Liveweight Changes and Reproductive Performance of Snails (Archachatina Marginata) Under Controlled and Naturally Induced Aestivation

*Omoyakhi1, J.M., Edo-Taiwo2, O., Aremu3, O.T., Okhale1, O.E.
1Department of Animal Science, Faculty of Agriculture, University of Benin, Benin City, Nigeria
2Department of Animal and Environmental Biology, Faculty of Life Science, University of Benin, Benin City, Nigeria
3Department of forestry and wildlife, Faculty of Agriculture, University of Benin, Benin City, Nigeria

Abstract: The liveweight changes and reproductive performance of snails (Archachatina marginata) under controlled (watered throughout the experimental period, aestivated for 6 and 12 weeks) and naturally induced aestivation were investigated. Eighty (80) reproductively matured snails with five snails replicated four times in a completely randomized design were used. Liveweight and mortality were monitored during aestivation. Eggs collected were incubated. While the continuous watered group (0A) gained 11.95 g, the groups aestivated for 6 weeks (6A), 12 weeks (12A) and by nature (N) respectively recorded -40.05, -88.28 and -51.29 g. Mortality records showed 1, 1, 2 and 0. Apart from clutch size and number of snaillet hatched, the investigated reproductive indices reflected significant (P<0.05) differences. The average clutch number was highest in 0A (5.65) followed by 12A (4.25), 6A (2.80) and N (2.67). Clutch weight was highest in N (25.17 g) which differ statistically from 0A (21.54). This was however not significantly different from 6A (22.88g) and 12A (24.32g). These respectively gave an average egg weight of 2.92, 2.91, 3.16 and 3.21 g for 0A, 6A, 12A and N. Hatchability was significantly highest in 6A (95.92%) which was not significantly different from 12A (93.69 %) and N (89.48 %). The later however, did not differ statistically from 0A. 0A nevertheless gave the highest snaillet weight (2.63 g) and the least was recorded from 6A (1.53 g). It is evident from this study that aestivation although depressed weight and overall number of eggs laid, it however favoured hatchability and incubation period in A. marginata.

Keywords: Aestivation, Archachatina marginata, liveweight, Reproductive performance, egg

I. Introduction

In Nigeria, snails are collected from the wild and there has been noticeable decline in wild stock. Of serious concern is that in recent time, the popular Achatina achatina has been observed in much short supply in Nigeria. Beside this decline in this and several other species, it is also noted that meeting the current and anticipated demand for snail meat and products cannot rely on wild supply (Omoyakhi and Okhale, 2015a). Clearly, availability of captive bred snails among other things will augment the exploitation of the wild stock.

Snails have been included to a group called miniframe species of livestock that are increasingly playing an important role in human nutrition (Eruvbetine, 2012). Snails are tractable, prolific and widely accepted for public healthy consumption and have been described as one of the surest ways of ameliorating the shortfall of animal protein in Nigeria (FAO, 1970; Mbah, 1989 and Onadeko, 2002). In view of this, the rearing of snail in captivity is attracting the keen interest of both scientists and farmers, suggesting the potential of this species as farm animal of the future in the West Africa sub-region.

In captive rearing and management of wild species of animals like snail, Okhale et al. (2015) asserted that utmost attention should be given to their life pattern as provided by nature in order to maximize their survival, growth and reproductive potential. Consequently, practices capable of adverse short or long term effects are avoided. This also enables conditions and factors vital for sustainable and optimal productivity to be integrated in captive rearing and management of wild species of animals. This study therefore seeks to investigate the liveweight changes and reproductive performance of snails (Archachatina marginata) under controlled (watered throughout the experimental period, aestivated for 6 and 12 weeks) and naturally induced aestivation.

II. Materials And Methods

Experimental Location

The research was carried out at the Snailry unit of the University Teaching and Research Farm, University of Benin, Edo State, Nigeria. The farm is located within the tropical rain forest vegetation zone of Southern Nigeria, lying between longitude 5°E and 6°42’E and latitude 5°45 and 7°34’N of the equator (NAA,
Liveweight Changes And Reproductive Performance Of Snails (Archachatina Marginata) Under Natural and Varying Aestivation Lengths

2015). On the north Edo State is bounded by Kogi State, to the east is Anambra State, south by Delta State and west by Ondo State. The climate of Edo is humid.

Materials
The materials used include a total of 80 healthy snails of Archachatina marginata with liveweight of 150 to 280 g. shallow containers for feed and water, a sensitive electronic weighing scale, identification materials and plastic incubators.

Experimental design
The experiment was laid out in a completely randomized design (CRD). Eighty (80) reproductively matured snails with five snails replicated four times were used. Liveweight and mortality were monitored during aestivation and the reproductive performance post aestivation.

Experimental procedure
The brick hutches (90 cm square with a depth of 54 cm) were prepared and assigned shallow plastic troughs (10 cm x 6 cm x 2 cm) for concentrate feed. The snails were balanced for liveweight and randomly allotted to the four treatments. Feeding and watering were provided as necessitated for the study. The feed materials supplied included fruits of pawpaw and water melon, leaves of cocoyam, pumpkin, plantain and water leaves. The concentrate was formulated with the following nutritional information (calculated); CP = 16 %, ME = 2,469.40 kcal/kg, CF = 5.79 %, EE = 3.91 %, Ash = 3.67 %, NFE = 49.05 %, Lysine = 0.59 %, Met + cyst = 0.50 %, Ca = 5.31 and P = 1.07.

Routinely, cleaning of troughs, regular clean-up and changing of the feeds as well as regular check for insects and other predators were ensured. The trench around the cells was also provided with water and suitable pesticides to prevent insect and other pests from crawling into the cells.

Snails in group 0A were fed, watered and moistened throughout the experimental period. Group 6A and 12A were allowed a 6 - and 12 - week unbroken aestivation period while snails in group N were controlled by natural weather elements. Group 6A was induced 6 weeks after Group 12A had been induced. This enabled termination of aestivation of all the groups and simultaneous data collection.

Two (2) weeks acclimatization (adjustment) period was allowed prior to the commencement of the research which was programmed for the month of December. This enabled the completion of all the aestivation lengths (including the 12 weeks aestivation length) within the pre-spawning period of GALS in the tropics which last from March to April as reported by Cobbinah (1992). Egg production in each group was monitored up to month of August. Eggs laid were collected and incubated immediately.

Statistical analysis
The data obtained for reproductive performance were subjected to one way analysis of variance (ANOVA) using Genstat, 2009 (12th Edition) and significant treatment means were separated by Duncan Multiple Range Test.

Experimental design and induction of aestivation
The experiment was conducted using 80 reproductively matured snails, laid out in a completely randomized design, with 5 snails replicated four times. Snails were induced to aestivate under prevailing atmospheric conditions in this study by discontinued moistening of soils and withdrawal of feed and water as reported in several researches on snail aestivation (Omo yakhi et al., 2008a, b, Ajayi et al., 2012; Adeoba et al., 2012).

III. Result
Weight change and mortality of Archachatina marginata Under Natural and Varying Aestivation Lengths
Analysis of variance as shown in table 4.1 reflects significant (P<0.05) weight loss of the aestivated (6 and 12 weeks) and naturally induced group. These were not significantly different from each other but differ significantly from the group that was watered throughout the experimental period. The continuous watered group however, only gained 11.95 g (6.01 %) throughout the investigation periods. The weight loss was highest in the 12th week aestivated group (-88.28 g) followed by the naturally controlled group (-51.29 g) and then the 6 weeks aestivation treatment group (-40.05 g). This accounted for about 35.41 %, 23.82 % and 16.41 % of their respective initial weight. The 12 weeks aestivation treatment group suffered 2 mortalities (in week 2 and 12) and one (1) each from the 6 weeks induced aestivation group and the continuous watering groups. No mortality was recorded in the naturally controlled group. The trend in the change in weight in the respective treatments groups is shown in figure 1. The continuous watered group experienced stable weight changes. Noticeable loss in weight was observed in week two and gain in the 12th week. The aestivated groups (including the naturally
Liveweight Changes And Reproductive Performance Of Snails (Archachatina Marginata) Under Controlled

Liveweight Changes And Reproductive Performance Of Snails (Archachatina Marginata) Under

controlled) had consistent drop in weight. Major weight loss was observed in week 2 and 8 in the 12 weeks
aestivation group. The naturally induced aestivation group had a slight gain in weight, thereafter a fairly stable
weight up to the 7th week. Only marginal weight loss was observed in the 2nd week of aestivation in the 6
weeks aestivation treatment group.

Table 1: Mean Weight Change and Mortality of Archachatina marginata Under Natural and Varying
Aestivation Lengths

<table>
<thead>
<tr>
<th>Initial Weight (g)</th>
<th>Continuous watered</th>
<th>6 weeks</th>
<th>12 weeks</th>
<th>Naturally controlled</th>
<th>SEM</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>199.00^a</td>
<td>244.10^a</td>
<td>249.30^a</td>
<td>215.30^b</td>
<td>13.97</td>
</tr>
<tr>
<td>Final Weight (g)</td>
<td>206.80^a</td>
<td>182.80^b</td>
<td>161.00^b</td>
<td>164.00^b</td>
<td>9.71</td>
</tr>
<tr>
<td>Weight change (g)</td>
<td>11.95^a</td>
<td>-40.05^b</td>
<td>-88.28^b</td>
<td>-51.29^b</td>
<td>16.40</td>
</tr>
<tr>
<td>Mortality</td>
<td>1</td>
<td>1</td>
<td>2</td>
<td>4</td>
<td></td>
</tr>
</tbody>
</table>

Values are least square means (± sem)
abc Means with different superscripts within the same parametric row differ significantly (P<0.05)

Figure 1: Trend in Weight Change over the weeks of Aestivation of Archachatina marginata Under Natural
and Varying Aestivation Lengths

Reproductive Performance of Archachatina Marginata Under Natural and Varying Aestivation Lengths

Apart from clutch size and number of snaillet hatched, the investigated reproductive performance
indices as shown in table 4.3 reflected significant (P<0.05) differences among the treatments. The averages
clutch number was significantly highest in the continuous watering group (5.65) and followed by the 12 weeks
aestivation group (4.25). The 6 weeks aestivation group recorded 2.80 average numbers of clutches. Clutch
weight was highest in the naturally controlled group (25.17 g), which was different statistically from the
continuous watering group (21.54). It was however not significantly different from the 6 (22.88 g) and the 12
weeks (24.32 g) groups. These respectively gave an average egg weight of 2.92 g, 2.907 g, 3.16 g and 3.21 g for
the continuous watering group, 6, 12 and naturally controlled aestivation groups. Hatchability was significantly
highest with the 6 weeks aestivation group (95.92 %) which was not significantly different from the 12 weeks
(93.69 %) and naturally controlled group (89.48 %). The later however did not differ statistically from the
continuous watering group (85.83 %). This nevertheless gave the highest snaillet weight (2.63 g) and the least
recorded from the 6 weeks aestivation group.
Liveweight Changes And Reproductive Performance Of Snails (Archachatina Marginata) Under Natural and Varying Aestivation Lengths

<table>
<thead>
<tr>
<th>Reproductive Indices</th>
<th>Aestivation treatments</th>
<th>Continuous watering</th>
<th>6 weeks</th>
<th>12 weeks</th>
<th>Naturally controlled</th>
<th>SEM</th>
</tr>
</thead>
<tbody>
<tr>
<td>Date of first egg lay</td>
<td>56.00°</td>
<td>92.00°</td>
<td>98.00°</td>
<td>64.00°</td>
<td>2.020</td>
<td></td>
</tr>
<tr>
<td>Clutch number</td>
<td>5.65°</td>
<td>5.80°</td>
<td>4.25°</td>
<td>2.66°</td>
<td>0.290</td>
<td></td>
</tr>
<tr>
<td>Clutch size</td>
<td>7.43°</td>
<td>7.49°</td>
<td>7.98°</td>
<td>7.54°</td>
<td>0.303</td>
<td></td>
</tr>
<tr>
<td>Clutch weight (g)</td>
<td>2.34°</td>
<td>2.88°</td>
<td>2.32°</td>
<td>2.71°</td>
<td>0.982</td>
<td></td>
</tr>
<tr>
<td>Average egg weight (g)</td>
<td>2.91°</td>
<td>2.90°</td>
<td>3.15°</td>
<td>3.21°</td>
<td>0.209</td>
<td></td>
</tr>
<tr>
<td>Incubation period (days)</td>
<td>29.33°</td>
<td>25.33°</td>
<td>25.33°</td>
<td>26.00°</td>
<td>0.408</td>
<td></td>
</tr>
<tr>
<td>Number hatched</td>
<td>5.93°</td>
<td>5.91°</td>
<td>6.25°</td>
<td>4.66°</td>
<td>0.583</td>
<td></td>
</tr>
<tr>
<td>Hatchability (%)</td>
<td>85.83°</td>
<td>95.92°</td>
<td>93.69°</td>
<td>89.48°</td>
<td>2.520</td>
<td></td>
</tr>
<tr>
<td>Weight of hatching</td>
<td>16.06°</td>
<td>10.48°</td>
<td>16.32°</td>
<td>15.18°</td>
<td>0.711</td>
<td></td>
</tr>
<tr>
<td>Weight of snail</td>
<td>2.63°</td>
<td>1.53°</td>
<td>2.22°</td>
<td>1.83°</td>
<td>0.128</td>
<td></td>
</tr>
</tbody>
</table>

Values are least square means (± sem)

Means with different superscripts within the same parametric row differ significantly (P<0.05)

IV. Discussion

There was significant decline in liveweight of snails in all the aestivated groups. The percentage decline correlates relatively to the aestivation lengths. This may be partly due to autophagy and/or the use of a large amount of energy reserves in different body tissue (Lukong and Onwubiko, 2004; Omoyakhi and Okhale, 2015b,c). In this study, while the continuous watered group gained 6.01 % of its initial weight, 35.41, 16.41 and 23.82 % weight loss were observed from 6A, 12A and N respectively. In a similar research, Emerson and Duer (1967) reported weight reduction of 62 % in Littorina planaxis. About 50 % loss in weight was also reported by Russel-Hunter and Eversole (1976) after 132 days of starvation in the fresh water pulmonate snail, Helisoma trivolvis. Lukong and Onwubiko (2004) observed a liveweight decline of 44.60 % in Archachatina achatina aestivated for 4 months. Omoyakhi (2008a) and Abdusamad et al. (2010) reported a liveweight reduction of 52.4 % and 35.6 % of initial weight for A. marginata after six weeks of aestivation respectively. Snails undergo aggressive recovery when the environment becomes favourable and water and feed are available post aestivation (Cobbina, 1992, Rizzatti and Romero, 2001). Omoyakhi and Osinowo (2010a,b) observed significant compensation for this loss in liveweight and other components studied after snails were arouse of aestivation. They noted that, within two weeks post-aestivation, the liveweight exceeded their pre-aestivation values by 192.6 % for Archachatina marginata. The first egg lay recorded was from the continuous watered group (25th February). This coincided with the date of the third rain of the year in the study area. The first two rains would probably have triggered the initiation of the reproductive behaviour and consequently the commencement of egg lay. The N group had its first lay on the 5th March. Bonnefoy-Claudet and Deray (1984) reported 52 and 85 days as day of first mating and egg lay respectively for Cantareus aspersus hibernated for 1.5 months. Omoyakhi and Osinowo (2012) reported 72 days for first record of egg lay in A. marginata and 86 days for A. achatina aestivated for 6 weeks. The delay may be due to necessary period required for reproductive tissue recovery and mobilization of necessary nutrient for egg formation. Omoyakhi and Okhale (2015b,c) observed diminished tissue content of the reproductive organs of snails that were aestivated for 6 weeks. They reported progressive recovery with re-appearance of the lumen and follicular enlargement within 3 and 6 weeks post aestivation when the histology of the reproductive organs was studied (Omoyakhi and Okhale, 2015d,e).

Depression in the number of clutches was observed in the aestivated groups. Omoyakhi and Osinowo (2012) reported similar trend with 6, 4, 4 and 3 clutches for 0, 2, 4 and 6 weeks aestivation lengths respectively. But 12A in this study however, recorded better result (4.25) than the 6A (2.88) by 51.79 % and by 59.36 % of the naturally controlled group (2.67). Bonnefoy-Claudet and Deray (1984) reported enhanced mating interval by hibernating Cantareus aspersus. The incubation period ranged from 25.33 days (in the 6A and 12A) to 29.33 days (recorded from the continuous watering group). The continuouswatered group (29.33) compared favourably with the records of Odaibo (1997), Akinmusi (1998), Akebejo-Samson and Akinmusi (2000) and Awah et al. (2001) who reported between 28 and 32.68 days for A. marginata. The restricted groups had significantly lower incubation period which may be connected with the source and management. The 0A has a history of continuous watering over time but the other groups were collected from the wild and were naturally controlled. Omoyakhi (2007) similarly recorded slightly lower incubation period in the aestivated group as compared with the watered group. This may therefore be added to the phenomenon of aestivation but the physiological reason for this observation was not identified in this study. Clutch size was not significantly affected by aestivation. Percentage hatchability was however improved. The values obtained generally in this study was higher than the ranges of 42.86 to 65.71 % (Omoyakhi and Osinowo, 2012), and 35 to 71.9 % (Adeniran, 2000) earlier reported. It was comparable with the ranges of 73.4 to 98 % (Akebejo-Samson and Akinmusi, 2001) and 84 to 100 % recorded by Akinmusi (2000). It is evident from this study that aestivation although depressed weight and overall number of eggs laid, it however favoured hatchability and incubation period in A. marginata.
Liveweight Changes And Reproductive Performance Of Snails (Archachatina Marginata) Under

References


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