**Cyclin B1 Protein Expression in Oral Dysplasia and in Oral Squamous Cell Carcinoma – “An immunohistochemical study”**.

Dr. Santosh Kumar Kotnoor MDS, Dr. Bhalkešwar,
Dr. Rohini V Junjunappanavar MDS, Dr. Gurucharan Donekāl,
Dr. Pavankichade, Dr. Shivraj S Patil, Dr. Abhishek Bansal MDS

1. Senior Lecturer, Dept Of Oral Pathology, Hyderabad Karnataka education Trust Dental College, Humnabad, Karnataka.
2. Senior Lecturer, Dept of Conservative and Endodontics, HKDET’s Dental College, Hospital and Research Institute, Humnabad, Karnataka.
3. Private Practitioner
4. PG Student, Dept. Of Oral Surgery, Al-Badar Dental College and Hospital, Gulbarga.
5. PG Student, Dept. Of Oral Surgery, Al-Badar Dental College and Hospital, Gulbarga.
6. PG Student, Dept. Of Oral Surgery, Al-Badar Dental College and Hospital, Gulbarga.
7. Senior Lecturer, Dept of Orthodontics and Dentofacial Orthopedics, Vaidik Dental College And Research Center, Nani Daman, U.T.

**Abstract:** Cyclin B1 is a member of the cyclin family of protein whose level vary during the cell cycle in order to activate specific cyclin-dependent kinases (CDKs) required for the proper progression through the cell cycle. The present study is to compare the expression of cyclin B1 in normal, dysplastic and squamous cell carcinomas.

**Experimental design:** To determine the role of cyclin B1 in normal, dysplastic and squamous cell carcinomas, we analysed 60 patients with normal, dysplastic and squamous cell carcinomas.

**Results:** In control patients that is of twenty patients, six were negative for cyclin B1 expression in basal and parabasal layers and 14 were positive for cyclin B1 expression in basal and parabasal layers. In other group (dysplastic) of twenty patients, 3 were negative and 17 were positive for cyclin B1 expression. In other group with squamous cell carcinomas, 7 were negative and 13 were positive for cyclin B1 expression.

**Conclusion:** Our study concludes that although the expression of cyclin B1 in dysplastic epithelium is below the cut of point (15%) but the expression was considerably high and this indicates that the patients with dysplastic are more susceptible for further changes, including progression and development of carcinomas.

**I. Introduction**

Carcinogenesis is literally the origination of cancer. It is a process by which normal cells are transformed into malignant cells, characterized by a progression of changes at the cellular, genetic and epigenetic level that ultimately reprogram a cell to undergo uncontrolled division, thus leading to formation of a malignant mass. Recent insights in the field of cell cycle regulation and cancer have provided prime examples of research at the “frontiers of science”.

The eukaryotic cell cycle is divided into four stages; G1, S, G2, and M. G1 is a gap phase during which cells prepare for the process of DNA replication. It is during the G1 phase that the cell integrates mitogenic and growth inhibitory signals and makes the decision to proceed, pause, or exit the cell cycle. S phase is defined as the stage in which DNA synthesis occurs. G2 is the second gap phase during which the cell prepares for the process of division. M stands for mitosis, the phase in which the replicated chromosomes are regulated into separate nuclei and cytokinesis occurs to form two daughter cells. G0 term is used to describe cells that have excised the cell cycle and become quiescent.

Cyclins A, D and E regulates the passage from G1 phase to S phase, where as cyclins A and B direct the transition from G2 phase to M phase. Specifically, cyclinB1 and cdc2 are the components of the maturation/mitosis-promoting factor, which plays an important role from G2 – M phase transition. Cyclin B1 binds to cyclin-dependent kinase2, which then becomes dephosphorylated and relocated to the nucleus.

Expression of cyclin B1 is cyclic with a minimal expression in G1 phase, an increased level in S phase, and peak at the G2-M transition. However, activecyclin B1 / (Cyclin-dependent kinase2 complex has been detected in the cytoplasm and is implicated in the formation of mitotic spindle. Thus Cyclin B1 through its trafficking between the cytoplasm and the nucleus, seems to play a role in coordinating the mitotic process in both components.
Cyclin B1 is overexpressed in a variety of cancers compared to normal cells and tissues. In normal tissues, low levels of cyclin B1 are detected in testis, thymus, bone marrow, and smooth muscle (CCNB1 expression).

Overexpression of cyclin B1 has been reported in lung, breast, colon, prostate, and head and neck cancers. Oral habits such as alcohol consumption and tobacco chewing are considered to be initiators of dysplastic changes in the oral mucosa. The expression of cyclin B1 in clinically normal oral mucosa, and in dysplastic changes has not been studied. The present study is to compare the expression of cyclin B1 in normal, dysplastic, and squamous cell carcinomas, so to fill the void in this important field.

II. Materials And Methods

Study population.

The patients for the present study are drawn from the outpatient department of S. Nijalingappa institute of Dental Sciences and Research and Basaweshwar Teaching and General Hospital, Gulbarga.

Patient Selection

The patients selected for the present study were with the following parameters.

Group I: Twenty patients without tobacco chewing or smoking habits and with clinically normal oral mucosa, served as a control group.

Group II: Twenty patients with tobacco chewing or smoking habits. Oral biopsy was done and stained with H and E staining to ascertain, the status of epithelial cells.

Group III: Twenty patients with the habit of tobacco chewing with clinical and histopathologically confirmed cases of oral squamous cell carcinoma.

The tissue biopsies of all three groups of the patients were stained with haematoxylin and eosin and subsequently subjected to immunohistochemical staining. Paraffin embedded, 3μm thick tissue section from all 60 patients were stained for cyclin B1 using primary mouse monoclonal antibody (CGN51) (Santa Cruse United Kingdom). The samples were rehydrated in graded alcohols. To retrieve the antigenicity, the tissue section were treated with pressure cooker using EDTA at pH 8. The pressure cooker was allowed to reach to full pressure (15 psi) and then incubated for 2 minutes. To block Endogenous peroxidase the tissue were covered with peroxide block reagent according to tissue size. Then it is incubated for 10 minutes at room temperature in humid chamber. Section was incubated overnight at 4°C with primary anticyclin B1 at a 1:15 dilution. The sections were processed using standard avidin-biotin immunohistochemistry according to the manufacturers recommendation. DAB – 3, 3′-diaminobenzidinechromozen (HK124 – 5k) was used as a chromo Zen& commercial hematoxylin was used for counter staining. The recommended positive control tissue for this antibody was thyroid carcinoma tissue.

The cyclin B1 labeling index was defined as the percentage of tumor cells displaying cytoplasmic or nuclear immune-reactivity and was calculated by counting the number of cyclin B1 stained tumor cells ≥ 1000 cells from representative areas of each tissue section we used 15% labeling index as a cutoff point. Cells were counted in ≥ 4 fields (at ×100) in these areas. All slides were scored concomitantly by two investigators.

III. Results and Observations

The expression of cyclin B1 in oral mucosal biopsies of different category of patients was assessed by immunohistochemical method by using mouse monoclonal antibody to cyclin B1 protein. The different groups of patients include Group(I) patients with clinically normal mucosa, without the habit of either tobacco chewing or smoking.

Group (II) patients were tobacco chewers and/or smokers

Group (III) patients included were histopathologically confirmed oral squamous cell carcinoma cases with habit of tobacco chewing and smoking.

Cyclin B1 expression in various groups of patients (Among 1000 cell counted more than four fields) in %.

In group I, (control) patients, that is of twenty patients, six were negative for cyclin B1 expression in the basal and parabasal layers.

In group II, three patients were negative for cyclin B1 and seventeen patients were positive for cyclin B1 expression.

In group III, seven patients were negative for cyclin B1 expression and thirteen were positive for cyclin B1 expression. Comparison of cyclin B1 expression in different category of patients (Table II, III, IV, IX) Comparison between group I and group II (Table 3) using student T test, T value was 6.90 (p<1.96 for p=0.05). Comparison between group I and group III (Table 3) using student T test, T value was 5.13 (p<1.96 for p=0.05).
Comparison between group II and group III (Table 3) using student T test, T value was 2.61 (> 1.96 for p = 0.05).

Moderately Differentiated oral Squamous
Well Differentiated oral Squamous cell Carcinoma cell (H & E stain X40)
Carcinoma (H & E stain X40)
A group of biologically similar cancers that start in the lips, mouth (oral cavity), nose (Nasal cavity), Paranasal sinuses, pharynx, and larynx are referred to as head and neck cancer. In head and neck region, squamous cell carcinoma is the most frequent malignant tumor and account for 90% of all malignancies. The role of exogenous agents in the etiology of head and neck cancers are tobacco smoking, alcohol intake, oral snuff, dietary factors, occupational exposures, and oral hygiene.

Cigarette smoke contains at least 3500 unidentified chemical constituents, of which nearly 40% being carcinogenic like polycyclic aromatic hydrocarbons (PAH), such as benzo(a)pyrene (Bap), and 4(α-methyl nitrosamine)-(3-pyridil)-1-butanone (NNK). Relatively fewer carcinogens are present in smokeless tobacco. About 30 carcinogens have been identified, the major contributor being nicotine derived nitrosamines (TSNA). Smokeless tobacco is often combined with other reagents including betel leaf, sliced areca nut and powdered slake lime. These additives used not only enhance the psychotropic effects of nicotine but also make the combination more genotoxic than the tobacco alone. The accompanying use of tobacco and alcohol contributes to increased incidence of head and neck cancers.

The process by which normal cells are transformed into cancer cell is known as carcinogenesis or oncogenesis or tumorigenesis. It is characterized by a progression of changes at the cellular, genetic and epigenetic level that ultimately reprogram a cell to undergo uncontrolled cell division, thus forming a malignant mass. Eukaryotic cell cycle is divided into four stages, G1, S, G2 and M. G1 - the cell prepares for the process of division. S- DNA synthesis occurs. G2 - the cell prepares for the process of division. M (mitosis) - the replicated chromosome are segregated into two separate nuclei to form two daughter cells. Cyclins were first identified in marine invertebrates as proteins whose accumulation and degradation oscillated during the cell cycle. 16 mammalian cyclins have been identified A, B1, B2, C, D1, D2, D3, E, F, G1, G2, H, I, K, T1 and T2.

Of these cyclin B1/cdc2 plays an important role in G2-M transition. The cellular proliferation occurs due to deregulation of this complex and results in uncontrolled cell growth. Cyclin B1 (48.337K Da) is a member of the cyclin family of proteins. Their level vary during the cell cycle, which is to activate specific cyclin dependent kinases (CDKs), which is essential for the proper progression through the cell cycle. In G2 phase cyclin B1 proteins begins to increase, becomes peak in mitosis, and is rapidly degraded before the cell cycle is completed. Cyclin B1 interacts with CDK1 to form a complex known as the maturation promoting factor (MPK), which is essential for cell cycle progression through mitosis.

After proper alignment of chromosome during the anaphase, the degradation of cyclin B1 occurs by anaphase promoting complex/cyclosome (APC/C) to exit mitosis, and thus cell cycle is completed. The overexpression of cyclin B1 may be due to improper degradation or may be due to increased synthesis, localization because of failure of nuclear/cytoplasm homeostasis. Over expression of cyclin B1 was seen to be positively correlating with other proliferation markers such as cyclin A, ki-67 in oral carcinomas. Over expression of cyclin B1 was noticed in squamous cell carcinomas of the tongue, cervical lymph nodes.

In this present study cyclin B1 expression was studied by immunohistochemical method using santa cruse mouse monoclonal antibody in dysplastic epithelium (tobacco and smokers) and oral cancer. The results were compared with immunohistochemically stained sections of twenty control subjects. Cyclin B1 expression was not observed in eight of the control cases. This is consistent with previous studies of absence of cyclin B1 expression in normal mucosal biopsies.
Twelve cases of control group showed cyclin B1 expression in nuclei of the cells in basal and parabasal epithelial layers consistent with the proliferative compartment of stratified squamous epithelium. This is consistent with previous study of cyclin B1 expression in basal and parabasal layers of normal mucosa.  

In the present study cyclin B1 expression was found in 17 (85%) of the twenty cases of tobacco users. Among six mild dysplasia cases four cases expressed for cyclin B1 accounting for 66.67%. Among the six moderate dysplasia cases, five expressed for cyclin B1 accounting for 83.33%. Among eight cases of severe dysplasia, seven expressed for cyclin B1 accounting for 87.5%. Khaled A et al conducted a study on ten cases of dysplastic epithelium and observed the expression for cyclin B1 in only three cases, that is 30%. In our study we found that there is step wise increase in the expression of cyclin B1 from mild to severe, although they are below cutoff point that is 15% to be considered as tumor (1000 cells counted in more than four fields).

Expression of cyclin B1 in oral squamous cell carcinoma patients habituated to tobacco in our study was observed in 13 of the 20 cases of histopathologically confirmed cases (65%). And for tumor cells the cutoff point we included was 15% (1000 tumor cells in more than four fields, and remaining were taken as negative).

In our studies the expression of cyclin B1 was observed in both nucleus and cytoplasm. Studies done by khaled A et al., the cyclin B1 expression was observed in the cytoplasm. The nuclear expression of cyclin B1 may be due to cyclin B1 tends to move from cytoplasm to nucleus with grades of conventional oral squamous cell carcinomas thus increasing the mitotic index in higher grades. Gururaj B, Patil et al.

The phenomenon for the expression of cyclin B1 in the cytoplasm is unclear, but one explanation could be that nuclear localization could be transient, where as cytoplasmic accumulation is continuous throughout the cell cycle in premalignant and malignant lesions. Khaled A et al. Another explanation may be that anticyclin B1 antibody has a higher affinity to cytoplasmic cyclin B1 because of epitopicmodifications. Most of the well differentiated oral squamous cell carcinomas, the cyclin B1 expression was detected in six cases with percentage being 75%. Among seven moderately differentiated oral squamous cell carcinomas, three cases were detected as cyclin B1 positive with the percentage being 42.85%. Among five cases of poorly differentiated carcinomas, the cyclin B1 expression was detected in four cases with the percentage being 80%. But when we compared statistically by student T test between moderately differentiated and poorly differentiated oral squamous cell carcinomas it showed no significance.

The percentage of cyclin B1 expression in OSCC was (65%) compared with the percentage of dysplastic lesions (85%), and we also found a large percentage of cyclin B1 expression in high dysplasias (87%) which may be indicative of increased traverse through the cell cycle, occurring early in tumor progression.

With regards to cyclin B1 expression and grading in oral squamous cell carcinomas, cyclin B1 expression was increased with decrease in differentiation except in moderately differentiated oral squamous cell carcinomas. We observed a predominant cyclin B1 expression in the cytoplasm as well as in the nucleus of the tumor cells. This is in contrast to the study done by Khaled et al where predominant expression was in the cytoplasm of tumor cells, rather than in the nucleus.

The significance of this phenomenon is unclear but one explanation could be that nuclear localization of cyclin B1 is transient where as cytoplasmic accumulation is continuous throughout the cell cycle in premalignant and malignant cells. Although there have been few studies on cyclin B1 expression on oral biopsies of squamous cell carcinomas, the expression of cyclin B1 in clinically normal mucosa of dysplastic (tobacco habituated) individual has not been studied.

Bibliography

[2]. Carcinogenesis from Wikipedia, the free encyclopedia. Kataline Collins, tyler jacks, nikklap pavletics, the cell cycle and cancer, science sessions, the PNAS podcast program.

DOI: 10.9790/0853-15076149154

153 | Page
Cyclin B1 Protein Expression in Oral Dysplasia and in Oral Squamous Cell Carcinoma — …

[10] Frieddilewin, M.D.1,2, staffan E. norell , M.D, hemming Johansson, B.Sc,1 per gustavsson, M.D,1 johanwennerberg, M.D,1 smoking tobacco, oral snuff, and alcohol in the etiology of squamous cell carcinoma of the head and neck.


[16] Urmila J nair1,2, gunte obe1, marlin friessen1, mark T goldberg4, and helmat bartsch1, role of lme in the generation of reactive oxygen species from betel-quid ingredients. Journal of environmental health perspectives. 1992 ; 98: 203-205.


[26] Urmila J nair1,2, gunte obe1, marlin friessen1, mark T goldberg4, and helmat bartsch1, role of lme in the generation of reactive oxygen species from betel-quid ingredients. Journal of environmental health perspectives. 1992 ; 98: 203-205.


[34] Jatin k nagpal; sriivas pathak1, bibhu R das2,3, prevalence of high risk human papilloma virus types and its association with psx cdon 72 polymorphism in tobacco addicted oral squamous cell carcinoma (oscc) patients of eastern Indian, int.j.cancer. 2002;97:649-653.


[38] Winston Patrick, kuo,DDS,Ms, tor-kristianjensen,Msc, peter j.park,phd, marks W.ingen, DDS,phd, rifatharinaDDS,phd, lucia duo-rochado,MD,phd1. Journal of annual symposium. 2002; 415-419.


