found to be VISA nor VRSA when confirmed with E-Test with all having MIC < 2 . Though there was no incidence of VRSA but MIC of 1.5 $\mu\text{g/ml}$ in our study rang an alarm to the infection control committee of this tertiary care hospital.

Keywords: MIC, S. aureus, VISA, VRSA, Vancomycin Screen Agar

Date of Submission: 05-12-2018

Date of acceptance: 21-12-2018

I. Introduction

Staphylococcus aureus is one of the most notorious and important human pathogen that has long been recognized for its ability to cause serious and invasive diseases. Staphylococcus aureus is the most pathogenic member of Staphylococcus genus. This microbe is ubiquitous in nature which resides in the human as a commensally. Some of the commoner infection caused by Staph. aureus include Pyogenic infection (eg. Folliculitis impetigo, breast abscess, post operative wound infection, osteomyelitis, septic arthritis, bronchopneumonia, lungs abscess, empyema). Disseminated infection such as Septicemia. Toxic shock syndrome (TSS), staphylococcal scalded skin syndrome (SSSS), staphylococcal food poisoning.^[1] A wide range of antibiotics are used to treat the staphylococcal infections including penicillin, cephalosporin, macrolide, fluoroquinolone and glycopeptide group of antibiotics.^[3] In early 90s, the major treatment available to combat this organism was Penicillin and all isolates were sensitive to penicillin. In the 1960s, a new semisynthetic penicillinase-resistant antimicrobial drugs methicillin (formerly named as celbenine) and oxacillin were developed to treat staphylococcal infections caused by β -lactamase-production.^[4] But at present, some S. aureus strains also show resistant to methicillin.^[2]

Methicillin-resistant Staphylococcus aureus (MRSA)^[5] has been recognized as one of the major pathogens in both hospital and community settings. The first case of MRSA was isolated way back in 1961. Since then, there has been an escalating rate of infections caused by MRSA worldwide resulting in increased mortality and morbidity statistics. In India, the prevalence of nosocomial infections caused by MRSA varies between 20 and 40%. Vancomycin, a glycopeptides antibiotics had been considered to be the "gold standard antibiotic" and the drug of choice over the last 3 decades.^[6]

Apart from vancomycin, other effective drugs as linezolid and teticolanin are also widely used. Injudicious and infrequent use of vancomycin even in methicillin sensitive Staphylococcal infection has resulted in emergence of the strains with higher vancomycin MIC.^[7] At present few vancomycin resistant S. aureus strains have been reported. Initially in early twenties, only VISA (Vancomycin intermediate S. aureus) strains

were reported by CDC but even now in India, there is emergence of *S.aureus* strain with higher vancomycin MIC.

This higher vancomycin MIC has been attributed by increased thickness of cell wall as in case of Vancomycin Intermediate *Staphylococcus aureus*.^[8] Vancomycin resistance is acquired by mutation and thickening of cell wall due to accumulation of excess amounts of peptidoglycan.^[9]

Keeping the above points in view and also that documented reports of VISA and VRSA in India including Rajasthan are very few, the present study was planned to find out the prevalence of vancomycin intermediate *Staphylococcus aureus* and vancomycin resistant *Staphylococcus aureus* among isolates of *Staphylococcus aureus* in various clinical specimens along with their antibiotic sensitivity pattern so as to guide the clinicians of our hospital to select appropriate antimicrobial agents and also to make them aware, that if inappropriate use of vancomycin is continued it may lead to impending public health disaster. Therefore the present study is designed to find out the prevalence of vancomycin intermediate and resistant *Staphylococcus aureus* in clinical specimen and to determine the antibiotic susceptibility pattern of vancomycin intermediate and resistant *Staphylococcus aureus* and to measure the MIC values of vancomycin intermediate and resistant *Staphylococcus aureus* by E-test.

II. Material And Methods

The present study was conducted in the Department of Microbiology, NIMS Jaipur during the year 2017-2018. A total of 287 non- duplicate *Staphylococcus aureus* isolates from various clinical specimens [pus, wound or vaginal swabs, blood, body fluids (csf, pleural fluid, ascitic fluid) urine, sputum, ET secretion etc. were included in the study. Isolates from both in-patients and out-patients were considered. Institutional Ethical clearance was obtained. Data regarding age, sex, etc was obtained from the requisition form submitted to microbiology Department, NIMS, Jaipur.

2.1 Isolation & Identification, of *Staphylococcus aureus*^[10,11]

Smears were prepared from pus, wound swab, sediments of body fluids and respiratory samples, stained by Gram staining and examined for presence of inflammatory cells, epithelial cells and the type of microbial flora. As soon the samples received in the laboratory, Streak culture method was employed for sample inoculation on Blood agar, Nutrient agar. Culture plates were incubated at 37° C aerobically for 24-48 hours. Plates were observed for typical colony characteristics of *Staphylococcus aureus* on Blood agar (β haemolysis) and Nutrient agar (yellow pigment). For plates with mixed culture growth, sub culture was done on blood agar and incubated for 18-24 hours aerobically at 37°C. Sub culture was also done for *Staphylococci* colonies more than 48 hours old, where ever required. Gram's staining was performed from representative colonies, and observed for Gram positive cocci in clusters, under oil immersion microscope. Catalase test was performed from Nutrient agar plate. This test is positive for *Staphylococcus aureus* and differentiates from *Streptococci*. Slide coagulase test was employed to detect presence of clumping factor (bound coagulase) which was indicated by prompt clumping in positive isolates. Irrespective of result of slide coagulase test, tube coagulase test was done for presence of coagulase enzyme (Free coagulase). *Staphylococcus aureus* is positive. For all *Staphylococcus aureus* strains mannitol fermentation was observed on Mannitol salt agar.

2.2 Antibiotic Sensitivity testing of *Staphylococcus aureus*^[12]

Antibiotic susceptibility to a panel of drugs was tested using (Modified Kirby Bauer's method. Antibiotic discs procured commercially [Hi-media Laboratories, Mumbai] and were placed on inoculated MHA plates using forceps. Plates were incubated at 35±2PC for 18-24 hours. Zone of inhibition of all the antibiotics were measured with scale in reflected light against a black background, to the nearest mm. Interpretation was done according to the guidelines of Clinical Laboratory Standards Institute (2012). All *Staphylococcus aureus* isolates were subjected to predetermined panel of antibiotics which includes cefoxitin[30ug], Penicillin [10µgm], Cotrimoxazole [1.5/23.75µgm], Erythromycin[15µgm], Ciprofloxacin [5µgm], Clindamycin [2 µgm], Gentamycin [10 µgm], Levofloxacin [5 µgm], Tetracycline [30 µgm], Vancomycin [30 µgm]. *Staphylococcus aureus* ATCC 25923 was used as control. Cefoxitin disc (30 µgm) was used along with other antibiotics, to detect methicillin resistant isolates as it is a potent inducer of *mec A* gene mediated resistance^[12].

2.3 Detection of Vancomycin resistance

Following 3 methods were used and comparison between modified Kirby -Bauer disc diffusion method using 30µg Vancomycin disc and BHI vancomycin screen agar (6 µg/ml) was done keeping in view E-test as gold standard for detection of vancomycin resistance.

2.3.1 Disc diffusion by Modified Kirby Bauer's Method^[13]

It was performed using Vancomycin 30µg disc. The diameter of-zone of inhibition was measured and interpreted according to CLSI guidelines 2007^[13]. B.BHI vancomycin screen agar^[12] BHI-VSA (Hi-Media, India) plates containing 6µg/ml vancomycin were prepared. Inoculum suspensions were prepared by selecting colonies from subcultured colonies on nutrient agar plates. The colonies were transferred to sterile saline to produce a suspension that matches the turbidity of a 0.5 McFarland standard. Using a micropipette, spot of 10-µL drop onto agar surface was done and were incubated aerobically at 35±2°C for 24 hrs^[14]. Any growth is examined carefully with transmitted light.

2.3.2 Determination of Minimum Inhibitory Concentration (MIC) values^[15]

The MIC value of vancomycin was determined by E-test [Epsilometer-test]. Inoculum suspensions were prepared by selecting colonies from overnight growth on nutrient agar plates. The colonies were transferred to sterile saline to produce a suspension that matches the turbidity of a 0.5 McFarland standard and a lawn culture was prepared by pouring the growth suspension on the surface of the BHI agar plate. After drying the surface for half an hour, the E-strips were placed over the surface and incubated over night at 35°C. The plates were read only when sufficient growth was seen and the MIC values were recorded where the ellipse intersects the MIC scale on the strip. If the ellipse intersects the strip in between 2 dilutions MIC was recorded as the value which is nearest to the intersection. For classifying isolates of *Staphylococcus aureus* with reduced susceptibility to vancomycin based on the laboratory breakpoint published by the clinical and laboratory standards institute [CLSI guidelines]^[12]

Vancomycin sensitive *Staphylococcus aureus* [VSSA]: ≤2 µg/ml.

Vancomycin intermediate *Staphylococcus aureus* [VISA]: 4-8 µg/ml.

Vancomycin resistant *Staphylococcus aureus* [VRSA]: ≥16 µg/ml.

III. Observation And Results

During the study period, a total of 287 non duplicate *Staphylococcus aureus* isolates were obtained from various clinical specimens. Methicillin resistance was detected in 146 (51%) of the total isolates. Isolates that were sensitive to methicillin were 141 (49%) in 287 strains.

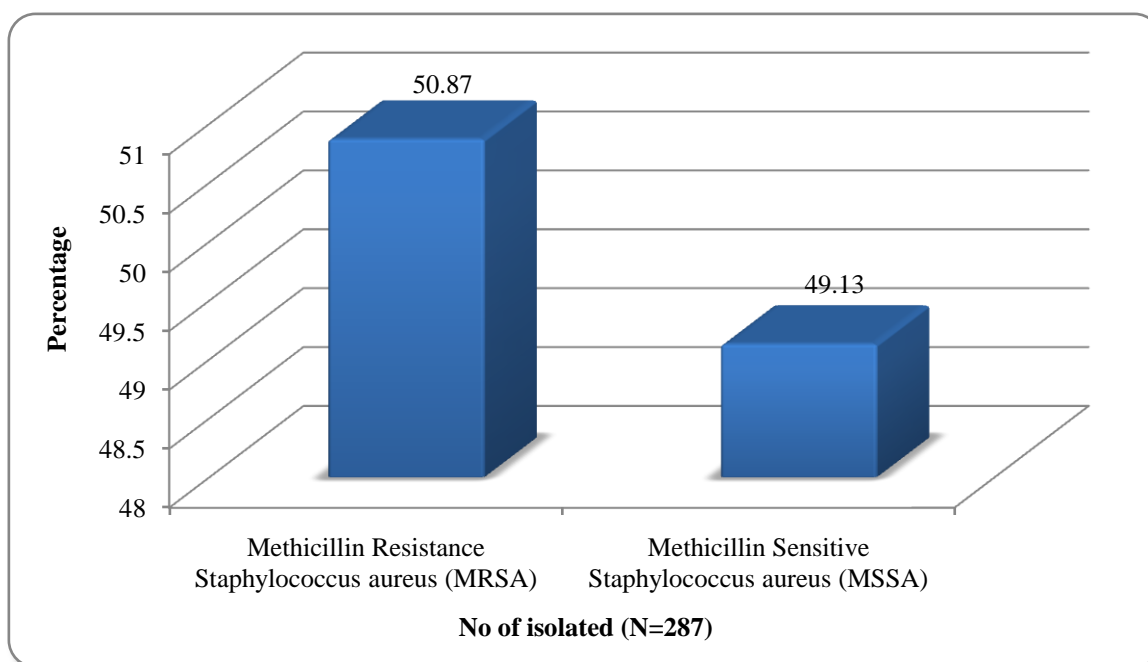


Fig. 1: Prevalence of MRSA and MSSA in total of 287 *Staphylococcus aureus* isolates

3.1 Vancomycin Resistance

Three phenotypic methods employed in detection of Vancomycin resistance,

3.1.1 Disc Diffusion method

In a total of 287 *S.aureus* stains, 9 (3%) isolates had shown resistance towards Vancomycin in Disc diffusion.

3.1.2 BHI Vancomycin screen agar (6 ugtn/ ml)

None of the isolates grew on So no resistance is reported by this method.

3.1.3 Epsilometer [E-test] for Vancomycin

It was considered as Gold Standard. MIC values of all the 287 isolates fall in between range of $\leq 2\mu\text{g}$ /ml, which is category of sensitive. All of the 9 isolates, which were resistant by Vancomycin disc diffusion, are in range of sensitive MIC.

Table No 1: Detection of Vancomycin resistance by different phenotypic methods

| Methods | VISA | VRSA | VSSA | Total |
|--|------|------|------|-------|
| Vancomycin disc diffusion (30 ug/disc) | NA | 9 | 278 | 287 |
| Vancomycin Screen Agar(6ug/ml) | 00 | 00 | 287 | 287 |
| Vancomycin E Test | 00 | 00 | 287 | 287 |

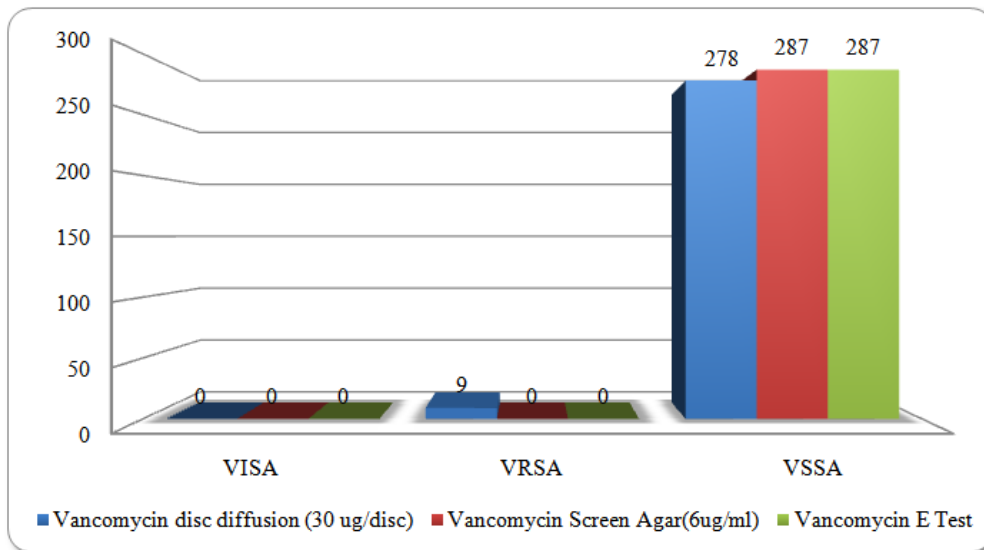


Fig. 2: Detection of Vancomycin resistance by different phenotypic methods

Table No 2: Distribution of the cases according to MIC Values (ugm/ml) against vancomycin

| MIC Values (ugm/ml) | Total | |
|---------------------|-------|--------|
| | No | % |
| 0-0.5 | 169 | 58.89 |
| >0.5-1 | 112 | 39.02 |
| >1-1.5 | 6 | 2.09 |
| Total | 287 | 100.00 |

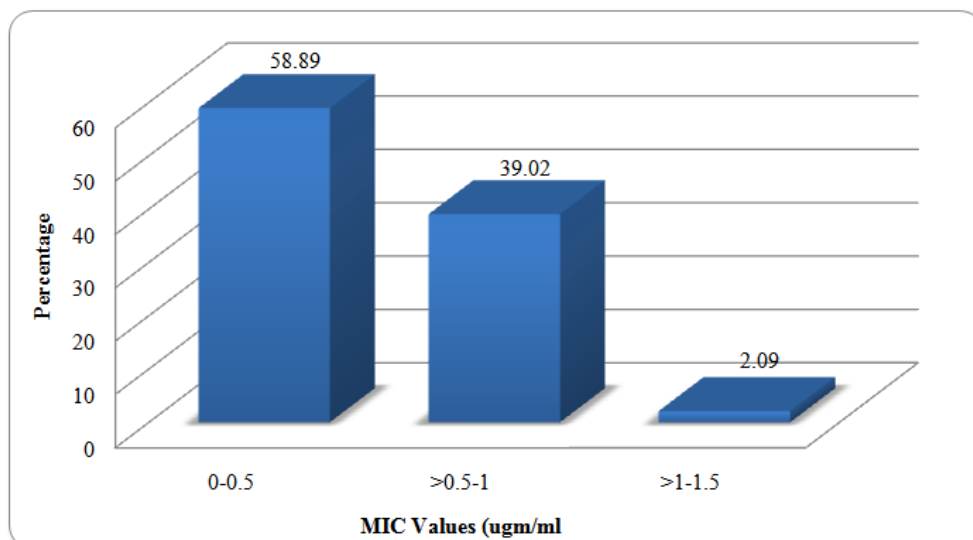


Fig.3: Distribution of the cases according to MIC Values (ugm/ml) against vancomycin

3.2 Antibiogram

All 287 *Staphylococcus aureus* strains were tested for antibiotic susceptibility pattern against a panel of predefined antibiotics, which was described earlier.

Staphylococcus aureus showed 88% resistance to Penicillin G, followed by 51% resistance to Cefoxitin, 43% each to Co-trimazole and Erythromycin, 66% to Ciprofloxacin, 31% to Clindamycin, 16% to Gentamycin and 28% to Levofloxacin and 3% each for both Tetracycline and Vancomycin.

IV. Discussion

Staphylococcus aureus is a major human pathogen and is one of the commonest causative agent of Community and Hospital acquired infections^[11]. The treatment of *Staphylococcus aureus* infection has become problematic because of emergence of resistance to Penicillin, Methicillin, Vancomycin and many other antibiotics, by acquiring several resistance mechanisms. Increased antimicrobial resistance for such an organism is, therefore a cause of concern.

In the past few decades MRSA has emerged as an important nosocomial pathogen worldwide. In India, prevalence rate varies from 30-85% in different parts and has now become endemic.^[16,17,18] A multicentric study done in India involving 17 tertiary care Hospitals reported MRSA prevalence of 41% in 2008-2009.

MRSA is of serious therapeutic concern not only due to its resistance to Methicillin, but also because of resistance to many other antimicrobials that are used on regular basis in Hospitals. Therefore, the most reliable and sustained therapeutic agent against methicillin-resistant *Staphylococcus aureus* (MRSA) strains is Vancomycin^[19]. There is still controversy in clinicians regarding the outcome of Vancomycin treatment in MRSA. Increasing prevalence of MRSA, lead to the extensive use of vancomycin. This in turn lead to the decreased susceptibility to Vancomycin all over the World including India, this was soon followed by strains of *Staphylococcus aureus* that were totally resistant to vancomycin^[20,21]. Such resistance resulted in serious clinical and public health consequences because currently few licenced alternatives are available. to treat vancomycin resistant *Staphylococcus aureus* infections.

Thus the present study was undertaken to determine the current status of Vancomycin susceptibility in our Hospital setup. to compare the Vancomycin disc diffusion test with BHI- VSA [6µg/ml] for detecting Vancomycin resistance considering E-Test as gold standard and also to determine the antibiotic susceptibility pattern of these isolates.

In the present study a total of 287 non-duplicate staphylococcus strains were isolated from various clinical specimens. Among all these samples highest isolation was from pus 175 (61%). Harcharan Singh et al in Udaipur (65%)^[22], Manu Chaudhary et al in H.P (63%)^[23] and Ankur Goyal et al in Agra (66.03%)^[24], also reported the highest isolation of *Staphylococcus aureus* from pus.

In our study 146 (51%) isolates turned out to be MRSA and 141 (49%) as MSSA. from a total of 287 *Staphylococcus aureus* strains. The prevalence rate of MRSA in our institute is 51%, which is similar to the studies conducted by S.Vidhani and P.L. Mehndiratta et al in 2001^[25] showing a prevalence rate of 51.6% and almost comparable -to the study conducted by Majumdar et al in 2001^[26] and Assadullah et al in 2003^[27] showing 52.9% prevalence rate. The higher rate in their studies may be attributed to the fact that the studies were conducted at a tertiary care multispecialty center with more and more patients coming from periphery and small nursing homes, where injudicious use of antibiotics and inadequate infection control policies are prevalent.

In the present study, maximum MRSA were isolated from pus 89(61%), followed by blood 22(15%), respiratory secretions 15(10%), Swabs and body fluids 7(5%) each and least from Urine 6 (4%). This pattern correlates with studies conducted by Vidya Pai et al in 2010^[28] and Nitish Kumar Sharma et al 2013^[18]. This is due to the reason that *Staphylococcus aureus* accounts for most of the skin and soft tissue infections, septicemia and also respiratory tract infection.

Comparatively MRSA prevalence was more in males. (71%) than in females (29%) in our study: Similar findings was also reported by Rao BN et al^[29] and Abhishek Mewar et al^[30]. The increased rate of MRSA infections among males could be due to their more outdoor activities, in turn exposing them to contaminated environment and also compared to females, accidental injuries are more common among men.

Most of the MRSA strains were isolated from 21-30 yrs of age group (ie 23.69%) and in 31-40yrs (ie 17.77%), indicating MRSA infection is more common in working and old age group. The reason for this may be that younger age group are more involved in outdoor activities in turn exposing them to contaminated environment and in older age group it may be due to waning immunity, hormonal abnormalities and co-morbid conditions. Similar pattern of affected age group. was also reported by Ankur Goyal et al in 2013^[24].

In the present study all 287 *Staphylococcus aureus* strains were screened for vancomycin resistance by Vancomycin disc diffusion method [30g/disc] and BHI-Vancomycin screen agar [6µg/ml]. These were further confirmed by Vancomycin E-test. All 287 *Staphylococcus aureus* isolates had MIC values $\leq 2\mu\text{g/ml}$, hence all were sensitive to Vancomycin and were labeled as VSSA according to the CLSI guidelines 2012^[12]. Among

these 169(59%) isolates had MIC of < 0.5µg/ml, 112[39%] had MIC of >0.5-1 µg /ml. Only 6[2%] had MIC of >1-1.5 µg /ml and none of the isolates had MIC values of >1.5 µg /ml.

In our study no VISA. and VRSA found. This may be due to the fact that the community acquired MRSA (CA-MRSA) unlike the hospital acquired MRSA (HA-MRSA) are known to be sensitive to drugs other than vancomycin. Because of its. high cost, vancomycin may not be in use in the peripheral rural setups, thus decreasing the selection pressure for vancomycin resistance^[31].

The current study only indicates the tip of iceberg. More and more studies should be undertaken in future to 'monitor the emergence of resistance to these. antibiotics. This also necessitates to find out better treatment policies and also to use cheaper and effective alternative anti - MRSA drugs so as to reduce the antibiotic pressure on vancomycin. Also clinicians should continue to exercise caution in their use of vancomycin in order to preserve this useful antibiotic and prolong its therapeutic usefulness.

V. Conclusion

To conclude, the result of our present study indicated high antibiotic resistance in commonly used antibiotics by MRSA isolates. The increased use of vancomycin drug has worsened the sensitivity.

We should undertake more and more such studies in future to fight against rising menace of antibiotic resistance, Also more research should be done to find better treatment policies, effective and cheaper alternative antibiotics in developing countries like ours. Even the clinical microbiology laboratories must ensure using detection methods with good sensitivity and specificity. We should also undertake more studies to find out the accurate screening method for VISA and VRSA. The findings of the studies should be shared with hospital infection control committee to help in the formulation of infection control polices and also antibiotic policies. So that the primary care givers can use antibiotics rationally.

However, *S. aureus* with reduced susceptibility was not observed in the present study. So, as a precautionary measure before starting the patient on vancomycin, the clinicians should seek the help of Clinical Microbiologist to determine the MIC of such strains so that emergence of vancomycin resistance can be prevented. All the laboratories should routinely test the MIC of vancomycin for all *Staphylococcus aureus* infection for appropriate treatment of patients and also for implementation of infection control.

Acknowledgement

We are thankful to Dr. Suman Rishi and Dr. Anjali for their guidance and help.

References

- [1]. Koneman E, Procop G, Schreckenberger P et al. Taxonomy of Staphylococci and related Gram Positive Cocci, clinical significance of Staphylococci and related Gram Positive Cocci. Koneman's Colour Atlas and Text book of practical Microbiology; 6th Edn USA 2007;624-642.
- [2]. Mackie and McCartney. In: Practical Medical Microbiology 14th edition, South Asia: Churchill Livingstone Elsevier. 2006;246.
- [3]. Dhawan B, Gadepalli R, Rao C, Kapil A, Sreenivas V. Decreased Susceptibility to Vancomycin in Methicillin-Resistant *Staphylococcus aureus*: A 5 year study in an Indian tertiary hospital. Journal of Medical Microbiology. 2010; 59: 375-376.
- [4]. Oliveria DC, Tomasz A, de Lencastre H. Secrets of success of a human pathogen: molecular evolution of pandemic clones of methicillin-resistant *Staphylococcus aureus*. Lancet Infect Dis 2002;2:180-9. Barber M. 1961. methicillin-resistant staphylococci. J Clin Pathol 14:385-393.
- [5]. Priya datta, Neelam Gulati, Nidhi Singla, Hena Rani Vasdeva, Kiran Bala, Jagdish Chander and Varsha Gupta. Evaluation of various methods for the detection of Methicillin-resistant *Staphylococcus aureus* strains and susceptibility patterns. Department of Microbiology, Government Medical College Hospital, Chandigarh, India, Journal of Medical Microbiology (2011),60,1613-1616.
- [6]. Peppard WJ, Daniels A, Fehrenbacher L, Winner J. Evidence based approach to the treatment of community -associated methicillin -resistant *Staphylococcus aureus*. Infect Drug Resist, 2009;2:27-40.
- [7]. Hiramatsu K. Vancomycin -resistant *Staphylococcus aureus*: a new model of antibiotic resistance. lancet Infectious Diseases. 2001;1(3):147-155.
- [8]. Cui, L., X. Ma, K. Sato, K. Okuma, F.C. Tenover, E.M. Mamizuka, C.G. Gemmell, M.N. Kim, M.C. Poly, N. El -solh, V. Ferraz, and K. Hiramatsu. Cell wall thickening is a common feature of vancomycin resistance in staphylococcus aureus. Journal of Clinical Microbiology. 2003;41:5-14.
- [9]. Edmond MB, Wenzel RP, Pasculle AW. Vancomycin- resistance in staphylococcus aureus: perspectives on measures needed for control. Annals of Internal Medicines. 1996;124:329-334.
- [10]. Baird. *Staphylococcus*: Cluster-forming gram-positive cocci. In: Collee JG, Fraser AG, Marmion BP, Simmons A, editors. Mackie and McCartney Practical Medical microbiology. 14th edn. Edinburgh: Churchill Livingstone. 1996:245-261.
- [11]. Koneman Elmer, Winn Washington, Allen Satphen, Procop Gary editors. Color Atlas & Textbook of Diagnostic Microbiology, 6th edition. 2006:643 -648.
- [12]. Clinical and Laboratory Standards Institute [CLSI]. Performance Standards for Antimicrobial Susceptibility Testing. Twenty-second - Informational Supplement. M100-S22. 2012; 32(1).
- [13]. Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing 17th informational supplement M100-SI7, CLSI, Wayne, PA, 2007.
- [14]. Kuusela P, Hilden P, Savolainen K et al. Rapid detection of methicillin-resistant *Staphylococcus aureus* strains not identified by slide agglutination tests. J Clin Microbiol 1994; 32: 143-47.
- [15]. Centers for Disease Control and Prevention. Investigation and Control of Vancomycin-Intermediate and - Resistant *Staphylococcus aureus* (VISA/ VRSA). A Guide for Health Departments and Infection Control Personnel. Updated on August 6 2012.

- [16]. Priyanka Chauhan, Prabhakar S. Bais and Nidhi Gupta et al. Prevalence of Methicillin resistant Staphylococcus aureus (mac A gene) among the patients admitted in Intensive care Unit. *Int. J. Bioassays*, 2013; 02 (09), 1256-1259.
- [17]. Hafiz S, Hafiz AN, Ali L, Chughtai AS, Memon B: Methicillin resistant Staphylococcus aureus: a multicentre study. *JPMA* 2002; 52:312.
- [18]. Nitish K S, Raina G, Shrikala B and Gopalkrishna B K: Nosocomial Infections and Drug susceptibility Patterns in Methicillin Sensitive and Methicillin Resistant staphylococcus aureus. *Clin Diagn Res.* 2013;7:2178-2180.
- [19]. HW Boucher, GR Corey, *Clin Infect Dis.* 2008,46,5,344-349.
- [20]. Centers for Disease Control and Prevention. Staphylococcus aureus resistant to vancomycin-United States, 2002. *Morb Mortal Wkly Rep MMWR.* 2002;51: 565-567.
- [21]. Tiwari HK, Sen MR et al. Emergence of vancomycin resistant Staphylococcus aureus (VRSA) from a tertiary care hospital from northern part of India. *infect Dis.* 2006;6:156.
- [22]. Harcharan Singh, Meena Atray, and Pankaj Kumar Modi et al. Antibiotic susceptibility pattern of Methicillin resistance Staphylococcus aureus in tertiary care center at Southern Rajasthan. *IJPSR*, 2014; 5(2): 607-611.
- [23]. Manu Chaudhary and Anurag Payasi. Prevalence of Icterogencous- Glycopeptide intermediate resistance in Methicillin resistant Staphylococcus aureus. *American Journal of Infectious' diseases.* 2013;9(3):63-70.
- [24]. Ankur Goyal, Manish Kumar Diwakar, Suneel Bhooshan, Sapna Goyal, Arti Agrawai, et al. Prevalence and Antimicrobial Susceptibility Pattern of Methicillin-resistant Staphylococcus aureus [MRSA] isolates at a Tertiary Care Hospital in Agra, North India - A systemic annual review. *Journal of Dental and Medical Sciences (IOSR-JDMS).* 2013;1 1(6): 80-84.
- [25]. Vidhani S, Mathur MD, Mehndiratta PL, Rizvi M. Methicillin resistant Staphylococcus aureus: the associated risk factors. *Indian J Pathol Microbiol* 2003;46(4):676-679.
- [26]. Majumder D, Samoa Bordoloi jN, Phukan AC, et al. Antimicrobial susceptibility pattern among methicillin resistant Staphylococcus isolates in Assam. *Ind. 3. Med. Microbiol.* 2001; 19(3):21-27.
- [27]. Assadullah S, Kakru DK, Thoker MA, Bhat FA, Hussain N, Shah A et al. Emergence of low level vancomycin resistance in MRSA *Indian J Med Microbiol.* 2003; 21.196-198.
- [28]. Vidya Pai, Venkatakrishna I Rao, Sunil P Rao. Prevalence and Antimicrobial Susceptibility Pattern of Methicillin resistant Staphylococcus Aureus MRSA Isolates at a Tertiary Care Hospital in Mangalore, South India. *Journal of Laboratory Physicians* 2010;2(2):82-4.
- [29]. Bandaru Narasinga Rao, Srinivas B. A prospective study of Methicillin resistant Staphylococcus aureus [MRSA] in a teaching Hospital of Rural setup. *Journal of Pharmaceutical and scientific innovation*, March -April 2012: 37-40.
- [30]. Mewara A, Gautam V, Kaur H, Ray P. In vitro evaluation of antibiotics for methicillin – resistant staphylococcus aureus from north India. *Indian J Med Res.* 2014;139:319-22.
- [31]. Dhanalakshmi T.A, Umopathy B.L, Mohan D.R, et al. Prevalence of Methicillin, Vancomycin and Nlultidrug Resistance among Staphylococcus aureus. *Journal of Clinical and Diagnostic Research.* 2012 August;6(6): 974-977.

Dr. Prassana Gupta. "A Study on Prevalence and Antibiotic Susceptibility Pattern of Vancomycin Intermediate and Resistant Staphylococcus Aureus in Clinical Specimen in a Tertiary Care Hospital and Detection of their MIC Values by E-test." *IOSR Journal of Dental and Medical Sciences (IOSR-JDMS)*, vol. 17, no. 12, 2018, pp 21-27.