IL28B Gene polymorphisms in prediction of PEG-IFN-α therapy in HCV Egyptian patients

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Abstract:

Background: Chronic HCV is one of the commonest reasons for ceaseless liver malady worldwide with around 15% of populace tainted in Egypt. Certain SNPs present close to the IL28B gene were found to affect the spontaneous clearance and also treatment result of HCV. To analyze the relationship between different IL28B variations and the relapse of HCV disease after consolidated treatment with ribavirin and pegylated interferon (pegIFN).

Methods: Forty HCV genotype all patients got 1.5 mg/kg/week peg-interferon alfa-2b in addition to 800-1400 mg/d ribavirin (weight-balanced) for 48 weeks

IL28B polymorphism rs12979860 was concentrated on in responders and relapsers at week 72.

Results: Out of 40 patients receiving treatment, 7 (17.0 %) were relapsers. By stratifying patients on the principle of the IL-28/60 genotype CC versus (CT&TT), CC patients demonstrated lower relapse rates (12.5%) compared to CT/TT patients (87.5%) (P < 0.05).

Conclusions: IL28B polymorphism can forecast response to PEG-IFN-a treatment in HCV patients List of abbreviations:

- HCV Hepatitis C infection pegIFN pegylated interferon
- **RBV**: ribavirin
- SVR: supported virological reaction
- DAAs: Direct acting antivirals
- SNP: single nucleotide polymorphism
- IL28B: Interleukin 28B
- IP10 : Interferon-gamma (IFN-gamma)-inducible protein-10

Key Words: HCV; IL28B polymorphism; relapsers

Background I.

In the recent decades Hepatitis C disease (HCV) has tainted more than 170 million people in the world as assessed by sheet (1). The pervasiveness of HCV is high in Egypt with around 14.7% of people polluted (2-4).

HCV is classified according sequence similarities within sequences from core, NS5 areas, into seven genotypes (with 60-70% grouping similarity) and various subtypes (with 75-85% sequence similarity) (5). In the average of 80% of patients with HCV infection get to be chronic which may advance to cirrhosis and even hepatocellular carcinoma significant proportion of patients (6-8). The basic components for the diligence and pathogenesis of the infection are not completely understood.

Nevertheless, one of the clarifications may be the viral avoidance of the natural invulnerable reaction (9). The infection meddles at various levels with the different flagging pathways (10, 11). The suggested treatment of unending HCV contamination in the previous couple of years was basically a mix of pegylated IFN-a (pegIFN) and ribavirin (RBV), which gives variable rates of achievement. The viral genotype notwithstanding has variables as IL-28 B genotype assumes a vital part in building up a supported virological reaction (SVR) (12).

Direct acting antivirals (DAAs) are now the new hope for chronic hepatitis patients. The principal model of DAAs were the NS3/4A protease inhibitors telaprevir and boceprevir, which were used in combination with pegIFN and RBV for genotype 1 in (13, 14), this expanded the SVR rate from 40% to 75%. The second one, simeprevir, which is another protease inhibitor, extended the SVR rate up to 90% when used as a piece of blend with pegIFN and RBV (15). Without interferon treatment has been moved with the use of sofosbuvir, the primary nucleotide NS5B inhibitor (16-21).

As in various sicknesses, where the genetic beautifiers of an individual impacts the prescription result, certain single nucleotide polymorphisms (SNPs) lying close the IL28B quality were found to impact the unconstrained flexibility and also treatment aftereffect of HCV (12, 22-24). Considerably more, the CC genotype of the single nucleotide polymorphism (SNP) rs12979860 was a decent indicator for SVR in early trials of the DAA boceprevir (25). IL28B quality encodes sort III interferon IFN-13 furthermore, is connected with more than a 2 times contrast in light of HCV medication treatment (12). In any case, the significance of IL28B polymorphism to viral backslide has not been assessed in Egypt in this way.

The vast majority of the past studies concentrated on SVR as an end point, however did not examine the status of backslide in those turned out viremic after leeway from viremia at end purpose of treatment at week 48. The present study was directed to decide the relationship of IL-28B rs12979860 quality polymorphism with the reaction (counting backslide) taking after double pegIFN what's more, ribavirin treatment in genotype 4 HCV Egyptian patients.

II. Patients and Methods:

This is a cross-sectional study conducted on 40 patients attending at the EL-Kaser El-Ainy hospital between 2013 and 2015, who received 1.5 mg/kg/week pegIFN alfa-2b in addition to 800-1400 mg/d ribavirin (weight- adjusted) for 48 weeks. IRB endorsement was gotten and patients gave composed educated agree to make their medical records available for this study.

III. Sample Collection:

At the beginning and before the start of treatment samples were collected (at zero point) and at week 72. After patients had end of the treatment period (ETR; at week 48), they were later partitioned into two groups as indicated by the SVR (Supported Virological Reaction; at week 72).

Group A: included 33 patients with maintained viral reaction (SVR) and Group B: included 7 relapsers.

RNA was extracted by using QIAmp Viral RNA Kit (QIAGEN, Santa Clarita, CA) according to the manufacturer's instructions and the RNA levels were quantified by using Abbott Real Time HCV amplification reagents kit (Abbott Laboratories. Abbott Park, IL), according to the manufacturer's instructions (detection limit 12 IU/ml). Relapse rates during the follow-up period were prospectively calculated according to baseline viral load (</>800.000IU/ml) and to viral declineat weeks 4 and 12 of therapy, respectively.

HCV genotyping

Viral RNA was extracted by using QIAmp Viral RNA Kit (QIAGEN, Santa Clarita, CA) From the nested PCR results of 237 bp, 10 ml were processed by restriction endonuclease enzymes for 2 hr at 37_oC by both MvaI/HinfI in buffer H and RsaI/HaeIII in buffer L (Boehringer Manheim, Germany) (26, 27) in a total volume of 20 ml. All enzymes were utilized at 7.5 units/response. Electrophoresis was done in a 4% Metaphore gel in 0.5X TBE buffer (FMC Bio *items, Rockland, ME*).

• Isolation of genomic DNA and SNP genotyping

IL28B SNP rs12979860 genotype were determined using PCR-RFLP. Genomic DNA was removed from 200 ml whole blood using the QIAamp DNA Blood Mini unit (Qiagen, Hilden, Germany) as per producer's guidelines. primers are listed in Table I (28, 29).

SNP	Primer	Sequence
rs12979860	Forward SCCAGGGCCCCTAACCTCT-GCA 3	
	Reverse	ЗGGGAGCGCGGAGT-GCAATTCA З

Table I. Primers for PCR Genotyping of rs12979860,

PCR was performed by using 25 ml reaction tube. PCR mixture containing 2.5 ml of 10X response buffer with MgCl2 (Fermentas, Vilnius, Lithuania), 10 pmol of primers (forward and invert), 0.2 mM/L dNTPs (Fermentas, Vilnius, Lithuania), 2 units Taq DNA polymerase (Fermentas, Vilnius, Lithuania) and 100-200 ng genomic DNA format. The cycling conditions began with beginning denaturation at 95_oC for 5 min, trailed by 35cycles under the accompanying conditions: • Denaturation at 94_oC for 30 sec.

• Annealing at 66°C for 30 sec.

• Extension at 72°C for 30 sec.

• Final extension at 72°C for 5 min.

Successful amplification was by the presence of right size bands on 2% agarose gel electrophoresis.

• Digestion with Restriction Enzyme

Ten micro liters of amplicons were processed with 10 units of fast digest BseMI (BsrDI) restriction endonuclease (Fermentas, Vilnius, Lithuania) at 37_oC for 15 min. Digested products were isolated on 3% agarose gel together with the PhiX174 DNA/HaeIII digest molecular weight marker (Finzyme) which incorporates 11 separate fragments (1353, 1078, 872, 603, 310, 281, 271, 234, 194, 118, and 72 bp). To confirm the digestion, the digested fragments were stained and visualized on UV transilluminator.

IV. Statistic and data processing

Mean and stander deviation were used to expressed quantitative variables normally distributed data and as middle and interquartile for other data, student t-test or Mann-Whitney test as appropriate. Qualitative variables were expressed by number (frequency) and rate analyzed between gatherings utilizing Chi-square test or Fisher definite likelihood test as suitable. Binary logistic regression analysis was done with non-response to treatment as the dependent factor. Adjusted odds ratios (aORs), 95% Confidence Intervals (CIs). AORs were balanced for age, pre-treatment and viral load. In all tests, P quality was viewed as noteworthy if under 0.05.

V. Results

This study involved 40 patients, 29 males and 11 females; whose ages went from 25 to 58 years with a mean of (42.8 ± 8.2) . Patients were subdivided according to SVR at week 72 into two groups; group 1: (responders) 33 (82.5%) and group 2: (relapsers) 7 (17.5%).

There was no significant change between the responders and relapsers in regards age, sex, IP10, ALT, and AST levels. All patients of responders and relapsers were genotype 4. A significantly higher viral load was observed in relapse group (Table II, Table III).

A non-significant difference was seen in the allele's dissemination of IL-28/60 amongst responders and relapsers (Table IV). The frequency of the IL-28/60 genotype CC, CT and TT are 27.5, 57.5 and 15 % respectively.

The percentage of different genotypes in responders and non-responders are shown in table IV. The different alleles of IL28/60 responders and non-responders are shown in Table VI. In the cross tabulation between (CC vs. CT/TT), the CC genotype didn't achieve a significantly change regarding response rates (P > 0.05) (Table VI).

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Table II.	Baseline	Charac	cteristics	of the	Studied	Group

	Groups			
	Responders (n=33)	Relapsers (n =7)	<i>r</i> -value	
Age (years) [#]	40.1 ± 9.78	44.3 ± 4.46		
Sex				
Males	45 (80.4)	11 (84.6%)	0.7	
Females	11 (19.6)	2 (15.4)		
Pre ALT (U/L)*	64 (42–79.8)	53 (31–102)	0.9	
Pre AST (U/L)*	39 (29–58)	40 (31–92)	0.6	
Viral load (copies/ml) [*]	1005190 (49,268-5,736,527)	1161192 (130,872–4,231,010)	0.01	

*median and interquartile for non-distributed data.

**Categorical data are presented as number and (percentage).

Table III.	IP10 in	responder	and relapser
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IP-10	Mean <u>+ </u> SD	P-value
	226.02 <u>+</u> 245.3	>0.05

Using different genetic models for IL-28/60, carriers of all C alleles and CC genotype had a significant protective effect from relapse compared to T allele carriers (**Table IV**).



Fig.1a. IP10 in non- responder

IP10

Fig.1b. IP10 in responder

Table IV. Genotype	s in response and	non-response groups
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	Geno Type		
	C\T	C/C	т\т
Responders	51.5%	30.3%	18.2%
Non-Responders	85.7%	14.3%	0.0%

	Gen	P-value		
	CT+TT	CC	1-value	
Responders	71.9%	30.3%	>0.05	
Non-Responders	85.7%	14.3%	>0.03	

Table V. Cross tabulation CT + TT vs. C\C) response and non-responders

Table VI. Frequency of Alleles of IL28/60 in Responders and non-relapsers

	G		
Allele	Responders	Non-Responders	P-value
С	37 (53%)	8 (55)	
Т	27 (46%)	9 (45)	P > 0.05



Discussion

This study was conveyed in the period somewhere around 2012 and 2015 on HCV patients getting double treatment (pegIFN and ribavirin) before beginning the use of DAAs. The forecast of treatment result is critical to diminish antagonistic impacts and costs (30). Different studies from European and middle Eastern contries demonstrated that SVR in genotype 4 after mixed treatment with pegIFN and ribavirin; ranges somewhere around 43% and 70% (31, 32).

Polymorphisms present close to the gene encoding IL28B were found to be the best indicator of patient reaction to pegIFN and ribavirin for chronic hepatitis C virus (HCV) genotype 1 infection (23, 33, 34). The majority of the past studies did not address the determination of the polymorphism in the site in IL28B in cases of relapse.

As we probably know, this is one of the primary studies intended to examine the part of SNP of the IL28B gene, the rs12979860, genotype, as indicators of relapse after pegIFN in addition to ribavirin in HCV genotype 4 patients of Egyptian race. In this study, time of relapsed patients ranged from 21 to 58 years with a mean \pm SD of (40.8 \pm 9.2) with no significant difference between the responders and relapsers in regards to age and sex. This is concurred with Khairy (35) who demonstrated that there was no significant relationship between age or sex amongst responders and relapsers. Genome wide affiliation examines (GWAS) have demonstrated that hereditary polymorphism at rs12979860 close IL28 quality on chromosome 19 is connected with variable reactions to antiviral treatment with an a few fold increments in SVR in the CC genotype at rs12979860 genotype of IL-28B contrasted with CT or TT genotypes (12, 36).

In this study, CT genotype was the most genotype in non-responders (85.7%) regards to IL-28/60, while T\T is the most in responders. The T allele was more persistent among non-responders than the C allele, which is predictable with numerous studies (37-39). Our outcomes are matched with Esmail This is rather than Khairy (35), who found that every one of the genotypes; CC CT and TT, were similarly significantly correlated with SVR.

The absence of information identified with cirrhosis and fibrosis was an impediment of this study as the nearness of cirrhosis is connected with a low reaction to antivirals.

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Conflict of Interest

The authors declare no conflict of interest. This research received no specific grant from any funding agency in the public or commercial sphere.

Availability of Data and Materials

All availability of data and materials are including in the tables under results section

References

- sheet Wf. World Health Organization. WHO fact sheet 2000;164. [1].
- [2]. El-Zanaty F, Way A. Feeding practices and micronutrient supplementation. Egypt Demographic and Health Survey 2008. 2009:166-75.
- Health. EMo. Report Annual. Accessed October. 2013;13. [3].
- [4]. Lehman EM, Wilson ML. Epidemiology of hepatitis viruses among hepatocellular carcinoma cases and healthy people in Egypt: A systematic review and meta-analysis. International journal of Cancer. 2009;124(3):690-7
- [5]. Nakano T, Lau GM, Lau GM, Sugiyama M, Mizokami M. An updated analysis of hepatitis C virus genotypes and subtypes based on the complete coding region. Liver international. 2012;32(2):339-45.
- Seeff LB. The natural history of chronic hepatitis C virus infection. Clinics in liver disease. 1997;1(3):587-602. [6].
- [7].
- Sen GC. Viruses and interferons. Annual review of microbiology. 2001;55:255-81. Micallef JM, Kaldor JM, Dore GJ. Spontaneous viral clearance following acute hepatitis C infection: a systematic review of longitudinal studies. [8]. Journal of viral hepatitis. 2006;13(1):34-41.
- Bode JG, Brenndorfer ED, Haussinger D. Subversion of innate host antiviral strategies by the hepatitis C virus. Archives of biochemistry and [9]. biophysics. 2007;462(2):254-65.
- [10]. Li K, Foy E, Ferreon JC, Nakamura M, Ferreon AC, Ikeda M, et al. Immune evasion by hepatitis C virus NS3/4A protease-mediated cleavage of the Toll-like receptor 3 adaptor protein TRIF. Proceedings of the National Academy of Sciences of the United States of America. 2005;102(8):2992-7.
- [11]. Meylan E, Curran J, Hofmann K, Moradpour D, Binder M, Bartenschlager R, et al. Cardif is an adaptor protein in the RIG-I antiviral pathway and is targeted by hepatitis C virus. Nature. 2005;437(7062):1167-72.
- Ge D, Fellay J, Thompson AJ, Simon JS, Shianna KV, Urban TJ, et al. Genetic variation in IL28B predicts hepatitis C treatment-induced viral [12]. clearance. Nature. 2009;461(7262):399-401.
- Jacobson IM, McHutchison JG, Dusheiko G, Di Bisceglie AM, Reddy KR, Bzowej NH, et al. Telaprevir for previously untreated chronic hepatitis C [13]. virus infection. The New England journal of medicine. 2011;364(25):2405-16.
- [14]. Poordad F, McCone J, Jr., Bacon BR, Bruno S, Manns MP, Sulkowski MS, et al. Boceprevir for untreated chronic HCV genotype 1 infection. The New England journal of medicine. 2011;364(13):1195-206
- Fried MW, Buti M, Dore GJ, Flisiak R, Ferenci P, Jacobson I, et al. Once-daily simeprevir (TMC435) with pegylated interferon and ribavirin in [15]. treatment-naïve genotype 1 hepatitis C: The randomized PILLAR study. Hepatology. 2013;58(6):1918-29.
- Gane EJ, Stedman CA, Hyland RH, Ding X, Svarovskaia E, Symonds WT, et al. Nucleotide polymerase inhibitor sofosbuvir plus ribavirin for hepatitis [16]. C. New England Journal of Medicine. 2013;368(1):34-44.
- [17]. Feld JJ, editor Interferon-free strategies with a nucleoside/nucleotide analogue. Seminars in liver disease; 2014.
- Gentile I, Buonomo AR, Zappulo E, Borgia G. Interferon-free therapies for chronic hepatitis C: toward a hepatitis C virus-free world? Expert review [18]. of anti-infective therapy. 2014;12(7):763-73.
- Sofia MJ. Beyond sofosbuvir: what opportunity exists for a better nucleoside/nucleotide to treat hepatitis C? Antiviral research. 2014;107:119-24. [19]. [20]. Sulkowski MS, Gardiner DF, Rodriguez-Torres M, Reddy KR, Hassanein T, Jacobson I, et al. Daclatasvir plus sofosbuvir for previously treated or
- untreated chronic HCV infection. The New England journal of medicine. 2014;370(3):211-21.
- [21]. Welzel TM, Zeuzem S, editors. Interferon-free strategies without a nucleoside/nucleotide analogue. Seminars in liver disease; 2014.
- Suppiah V, Moldovan M, Ahlenstiel G, Berg T, Weltman M, Abate ML, et al. IL28B is associated with response to chronic hepatitis C interferon-a [22]. and ribavirin therapy. Nature genetics. 2009;41(10):1100-4. Tanaka Y, Nishida N, Sugiyama M, Kurosaki M, Matsuura K, Sakamoto N, et al. Genome-wide association of IL28B with response to pegylated [23].
- interferon-α and ribavirin therapy for chronic hepatitis C. Nature genetics. 2009;41(10):1105-9
- [24]. Thomas DL, Thio CL, Martin MP, Qi Y, Ge D, O'hUigin C, et al. Genetic variation in IL28B and spontaneous clearance of hepatitis C virus. Nature. 2009;461(7265):798-801.
- [25]. Gordon SC. Utility of IL-28B Polymorphisms in the era of direct-acting antiviral therapy. Gastroenterology & hepatology. 2011;7(8):531.
- McOmish F, Yap PL, Dow BC, Follett EA, Seed C, Keller AJ, et al. Geographical distribution of hepatitis C virus genotypes in blood donors: an [26]. international collaborative survey. Journal of clinical microbiology. 1994;32(4):884-92. Constantine NT, Abdel-Hamid M, Oldach D. Rapid genotyping of hepatitis C virus. The New England journal of medicine. 1995;333(12):800
- [27].
- El-Awady MK, Mostafa L, Tabll AA, Abdelhafez TH, Bader El Din NG, Zayed N, et al. Association of IL28B SNP With Progression of Egyptian [28]. HCV Genotype 4 Patients to End Stage Liver Disease. Hepat Mon. 2012;12(4):271-7.
- [29]. Sharafi H, Pouryasin A, Alavian SM, Behnava B, Keshvari M, Mehrnoush L, et al. Development and Validation of a Simple, Rapid and Inexpensive PCR-RFLP Method for Genotyping of Common IL28B Polymorphisms: A Useful Pharmacogenetic Tool for Prediction of Hepatitis C Treatment Response. Hepat Mon. 2012;12(3):190-5.
- Derbala M, Rizk NM, Al-Kaabi S, John A, Sharma M, El-dweik N, et al. The predictive value of IL28B rs12979860, rs11881222 and rs8099917 [30]. polymorphisms and IP-10 in the therapeutic response of Egyptian genotype 4 patients. Virology. 2013;444(1):292-300.Hasan F, Asker H, Al-Khaldi J, Siddique I, Al-Ajmi M, Owaid S, et al. Peginterferon alfa-2b plus ribavirin for the treatment of chronic hepatitis C genotype 4. The American journal of gastroenterology. 2004;99(9):1733-7.
- [31]. Moucari R, Ripault M-P, Martinot-Peignoux M, Voitot H, Cardoso A-C, Stern C, et al. Insulin resistance and geographical origin: major predictors of liver fibrosis and response to peginterferon and ribavirin in HCV-4. Gut. 2009;58(12):1662-9
- [32]. Cieśla A, Bociąga-Jasik M, Sobczyk-Krupiarz I, Głowacki MK, Owczarek D, Cibor D, et al. IL28B polymorphism as a predictor of antiviral response in chronic hepatitis C. World journal of gastroenterology: WJG. 2012;18(35):4892.
- Coppola N, Marrone A, Pisatro M, Starace M, Signoriello G, Gentile I, et al. Role of interleukin 28-B in the spontaneous and treatment-related clearance of HCV infection in patients with chronic HBV/HCV dual infection. European journal of clinical microbiology & infectious diseases : [33]. official publication of the European Society of Clinical Microbiology. 2014;33(4):559-67.
- [34]. Khairy M, Fouad R, Mabrouk M, El-Akel W, Awad AB, Salama R, et al. The impact of interleukin 28b gene polymorphism on the virological response to combined pegylated interferon and ribavirin therapy in chronic HCV genotype 4 infected egyptian patients using data mining analysis. Hepatitis monthly. 2013;13(7).
- Thompson AJ, Muir AJ, Sulkowski MS, Ge D, Fellay J, Shianna KV, et al. Interleukin-28B polymorphism improves viral kinetics and is the strongest [35]. pretreatment predictor of sustained virologic response in genotype 1 hepatitis C virus. Gastroenterology. 2010;139(1):120-9. e18. Stättermayer AF, Stauber R, Hofer H, Rutter K, Beinhardt S, Scherzer TM, et al. Impact of IL28B genotype on the early and sustained virologic
- [36]. response in treatment-naive patients with chronic hepatitis C. Clinical gastroenterology and hepatology. 2011;9(4):344-50. e2.
- Asselah T, De Muynck S, Broët P, Masliah-Planchon J, Blanluet M, Bièche I, et al. IL28B polymorphism is associated with treatment response in [37]. patients with genotype 4 chronic hepatitis C. Journal of hepatology. 2012;56(3):527-32.
- [38]. De Nicola S, Aghemo A, Grazia Rumi M, Galmozzi E, Valenti L, Soffredini R, et al. Interleukin 28B polymorphism predicts pegylated interferon plus ribavirin treatment outcome in chronic hepatitis C genotype 4. Hepatology. 2012;55(2):336-42.